

Optimizing Harvest of Corn Stover Fractions Based on Overall Sugar Yields following AFEX Pretreatment and Enzymatic Hydrolysis

Rutgers University has made this article freely available. Please share how this access benefits you.
Your story matters. [\[https://rucore.libraries.rutgers.edu/rutgers-lib/50910/story/\]](https://rucore.libraries.rutgers.edu/rutgers-lib/50910/story/)

This work is an **ACCEPTED MANUSCRIPT (AM)**

This is the author's manuscript for a work that has been accepted for publication. Changes resulting from the publishing process, such as copyediting, final layout, and pagination, may not be reflected in this document. The publisher takes permanent responsibility for the work. Content and layout follow publisher's submission requirements.

Citation for this version and the definitive version are shown below.

Citation to Publisher Garlock, Rebecca J., Chundawat, Shishir P. S., Balan, Venkatesh & Dale, Bruce E. (2009).
Version: Optimizing Harvest of Corn Stover Fractions Based on Overall Sugar Yields following AFEX Pretreatment and Enzymatic Hydrolysis. *Biotechnology for Biofuels* 2(1), Article number 29. <http://dx.doi.org/10.1186/1754-6834-2-29>.

Citation to this Version: Garlock, Rebecca J., Chundawat, Shishir P. S., Balan, Venkatesh & Dale, Bruce E. (2009). Optimizing Harvest of Corn Stover Fractions Based on Overall Sugar Yields following AFEX Pretreatment and Enzymatic Hydrolysis. *Biotechnology for Biofuels* 2(1), Article number 29. Retrieved from [doi:10.7282/T3NC63GK](https://doi.org/10.7282/T3NC63GK).

Terms of Use: Copyright for scholarly resources published in RUcore is retained by the copyright holder. By virtue of its appearance in this open access medium, you are free to use this resource, with proper attribution, in educational and other non-commercial settings. Other uses, such as reproduction or republication, may require the permission of the copyright holder.

Article begins on next page

1
2
3 **Optimizing Harvest of Corn Stover Fractions Based on**
4
5
6 **Overall Sugar Yields following AFEX Pretreatment and**
7
8
9 **Enzymatic Hydrolysis**
10

11
12
13
14
15 Rebecca J. Garlock*, Shishir P.S. Chundawat, Venkatesh Balan, Bruce
16
17
18 E. Dale
19

20
21
22
23 Biomass Conversion Research Laboratory, Department of Chemical Engineering and Materials
24
25 Science, Michigan State University, 2527 Engineering Building, East Lansing, MI 48824-1226, USA
26

27
28
29 **Abstract**
30

31
32 Corn stover composition changes considerably throughout the growing season
33
34 and also varies between the various fractions of the plant. These differences can
35
36 impact optimal pretreatment conditions, enzymatic digestibility, and maximum
37
38 achievable sugar yields in the process of converting lignocellulosics to ethanol.
39
40 The goal of this project was to determine which combination of corn stover
41
42 fractions provides the most benefit to the biorefinery in terms of sugar yields, and
43
44 to determine the preferential order in which fractions should be harvested.
45
46 Ammonia Fiber Expansion (AFEX) pretreatment followed by enzymatic hydrolysis
47
48 was performed on early and late harvest corn stover fractions (stem, leaf, husk and
49
50 cob). Sugar yields were used to optimize scenarios for the selective harvest of
51
52
53
54
55

56 *Corresponding author. Tel.: +1-517-432-0157 Fax.: +1-517-337-7904
57 *Email address:* garlock1@msu.edu (R.J. Garlock)
58

1
2
3 corn stover assuming 70% or 30% collection of the total available stover. The
4
5 optimal AFEX conditions for all stover fractions, regardless of harvest period, were:
6
7 1.5 (g NH₃ g⁻¹ biomass), 60% moisture content (dwb), 90°C, and 5 min residence
8
9 time. Enzymatic hydrolysis was conducted using cellulase, β-glucosidase, and
10
11 xylanase at 31.3, 41.3, and 3.1 mg g⁻¹ glucan, respectively. The optimal harvest
12
13 order for selectively harvested corn stover (SHCS) was husk > leaf > stem > cob.
14
15 This harvest scenario, combined with optimal AFEX pretreatment conditions, gave
16
17 a theoretical ethanol yield of 1.62 t⁻¹ ha and 0.72 t⁻¹ ha for 70% and 30% corn
18
19 stover collection, respectively. In the end, resources may be more effectively spent
20
21 on improving sustainable harvesting and optimizing biomass processing rather
22
23 than focusing on the selective harvest of specific corn stover fractions.
24
25
26
27
28
29
30

31 **Keywords:** AFEX; *Zea mays*; Corn stover; Enzymatic hydrolysis; Fractionation;
32
33 Harvest scenario; Agricultural residue
34
35
36
37
38
39

40 **1. Introduction**

41
42 Corn stover, the aboveground, vegetative portion of maize (*Zea mays* L.),
43
44 makes up roughly 80% of all agricultural residues produced in the U.S. [1]. Data
45
46 on annual corn stover production in the U.S. are not readily available, so various
47
48 sources have independently estimated that anywhere from 200 to 250 million dry
49
50 tons of corn stover are produced per year [1-4]. Sustainably harvested corn stover
51
52 could be used as a feedstock for a variety of applications including lignocellulosic
53
54 ethanol production. It has been estimated that 35 GL of ethanol per year could be
55
56
57
58
59
60
61
62
63
64
65

1
2
3 produced from U.S. corn stover assuming 40% of the stover is collected [5]. It is
4
5 widely acknowledged that a percentage of the produced corn stover should be
6
7 retained on the field following harvest in order to prevent soil erosion and maintain
8
9 soil organic carbon (SOC) levels. The amount that can be sustainably harvested is
10
11 highly debated and depends heavily on cropping practices, climate, topography,
12
13 and soil type [4, 6-8]. Because of these factors, estimates on the amount of corn
14
15 stover that can be sustainably harvested vary widely, anywhere from 20-80% [1, 5-
16
17
18
19 6].
20

21
22 Lignocellulosic feedstocks, such as corn stover, derive their name from the
23
24 three primary components of the plant cell wall: cellulose, hemicellulose and lignin.
25
26 The complex polysaccharides, cellulose and hemicellulose, must be broken down
27
28 into monomeric form (primarily glucose and xylose) prior to microbial fermentation
29
30 into ethanol or other valuable products. High sugar yields require a two-step
31
32 process: generally a chemical and/or physical pretreatment step followed by
33
34 enzymatic hydrolysis of the polysaccharides. Previous work has shown that
35
36 ammonia fiber expansion (AFEX) is a promising pretreatment that can be used in
37
38 the process of converting corn stover polysaccharides into ethanol as a liquid fuel
39
40 source [9-12]. AFEX pretreatment uses concentrated ammonia-water mixtures
41
42 under moderate temperatures (60-180°C) and high pressures (200-1000 psi) to
43
44 disrupt the cellular structure of the plant material by decrystallizing the cellulose,
45
46 partially depolymerizing and solubilizing the hemicellulose, and altering the form,
47
48 location and structure of lignin [9-10].
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 The structure and composition of the plant cell wall depends on a number of
4
5 factors including: developmental stage at harvest, geographical origin, type of
6
7 tissue, and other external factors including season of harvest as well as
8
9 environmental conditions experienced during growth [13]. Corn stover, like most
10
11 grasses, experiences considerable compositional changes throughout the yearly
12
13 growth period as well as significant variation between the various fractions of the
14
15 plant (i.e. leaf vs. stem) [14-16]. Largely because of these differences in
16
17 composition, stover fractions have been shown to respond differently to
18
19 pretreatment and enzymatic hydrolysis, resulting in different sugar yields [17-19]. It
20
21 is reasonable to assume that differences in composition, due largely to differences
22
23 in morphology and cell and tissue organization, could cause different stover
24
25 fractions to have different optimal pretreatment conditions for maximizing sugar
26
27 yields. For example, wheat straw leaves, when treated with dilute NaOH, required
28
29 less severe pretreatment conditions to optimize glucan yields than stem internodes
30
31 and nodes [20], so the same might be true for corn stover pretreated with ammonia
32
33 (or AFEX). Maximum sugar yields from individual fractions would be one criterion
34
35 for determining which fractions should be left on the field following harvest.
36
37 Assuming no other constraining factors, it would be most logical to harvest the
38
39 least recalcitrant biomass and leave the remainder for erosion control and soil
40
41 organic carbon maintenance [21]. Crofcheck and Montross recommended, based
42
43 on glucose yields from fractionated corn stover, a roughly 30% corn stover harvest
44
45 scenario where the selectively harvested corn stover (SHCS) was composed of all
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 of the available cobs and 74% of the leaves and husks, leaving the most
4
5 recalcitrant stalks on the field [17].
6

7 For our experiment, AFEX followed by enzymatic hydrolysis was performed on
8
9 four different corn stover fractions (stem, leaf, husk and cob) from September
10
11 (early) and November (late) harvests. The objectives of this project were: 1) to
12
13 determine whether individual stover fractions have different optimal AFEX
14
15 conditions and whether this is different from previously optimized values for
16
17 homogeneously milled corn stover [9-10], 2) to discover which fractions give the
18
19 highest glucose and xylose yields at optimal pretreatment conditions, and 3) to
20
21 model optimal harvest scenarios, assuming 30% and 70% collection of total
22
23 available dry corn stover, based on the maximum monomeric glucose and xylose
24
25 yields from each fraction.
26
27
28
29
30
31
32
33

34 **2. Materials and methods**

35 *2.1 Harvest and milling*

36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Corn stover, from a variety intended for grain production, was manually harvested from the Michigan State University Agronomy Center in East Lansing, MI in September (early harvest) and November (late harvest) of 2006. The early and late stover harvests were separately hand-sorted into four individual fractions: stems, leaves with leaf sheaths, cobs and husks. The early husk and early cob fractions were not used due to spoilage of the material prior to use. All other fractions were air-dried, with stems split lengthwise to increase the drying rate. Fractions were then milled using a Fitzpatrick JT-6 Homoloid mill (Continental

1
2
3 Process Systems, Inc., Westmont, IL), with leaf, husk, and cob fractions passing
4
5 through a 4.763mm (3/16") mesh screen and stem fractions passing through a
6
7 3.175mm (1/8") mesh screen.
8
9

10 11 12 2.2 *Composition Analysis* 13

14 Biomass moisture content was determined using a moisture analyzer (A&D,
15 Model MF-50; San Jose, CA). The composition of each corn stover fraction (ash,
16
17 lignin, glucan and xylan content) was determined using the NREL standard
18
19 protocols [22]: Ash analysis, (LAP 005); Removal of Extractives (LAP 010); and
20
21 Structural Carbohydrates & Lignin (LAP 002, 003, 004, 007, 019). The acid
22
23 insoluble lignin analysis method was modified to use 47mm, 0.22µm pore-size,
24
25 mixed-cellulose ester filter discs (Millipore Corp. Bedford, MA) during the filtration
26
27 step instead of fritted crucibles. Due to problems with burning, these discs, with
28
29 the filtered lignin residue, could not be dried in the vacuum oven and so were dried
30
31 overnight in a desiccator prior to weighing. Soluble sugars could not be quantified
32
33 following the extraction due to difficulties in resolving distinct peaks using the
34
35 HPLC, and were not included in the composition.
36
37
38
39
40
41
42
43
44

45 2.3 *AFEX Treatment* 46

47 A small-scale benchtop reactor system, consisting of four separate 22 mL
48
49 stainless steel (#316) reaction vessels, was used for the pretreatment process.
50
51 Prior to biomass loading, the biomass was adjusted to the appropriate moisture
52
53 content with deionized water, after which 3.0 g (dwb) of biomass was added to
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 each reaction vessel. A metal screen was placed over the biomass inside of each
4
5 vessel, to prevent escape of biomass during venting. The loaded reactor units
6
7 were weighed and then attached to the reactor manifold, and any air within the
8
9 reactor vessels was then removed using a rotary vacuum pump. Liquid anhydrous
10
11 ammonia was dispensed into the manifold via Swagelok screw valves (Swagelok
12
13 Co., Solon, OH) and then added to the reactor vessels. The reactors were
14
15 weighed to determine the amount of ammonia added and then were vented slightly
16
17 to reach the appropriate ammonia loading. A heating mantle was used to raise the
18
19 reactors to the desired temperature and maintain it for the set residence time. At
20
21 the completion of the residence time, the reactor pressure was explosively
22
23 released via a stainless steel (#316) ball valve, and the reactor simultaneously
24
25 cooled. The pretreated biomass was removed from the vessel and left in the fume
26
27 hood overnight to allow the residual ammonia to evaporate.
28
29
30
31
32
33
34
35

36 *2.4 Enzymatic Hydrolysis*

37
38 NREL protocol (LAP 009) [22] was followed for the enzymatic hydrolysis of
39
40 pretreated and untreated (control) samples. All samples were hydrolyzed in 20mL
41
42 screw-cap vials at 1% glucan loading and a total volume of 15mL. Samples were
43
44 adjusted to a pH of 4.8 by 1M citrate buffer solution. Spezyme[®] CP (Genencor
45
46 Division of Danisco US, Inc.) cellulase at 15 FPU g⁻¹ glucan (31.3 mg protein g⁻¹
47
48 glucan) and β -glucosidase (Novozyme[®] 188, Novozymes Corp.) at 64 p-NPGU g⁻¹
49
50 glucan (41.3 mg protein g⁻¹ glucan) were added to each vial with a total protein
51
52 content of 72.6 mg protein g⁻¹ glucan. In addition, certain samples were also
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 hydrolyzed using xylanase (Multifect[®] Xylanase, Genencor Division of Danisco US
4
5 Inc.) at 10% of total cellulase protein (3.1 mg protein g⁻¹ glucan), giving a total
6
7 protein content of 75.7 mg protein g⁻¹ glucan. Samples were placed in a New
8
9 Brunswick Scientific (Edison, NJ) incubator shaker and hydrolyzed at 50°C and
10
11 150 rpm for 72 hr. The hydrolysates were sampled at 24 hr and 72 hr, following
12
13 which samples were heated at 90°C for 15 minutes, cooled, and centrifuged at 15K
14
15 for 5 minutes. The supernatant was filtered into HPLC shell vials using a 25mm,
16
17 0.2 µm polyethersulfone syringe filter (Whatman Inc. Florham Park, NJ) after which
18
19 samples were stored at -20°C until further sugar analysis.
20
21
22
23
24
25

26 2.5 Sugar Analysis

27
28 A high-pressure liquid-chromatography (HPLC) system was used to determine
29
30 the sample monomeric glucose and xylose concentrations following enzymatic
31
32 hydrolysis. The HPLC system consisted of a Waters (Milford, MA) pump, auto-
33
34 sampler and Waters 410 refractive index detector, equipped with a Bio-Rad
35
36 (Hercules, CA) Aminex HPX-87P carbohydrate analysis column with attached
37
38 deashing guard column. Degassed HPLC grade water was used as the mobile
39
40 phase, at 0.6 mL/min, with the column temperature set at 85°C. Injection volume
41
42 was 10 µL with a run time of 20 min per sample. Mixed sugar standards were used
43
44 to quantify the amount of monomeric glucose and xylose in each hydrolysate
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 **2.6 Statistical Analysis**
4

5 Monomeric glucose and xylose yields following enzymatic hydrolysis were
6 analyzed using multivariate analysis of variance (MANOVA) in Minitab15 Statistical
7 Software (Minitab Inc., 2006). The interactive effects plot of harvest period and
8 stover fraction between each other, the four AFEX pretreatment parameters
9 (moisture content, ammonia loading, temperature and residence time), and
10 xylanase addition, was also constructed using Minitab.
11
12
13
14
15
16
17
18
19
20
21

22 **3. Results**
23

24 **3.1 Composition Analysis**
25

26 The composition of each of the corn stover fractions from each harvest are
27 listed in Table 1. The value of the “other” column was determined by difference
28 from 100% and the standard deviation is representative of three replicates.
29

30 Statistically, the early and late stem and the late leaves and husk had the highest
31 glucan content, while the early leaves and late cob had the lowest glucan content.
32

33 The xylan content of the late fractions was significantly higher than their early
34 counterparts and tended to decrease from late cob > late husk > late stem > late
35 leaves > early stem > early leaves. The acid-insoluble lignin content was similar
36 for all fractions, except for the cob, which had the highest lignin content, and the
37 late husk which had statistically less lignin than the late stem. The ash content of
38 all fractions were statistically different and decreased from early leaves > late
39 leaves > early stem > late stem > late husk > late cob.
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

3.2 AFEX Pretreatment and Hydrolysis

Pretreatment conditions for AFEX-treated corn stover have been previously optimized at 1.0 (g ammonia g⁻¹ dry biomass), 60% moisture content (dwb), 90°C and 5 min. residence time [9-10]. These conditions were treated as the “base case” for analysis of pretreatment conditions. The effect of pretreatment conditions on monomeric glucose and xylose yields, following hydrolysis, was tested by varying one process parameter (temperature, ammonia loading, moisture content or residence time) at a time (e.g. raising the temperature from 90°C to 100°C). Once the preliminary data had been gathered, the untreated control, base case, and best case were supplemented with xylanase during hydrolysis to observe the effect on sugar yields.

Figure 1 shows the monomeric glucose and xylose yields for a variety of conditions with particular comparisons between untreated and AFEX treated materials at a range of ammonia loadings. The effect of xylanase addition to the enzyme cocktail can also be observed in this figure. Error bars in all figures represent the mean \pm 1 standard deviation. From the figure, it can be seen that AFEX substantially improves both glucose and xylose monomeric sugar yields for all harvest periods and corn stover fractions when compared to untreated materials. The xylose yields for the late fractions are especially improved by AFEX pretreatment, even at low ammonia loadings such as 0.5 (g ammonia g⁻¹ biomass).

The increase in ammonia loading from 0.5 to 1.5 (g ammonia g⁻¹ biomass) had different effects on early harvest and late harvest corn stover fractions. For the early harvest stover, without xylanase addition, glucose yields peak at 1.0 (g

1
2
3 ammonia g⁻¹ biomass). This optimum is similar to what has been seen previously
4
5 with AFEX-treated corn stover [9-10], which may indicate that that material was
6
7 from an earlier harvest. The xylose yields are relatively unaffected by any further
8
9 increase above 1.0 (g ammonia g⁻¹ biomass). However, when performing the
10
11 same experiment with the late harvest corn stover there is an increase in both
12
13 glucose and xylose yields for all fractions when increasing from 1.0 to 1.5 (g
14
15 ammonia g⁻¹ biomass).
16
17

18
19 Xylanase addition had little to no effect on increasing either glucose or xylose
20
21 sugar yields in untreated corn stover fractions. For AFEX-treated early harvest
22
23 fractions, the addition of xylanase at 1.0 (g ammonia g⁻¹ biomass) had no effect on
24
25 monomeric xylose yields, and it slightly lowered glucose yields. At 1.5 (g ammonia
26
27 g⁻¹ biomass), all fractions and harvests experienced an increase in both the
28
29 monomeric xylose and glucose yields with the addition of xylanase.
30
31

32
33 The leaf and stem, for both early and late harvests, have similar glucose yields
34
35 for 1.5 (g ammonia g⁻¹ biomass) ammonia loading, however the leaf glucan is more
36
37 digestible as seen by the greater yield (percent of maximum theoretical glucan
38
39 available). The late harvest husk approaches theoretical glucose yields at the
40
41 optimal condition of 1.5 (g ammonia g⁻¹ biomass). Because of this, the addition of
42
43 xylanase for this pretreatment condition increases husk xylose yields slightly, but
44
45 not glucose yields as is seen for the other fractions. With the addition of xylanase
46
47 at 1.5 (g ammonia g⁻¹ biomass), the cob and leaf also approach near theoretical
48
49 glucose yields.
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 Figure 2 shows the effect of pretreatment temperature on glucose and xylose
4 yields from corn stover fractions. Altering the temperature by 10°C from the base
5 case had little effect on glucose and xylose yields. There is a definite peak in
6 glucose yields at 90°C for the early harvest but the late harvest has no apparent
7 difference in yields for 80, 90 or 100°C. In previous work [10], raising the
8 temperature above 90°C had a negative impact on ethanol yields from
9 simultaneous saccharification and fermentation (SSF).
10
11
12
13
14
15
16
17
18

19 Decreasing the moisture content to 40% (dwb) and eliminating the residence
20 time (the time for which the reactor was held at the set temperature following heat-
21 up) each had a negative impact on glucose and xylose yields for all fractions
22 (Figure 3). For all stover fractions, except the late husk, it was more detrimental in
23 terms of sugar yields to decrease the residence time rather than the moisture
24 content.
25
26
27
28
29
30
31
32
33
34
35

36 3.3 *Statistical Analysis*

37
38 Multivariate analysis of variance (MANOVA) was conducted to determine the
39 significance of harvest date, corn stover fraction, AFEX parameters and xylanase
40 addition on both the 24 hour and 72 hour monomeric glucose and xylose yields.
41 Interactive effects were also examined between harvest date and stover fraction
42 and each of the other parameters. Because conclusions regarding significance
43 were the same for 24 hour and 72 hour yields for both glucose and xylose (Table
44 2), only the 72 hour yields were used for the interactive effects plot (Figure 4).
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 Glucose yields were significantly affected by three of the AFEX pretreatment
4
5 conditions: ammonia loading, moisture content, and residence time, but not by
6
7 temperature. Glucose yields were also dependent on corn stover fraction and
8
9 whether xylanase was added to the hydrolysis cocktail. Of the interactive effects
10
11 analyzed, only harvest date X ammonia loading had any significant effect on
12
13 monomeric glucose yields.
14
15

16
17 Xylose yields were significantly affected by all four AFEX pretreatment
18
19 conditions, including temperature. Unlike the case for glucose yields, xylose yields
20
21 were not significantly affected by corn stover fraction, but they were affected by
22
23 both the harvest date and the addition of xylanase to the hydrolysis cocktail. There
24
25 were also interactive effects on xylose yields from harvest date X ammonia loading
26
27 and corn stover fraction X xylanase addition.
28
29
30

31 When analyzing the interactive effects plot, significant interactive effects will
32
33 have very different slopes for the different lines in that portion of the graph. For
34
35 example, when observing the interactive effect of harvest X ammonia on xylose
36
37 yields, the slope of the early and late harvest lines are roughly the same when
38
39 increasing the ammonia loading from 0.5 to 1.0 (g ammonia g⁻¹ biomass).
40
41 However, when increasing the ammonia loading from 1.0 to 1.5 (g ammonia g⁻¹
42
43 biomass), the slope of the late harvest line is significantly steeper than the slope of
44
45 the early harvest line. This difference in slope signifies that most of the impact of
46
47 ammonia loading on this interaction is due to the second increase, not the first.
48
49 This implies that the higher ammonia loading has a greater effect on the later
50
51 harvest than the early harvest.
52
53
54
55
56
57
58
59
60
61
62
63
64
65

3.4 Optimization of Harvest Scenarios

Crofcheck and Montross [17] found that the weighted sum of the glucose yields from individual pretreated fractions was not statistically different from the glucose yield from whole pretreated corn stover. This means that glucose yields for selectively harvested corn stover (SHCS) could be predicted using glucose yields from individual fractions. Our estimate of the late harvest dry matter distribution of corn stover (Figure 5), was based on published data from four sources [15-17, 23], and is similar to standard estimates of corn stover dry matter distribution near corn harvest [24]. Corn stover dry matter yields, particularly of the husk and leaf, tend to decrease rapidly due to weathering over the course of the grain harvest season [15-16, 21, 23, 25-27]. This estimate attempts to account for both the effects of the late harvest date as well as our inclusion of the leaf sheath with the leaf fraction instead of the stem fraction, as is often the case [15-16].

Because of the wide range of opinions on how much corn stover can be sustainably harvested and because the amount will likely change for a given field depending on environmental conditions and agricultural practices [4, 6-8], we have modeled a number of corn stover harvest scenarios for both a liberal harvest estimate (70% dwb) and a conservative harvest estimate (30% dwb). The goal was to determine which combination of fractions provides the most benefit to the biorefinery in terms of sugar yields, and to determine the preferential order in which fractions should be harvested from the field. For this analysis, the sugar yields used were from 72 hour hydrolysis of AFEX-treated late corn stover. The option of an

1
2
3 early harvest was not analyzed because of the lack of data for husk and cob
4
5 fractions. Scenarios were analyzed with regard to the effect of increasing ammonia
6
7 loading from 1.0 to 1.5 (g ammonia g⁻¹ biomass) and for the maximized sugar yield,
8
9 either glucose or xylose. This gave four potential scenarios (1.0 + glucose, 1.5 +
10
11 glucose, 1.0 + xylose and 1.5 + xylose). All other AFEX and hydrolysis conditions
12
13 were held constant (60% dwb moisture, 90°C, 5 min residence time + 10%
14
15 xylanase addition). Because glucose yields were consistently higher than xylose
16
17 yields, the harvest conditions to obtain maximum glucose yields for all of the
18
19 scenarios also corresponded with the maximum total sugar yields. The conditions
20
21 selected resulted in three scenarios for selectively harvesting corn stover (Table 3),
22
23 because the harvest scenario to maximize glucose yields was the same for both
24
25 ammonia loadings. The relative amounts of harvested fractions for each scenario
26
27 are represented in Figure 5 for both the 70% and 30% harvests.
28
29
30
31
32

33
34 The estimated whole corn stover dry matter distribution was used to predict
35
36 monomeric glucose and xylose yields from the three different harvest scenarios for
37
38 both 70% (Table 4) and 30% (Table 5) harvest of on-field corn stover using
39
40 weighted averaging of individual fraction sugar yields. It is important to note that
41
42 values given in these tables do not attempt to take into account the ability or
43
44 inability to harvest the specific fractions or any losses due to inefficiencies in
45
46 harvest, transport, and storage of corn stover, which can be significant depending
47
48 on the methods used. A recent study found that the maximum amount of corn
49
50 stover was available at grain physiological maturity (15.6 t ha⁻¹) and steadily
51
52 decreased over the harvest period to a minimum of 8.6 t ha⁻¹ [27]. Because this
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 value takes into account the late season of harvest and because it is in the range
4
5 of most literature estimates of corn stover yields (7.8 – 8.8 t ha⁻¹) [1-2, 28], 8.6 t ha⁻¹
6
7 of available corn stover was chosen to estimate the total sugars that could be
8
9 produced per hectare for the given harvest scenario. The standard value of 0.51
10
11 (theoretical g ethanol produced g⁻¹ sugar consumed) was used to determine the
12
13 theoretical ethanol production from a hectare of SHCS and does not take into
14
15 account inefficiencies in fermentation.
16
17
18
19
20
21
22

23 **4. Discussion**

24 *4.1 Composition Analysis*

25
26
27 Fractions from the late harvest tended to have a slightly higher percentage of
28
29 cell wall components (although not always significant) and slightly lower
30
31 percentage of ash compared to their early harvest counterparts. For corn stover,
32
33 the increase in lignin and cellulose, and the decrease in ash have been observed
34
35 elsewhere [23, 25] as well as a general increase in all cell wall components with a
36
37 decrease in non-structural carbohydrates [26], and an increase in lignin and xylan
38
39 with a drop in soluble solids [15] with increasing maturity. This observed increase
40
41 in cellulose (glucan), hemicellulose (glucan and xylan) and lignin content is due to
42
43 the secondary thickening of the plant cell wall that continues to occur for as long as
44
45 the plant matures. During this same time there is also a decrease in ash content
46
47 [14]. However, while there is continual change in the dry matter composition until
48
49 late in the season, this tends to be very small during the grain harvest period [2,
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

4.2 AFEX Pretreatment

Based on the final total sugar yields, the optimal AFEX pretreatment conditions were observed to be consistent for all fractions for both early and late harvest corn stover: 1.5:1 (g NH₃ g⁻¹ biomass), 60% moisture content (dwb), 90°C, 5 min residence time; with 10% xylanase addition (mg xylanase protein mg⁻¹ cellulase protein) in addition to the standard enzyme mixture used during enzymatic hydrolysis.

For AFEX-treated early harvest fractions, the addition of xylanase at 1.0 (g ammonia g⁻¹ biomass) had no effect on monomeric xylose yields, and it slightly lowered glucose yields. This drop in glucose yields could be due to competition for binding sites on the cellulose chains between enzymes in the xylanase and cellulase mixtures. The fact that there is no increase in xylose yields with the addition of xylanase supports this conclusion.

The higher optimal ammonia loading for the late harvest fractions compared with the early harvest could be due to a number of reasons. AFEX, by ammoniation of the active methoxyl sites of lignin [29], may be preventing lignin from binding to the hydrolysis enzymes. It is likely that this mechanism is operating, and it may be one of the main reasons for the increase seen from 0.5 to 1.0 (g ammonia g⁻¹ biomass). However, if this were the reason for the difference in optimum ammonia loading between the early and late harvests, then the lignin content of the later harvest should be greater. However, this is not the case; statistically the lignin contents of the early and late fractions are identical. The difference in optimal ammonia loading is more likely due to the increase in xylan

1
2
3 content and possibly increased cross-linking between hemicellulose and lignin from
4
5 the early to late harvest. Ferulate cross-linking occurs between lignin and
6
7 arabinoxylan in the plant cell wall, with the ferulates ether-linked to lignin and ester-
8
9 linked to the arabinoxylan [30]. Ammonolysis of the ferulate ester linkages to
10
11 arabinoxylan side-chains is believed to be one major reactions occurring during the
12
13 AFEX process [31]. These mechanisms may be opening up the cell wall
14
15 ultrastructure more effectively at the higher ammonia loading, allowing the
16
17 enzymes greater access to cellulose, hence the optimum at 1.5 (g ammonia g⁻¹
18
19 biomass). Also, by allowing greater access to a larger amount of xylan, the
20
21 xylanase enzymes may be competing less with the cellulases for binding sites,
22
23 hence the increase in glucose yields with xylanase addition for the early harvests
24
25 at 1.5 (g ammonia g⁻¹ biomass).
26
27
28
29
30

31 It is worth noting that the husk, the material with the lowest lignin content, while
32
33 having the second-highest xylan content, is least affected by the combination of
34
35 increased ammonia loading and xylanase addition. At 1.5 (g ammonia g⁻¹
36
37 biomass), the xylose yield only increases by 6.1% with the addition of xylanase to
38
39 the hydrolysis cocktail. The late cob, which has a significantly higher lignin and
40
41 xylan content than all of the other materials, experiences the largest impact on
42
43 xylose yields due to the combination of increased ammonia loading and addition of
44
45 xylanase, a 22.5% increase at 1.5 (g ammonia g⁻¹ biomass) with the addition of
46
47 xylanase. The higher ammonia loading would cleave more linkages between the
48
49 hemicellulose and lignin, solubilizing more oligomeric and monomeric xylose and
50
51 perhaps some lignin as well. These exposed, solubilized sugars would be much
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 easier to hydrolyze with the xylanase. It might be possible, given the very high
4
5 xylan content of the cob, that more xylanase would be needed to achieve near
6
7 complete monomeric xylose yields. Because the xylanase loading was based on a
8
9 percentage of the cellulase loading (and therefore the glucan content), and
10
11 because the g glucan g⁻¹ xylan ratio for the cob (0.85 g glucan g⁻¹ xylan) is much
12
13 lower than the other fractions (1.47 – 1.85 g glucan g⁻¹ xylan), the xylanase loading
14
15 in terms of the xylan content (mg xylanase g⁻¹ xylan) is much lower for the cob
16
17 fraction (Table 6). This may be one reason for the much lower xylose yield relative
18
19 to the maximum theoretical xylose yield of the late cob fraction.
20
21
22
23
24
25

26 4.3 *Statistical Analysis*

27
28 All AFEX conditions had significant impacts on sugar yields, except for
29
30 temperature which had no effect on glucose yield. Based on least squares means
31
32 analysis (data not shown), the temperature effect on xylose yield is likely due to a
33
34 greater yield increase as the temperature is raised from 80 to 90°C rather than the
35
36 decrease in yield when temperature is raised from 90 to 100°C. For the range of
37
38 conditions tested, optimizing the ammonia loading, moisture content and residence
39
40 time are more important for maximizing sugar yields from corn stover. However,
41
42 this conclusion may change for a different range of temperatures and should not be
43
44 extrapolated to other conditions.
45
46
47
48
49

50 Harvest date had a significant impact on xylose yields but not on glucose yields.
51
52 This is largely due to the fact that the xylan content of the late fractions was greater
53
54 than the xylan content of the early fractions, whereas the glucan content was not
55
56
57
58
59

1
2
3 significantly different between harvests. The corn stover fractions tested had a
4
5 significant effect on glucose yields but not on xylose yields. The late stem, husk,
6
7 leaf and early stem fractions had no significant statistical difference in their glucan
8
9 contents, so their relative recalcitrance, in terms of glucose yields, can be inferred
10
11 from Figure 4. Because the husk has the highest glucose yield, it can be
12
13 considered the least recalcitrant followed by the leaf and then the stem. Inferences
14
15 cannot be made regarding the cob because its glucan content is statistically lower
16
17 than the other three fractions. However, because the cob approaches theoretical
18
19 glucan yields at optimal conditions while the stem does not (Figure 1), it may be
20
21 less recalcitrant in terms of the conversion of glucan.
22
23
24

25
26 For interactive effects, only two were significant: harvest date X ammonia
27
28 loading and corn stover fraction X xylanase addition. The harvest X ammonia
29
30 interaction was significant for both glucose and xylose yields. The increase in
31
32 ammonia loading from 1.0 to 1.5 (g ammonia g⁻¹ biomass) appears to have a
33
34 greater effect on the late harvest than the early harvest, but this may be due to the
35
36 lack of data for the early harvest cob. Because the cob is the fraction most
37
38 affected by the increase in ammonia loading, particularly for xylose yield, the lack
39
40 of early cob data may lead to an apparent difference in effects between harvests,
41
42 that is not actually present.
43
44
45
46

47
48 The second significant interactive effect was for corn stover fraction X xylanase
49
50 addition, but only for xylose yields. The main reason for this effect, as can be
51
52 observed from Figure 4, is due to the cob fraction which was much more strongly
53
54 affected by the addition of xylanase than all of the other fractions, whose
55
56
57
58
59
60
61
62
63
64
65

1
2
3 responses were fairly similar. This conclusion is supported by the fact that when
4
5 the data for the late cob was removed from the analysis, the fraction X xylanase
6
7 interaction became non-significant (data not shown). Taken together, these results
8
9 indicate that of all the corn stover components, the cob reacts the most differently
10
11 during enzymatic hydrolysis. As mentioned previously, the cob may require a
12
13 much higher xylanase loading than the other fractions in order to release xylose
14
15 remaining in the biomass or to convert the AFEX solubilized xylo-oligomers.
16
17
18
19
20

21 *4.4 Empirical Modeling of Harvest Scenarios*

22
23
24 Harvest scenario A, which selectively harvests the husk followed by the leaf,
25
26 stem and lastly cob, obtained the highest sugar and ethanol yields of all the
27
28 scenarios, and as a result was chosen as the optimal harvest scenario for AFEX-
29
30 treated corn stover. Harvest scenario A was also preferable to scenarios B and C
31
32 for a number of other reasons. First, optimizing collection for maximum glucose
33
34 yields is preferable because most current, relevant microbial strains selectively
35
36 utilize hexoses over pentoses as a carbon source during ethanolic fermentation
37
38 [11, 32]. Second, harvest scenario A selectively leaves behind the more lignified
39
40 fractions on the field which may prove more valuable for improving SOC levels due
41
42 to the longer half-life of lignin compared to cellulose and hemicellulose [4, 33].
43
44 Lastly, selectively leaving the least desirable cob and/or lower portion of the stem
45
46 behind may be feasible with current stover harvesting systems, but the downside
47
48 is, that depending on the method, complete collection of the leaves may not be
49
50 possible [34-35].
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 Crofcheck and Montross [17] recommended, based on glucose yields from
4
5 fractionated corn stover, a roughly 30% corn stover harvest scenario where the
6
7 selectively harvested corn stover (SHCS) was composed of all of the available
8
9 cobs and 74% of the leaves and husks, leaving the most recalcitrant stalks on the
10
11 field. The difference between their optimal harvest scenario and ours is most likely
12
13 due to their experimental methods for pretreatment and the subsequent analysis.
14
15 Pretreatment of lignocellulosic biomass using dilute sodium hydroxide, solubilizes
16
17 much of the lignin and some of the hemicellulose into the liquid pretreatment
18
19 stream [36, 37], so it is unlikely that glucan content of the pretreated corn stover
20
21 corresponds to glucan content of the untreated corn stover. For similar
22
23 pretreatment conditions of corn stover, Varga et al. found a 41.9% mass loss from
24
25 the untreated dry corn stover to the pretreated solids, and the composition of the
26
27 pretreated material shifted in favor of a higher glucan content [36]. The cob has a
28
29 significantly higher xylan and lignin content than the other fractions of the corn
30
31 plant, and so it is reasonable to assume that it will lose a greater proportion of its
32
33 mass following dilute alkali pretreatment. Because this mass loss wasn't taken into
34
35 account [17], the amount of glucan that could be obtained on a mass basis from
36
37 the untreated fractions was over-exaggerated, particularly from the xylan- and
38
39 lignin-rich cob. If the mass loss had been taken into account, it is likely that their
40
41 choice of optimal fractions for harvest would have been different. Because AFEX
42
43 is a dry-to-dry process with little mass loss during pretreatment, the glucan content
44
45 of the untreated material can be used as the glucan content of the pretreated
46
47 material [9].
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 A comparison of Table 4 and Table 5 reveals that the amount of corn stover
4
5 harvested has the largest impact on total sugars ha^{-1} and theoretical ethanol yield
6
7 ha^{-1} . Going from a 70% collection scenario to a 30% collection scenario cut the
8
9 ethanol yield by ~55% for all scenarios. Dropping the ammonia loading from 1.5 to
10
11 1.0 (g ammonia g^{-1} biomass) caused a ~15-20% decrease, while switching desired
12
13 sugars from glucose to xylose (*i.e.* changing harvest scenarios but keeping stover
14
15 collection and AFEX and enzymatic hydrolysis conditions constant) caused a ~2-
16
17 4% decrease. To determine the sensitivity of changing the harvest scenario, the
18
19 model was run assuming the “worst case scenario”, where the biomass was
20
21 harvested in the manner that would give the worst possible sugar yields (data not
22
23 shown). The worst case scenario led to a decrease in the total sugar yields and
24
25 theoretical ethanol yields per hectare ranging from ~4 – 18.5%. The drop was
26
27 more pronounced for the 30% stover harvest because the SHCS dry matter
28
29 distribution will be completely different for the best and worst case harvests, while
30
31 the 70% harvest will have some similarity in dry matter distribution between the
32
33 best and worst cases. As expected, the theoretical ethanol yield was substantially
34
35 lower (~70-85%), when comparing untreated corn stover to the AFEX-treated
36
37 cases (data not shown).
38
39
40
41
42
43
44

45 Shinnars et al. [35] analyzed the effect of cut height of corn stover on predicted
46
47 ethanol yields and found that the amount of ethanol produced was only ~3%
48
49 greater (L Mg^{-1} DM) for the low cut vs. the high cut. Because the cut height is
50
51 basically a harvest scenario that leaves a portion of the lower stem and leaves
52
53 behind, when basing the analysis on dry matter harvested, these results are
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 comparable to our results for the effect of harvested fraction type on ethanol yields.
4
5 Similarly to our results, they found that the proportion of fractions harvested had
6
7 little impact on predicted ethanol yields. However, when analyzed based on the
8
9 ethanol yield per hectare, the increase in total dry matter harvested with the lower
10
11 cut height increased the predicted ethanol yield by 52% compared to the higher cut
12
13 [35], which was similar to our results for increased dry matter collection.
14
15

16
17 Based on these results, optimizing the fractions collected during harvest has a
18
19 much smaller impact on potential yields than optimizing pretreatment and
20
21 hydrolysis conditions. The amount of stover harvested has the greatest impact on
22
23 theoretical ethanol production per hectare. It will be very important, in terms of
24
25 maximizing ethanol production, to develop methods to efficiently maximize harvest
26
27 of corn stover, while still maintaining soil productivity and preventing erosion.
28
29
30
31

32 33 34 **5. Conclusions** 35 36

37
38 Based on monomeric glucose and xylose yields, the optimal AFEX conditions,
39
40 for all stover fractions (leaf, stem, husk and cob) regardless of harvest period, were
41
42 found to be 1.5 (g NH₃ g⁻¹ biomass), 60% moisture content (dwb), 90°C, and 5 min
43
44 residence time; with enzyme loading during hydrolysis of 31.3 mg of cellulase
45
46 (Spezyme[®] CP), 41.3 mg of β-glucosidase (Novozyme[®] 188) and 3.1 mg xylanase,
47
48 g⁻¹ glucan. These conditions are different from those presented in previous
49
50 analyses [9, 10], largely due to the inclusion of xylanase in the hydrolysis cocktail.
51
52 The addition of xylanase was necessary to achieve high xylose yields at moderate
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 cellulase loadings and moderate AFEX conditions, particularly with respect to the
4
5 more recalcitrant cob and stem fractions.
6

7
8 The optimal harvest scenario for the collection of selectively harvested corn
9
10 stover (SHCS) would harvest the husk followed by the leaves, then the stem, and
11
12 lastly the cob. This harvest scenario was independent of ammonia loading during
13
14 AFEX pretreatment and maximized glucose and ethanol yield from SHCS. This
15
16 scenario, combined with the optimal AFEX pretreatment conditions for SHCS gave
17
18 a theoretical ethanol yield of $1.62 \text{ t}^{-1} \text{ ha}$ for the 70% dry matter harvest and 0.72 t^{-1}
19
20 ha for the 30% dry matter harvest. Decreasing the stover collection from 70% to
21
22 30% dropped the ethanol yield per hectare by ~55%. Maximizing stover collection
23
24 while protecting soil health will be the most important factor for maximizing ethanol
25
26 yields from corn stover.
27
28
29
30

31
32 Optimizing collection of corn stover fractions has little impact on the theoretical
33
34 ethanol yield per hectare (~2-4%), especially compared to the moderate effect of
35
36 optimizing pretreatment and hydrolysis conditions (~15-20%). The dry matter
37
38 distribution of collected corn stover fractions is generally much less important than
39
40 the optimization of the ethanol production process, however it is still something that
41
42 needs to be taken into account because harvesting the worst fractions can also
43
44 decrease ethanol yields, particularly for a more conservative harvest.
45
46
47
48
49
50

51 **Acknowledgements**

52 The authors gratefully acknowledge Michigan State University Research
53
54 Foundation for funding this project; Bill Widdicombe and the MSU Agronomy Farm
55
56
57
58

1
2
3 for providing the corn stover; Genencor Division of Danisco US Inc., for providing
4 commercial enzymes (Spezyme[®] CP Cellulase and Multifect[®] Xylanase); and
5
6
7 Derek Marshall for constructing the AFEX reactor system that was used for these
8
9
10 experiments.

11 12 13 14 15 16 **References**

- 17
18
19 [1] Kadam KL, McMillan JD. Availability of corn stover as a sustainable
20 feedstock for bioethanol production. *Bioresource Technology* 2003; 88:17-
21 25.
- 22
23 [2] Sokhansanj S, Turhollow A, Cushman J, Cundiff J. Engineering aspects of
24 collecting corn stover for bioenergy. *Biomass & Bioenergy* 2002; 23:347-55.
- 25
26 [3] Innovative Methods for Corn Stover Collecting, Handling, Storage and
27 Transport <<http://www.nrel.gov/docs/fy04osti/33893.pdf>> [Accessed 1 Jul
28 2008]
- 29
30 [4] Wilhelm WW, Johnson JMF, Hatfield JL, Voorhees WB, Linden DR. Crop and
31 soil productivity response to corn residue removal: A literature review.
32 *Agronomy Journal* 2004; 96:1-17.
- 33
34 [5] Kim S, Dale BE. Global potential bioethanol production from wasted crops
35 and crop residues. *Biomass & Bioenergy* 2004; 26:361-75.
- 36
37 [6] Kim S, Dale BE. Life cycle assessment of various cropping systems utilized
38 for producing biofuels: bioethanol and biodiesel. *Biomass & Bioenergy* 2005;
39 29:426-39.
- 40
41 [7] Wilhelm WW, Johnson JMF, Karlen DL, Lightle DT. Corn stover to sustain
42 soil organic carbon further constrains biomass supply. *Agronomy Journal*
43 2007; 99:1665-7.
- 44
45 [8] Mann L, Tolbert V, Cushman J. Potential environmental effects of corn (*Zea*
46 *mays* L.) stover removal with emphasis on soil organic matter and erosion.
47 *Agriculture, Ecosystems and Environment* 2002; 89:149-66.
- 48
49 [9] Teymouri F, Laureano-Perez L, Alizadeh H, Dale BE. Optimization of the
50 ammonia fiber explosion (AFEX) treatment parameters for enzymatic
51 hydrolysis of corn stover. *Bioresource Technology* 2005; 96:2014-2018.
- 52
53 [10] Teymouri F, Laureano-Pérez L, Alizadeh H, Dale BE. Ammonia fiber
54 explosion treatment of corn stover. *Applied Biochemistry and Biotechnology*
55 2004; 113-116:951-63.
- 56
57
58
59
60
61
62
63
64
65

- 1
2
3 [11] Wyman CE, Dale BE, Elander RT, Holtzaple M, Ladisch MR, Lee YY.
4 Comparative sugar recovery data from laboratory scale application of leading
5 pretreatment technologies to corn stover. *Bioresource Technology* 2005;
6 96:2026-2032.
7
8 [12] Lau MW, Dale BE, Balan V. Ethanol fermentation of ammonia fiber
9 expansion (AFEX) treated corn stover and distillers grain without
10 detoxification and external nutrient supplementation. *Biotechnology and*
11 *Bioengineering* 2008; 99:529-39.
12
13 [13] Nevins DJ. Analysis of forage cell wall polysaccharides. In: Jung HG, Buxton
14 DR, Hatfield RD, Ralph J, editors. *Forage cell wall structure and digestibility*.
15 Madison, WI: ASA-CSSA-SSSA; 1993, p. 105-29.
16
17 [14] Aman P. Composition and structure of cell wall polysaccharides. In: Jung
18 HG, Buxton DR, Hatfield RD, Ralph J, editors. *Forage cell wall structure and*
19 *digestibility*. Madison, WI: ASA-CSSA-SSSA; 1993, p. 183-99.
20
21 [15] Pordesimo LO, Hames BR, Sokhansanj S, Edens WC. Variation in corn
22 stover composition and energy content with crop maturity. *Biomass &*
23 *Bioenergy* 2005; 28:366-74.
24
25 [16] Shinnors KJ, Binversie BN. Fractional yield and moisture of corn stover
26 biomass produced in the Northern US Corn Belt. *Biomass & Bioenergy* 2007;
27 31:576-84.
28
29 [17] Crofcheck CL, Montross MD. Effect of stover fraction on glucose production
30 using enzymatic hydrolysis. *Transactions of the ASAE* 2004; 47:841-4.
31
32 [18] Montross MD, Crofcheck CL. Effect of stover fraction and storage method on
33 glucose production during enzymatic hydrolysis. *Bioresource Technology*
34 2004; 92:269-74.
35
36 [19] Hoskinson RL, Karlen DL, Birrell SJ, Radtke CW, Wilhelm WW. Engineering,
37 nutrient removal, and feedstock conversion evaluations of four corn stover
38 harvest scenarios. *Biomass & Bioenergy* 2007; 31:126-36.
39
40 [20] Duguid KB, Montross MD, Radtke CW, Crofcheck CL, Shearer SA, Hoskinson
41 RL. Screening for sugar and ethanol processing characteristics from
42 anatomical fractions of wheat stover. *Biomass & Bioenergy* 2007; 31:585-92.
43
44 [21] Akin DE, Morrison WH, Rigsby LL, Barton FE, Himmelsbach DS, Hicks KB.
45 Corn stover fractions and bioenergy. *Applied Biochemistry and Biotechnology*
46 2006; 129-132:104-16.
47
48 [22] NREL, Chemical Analysis and Testing (CAT) Standard Procedures; National
49 Renewable Energy Laboratory: Golden, CO, 2004.
50
51 [23] Tolera A, Sundstøl F. Morphological fractions of maize stover harvested at
52 different stages of grain maturity and nutritive value of different fractions of the
53 stover. *Animal Feed Science and Technology* 1999; 81:1-16.
54
55
56
57
58
59
60
61
62
63
64
65

- 1
2
3 [24] Myers DK, Underwood JF. Harvesting corn residue. Agronomy facts AGF-
4 003-92. The Ohio State University Extension; 1992.
5 <<http://ohioline.osu.edu/agf-fact/0003.html>> [Accessed 3 Jul 2008]
6
7 [25] Tolera A, Sundstøl F, Said AN. The effect of stage of maturity on yield and
8 quality of maize grain and stover. *Animal Feed Science and Technology*
9 1998; 75:157-68.
10
11 [26] Russell JR. Influence of harvest date on the nutritive value and ensiling
12 characteristics of maize stover. *Animal Feed Science and Technology* 1986;
13 14:11-27.
14
15 [27] Pordesimo LO, Sokhansanj S, Edens WC. Moisture and yield of corn stover
16 fractions before and after grain maturity. *Transactions of the ASAE* 2004;
17 47:1597-1603.
18
19 [28] Sokhansanj S, Turhollow AF. Baseline cost for corn stover collection.
20 *Applied Engineering in Agriculture* 2002; 18:525-30.
21
22 [29] Sewalt VJH, Fontenot JP, Allen VG, Glasser WG. Fiber composition and *in*
23 *vitro* digestibility of corn stover fractions in response to ammonia treatment.
24 *Journal of Agricultural and Food Chemistry* 1996; 44:3136-42.
25
26 [30] Hatfield RD, Ralph J, Grabber JH. Cell wall cross-linking by ferulates and
27 diferulates in grasses. *Journal of the Science of Food and Agriculture* 1999;
28 79:403-7.
29
30 [31] Tarkow H, Feist WC. A mechanism for improving the digestibility of
31 lignocellulosic materials with dilute alkali and liquid ammonia. *Advances in*
32 *Chemistry Series* 1969; 95:213-29.
33
34 [32] Zaldivar J, Nielsen J, Olsson L. Fuel ethanol production from lignocellulose: a
35 challenge for metabolic engineering and process integration. *Applied*
36 *Microbiology and Biotechnology* 2001; 56:17-34.
37
38 [33] Reijnders L. Ethanol production from crop residues and soil organic carbon.
39 *Resources, Conservation and Recycling* 2008; 52:653-8.
40
41 [34] Shinnors KJ, Binversie BN, Muck RE, Weimer PJ. Comparison of wet and
42 dry corn stover harvest and storage. *Biomass & Bioenergy* 2007; 31:211-21.
43
44 [35] Shinnors KJ, Adsit GS, Binversie BN, Digman MF, Muck RE, Weimer PJ.
45 Single-pass, split-stream harvest of corn grain and stover. *Transactions of*
46 *the ASABE* 2007; 50(2):355-63.
47
48 [36] Varga E, Szengyel Z, Réczey K. Chemical pretreatments of corn stover for
49 enhancing enzymatic digestibility. *Applied Biochemistry and Biotechnology*
50 2002; 98-100:73-87.
51
52 [37] Silverstein RA, Chen Y, Sharma-Shivappa RR, Boyette MD, Osborne J. A
53 comparison of chemical pretreatment methods for improving saccharification
54 of cotton stalks. *Bioresource Technology* 2007; 98:3000-11.
55
56
57
58
59
60
61
62
63
64
65

1
2
3 **Figure Legends**
4
5

6 **Figure 1:** Effect of AFEX pretreatment ammonia loading and addition of xylanase
7 during enzymatic hydrolysis on glucose (A) and xylose (B) monomeric sugar yields.
8 All AFEX runs were kept at constant moisture content (60% dwb), temperature
9 (90°C) and residence time (5 min). Yields are in terms of untreated dry biomass.
10

11
12 **Figure 2:** Effect of AFEX pretreatment temperature on glucose and xylose
13 monomeric sugar yields following enzymatic hydrolysis. All AFEX runs were kept
14 at a constant moisture content (60% dwb), ammonia loading (1.0 g NH₃ g⁻¹ dry
15 biomass) and residence time (5 min). Yields are in terms of untreated dry
16 biomass. Glu = glucose, Xyl = xylose, MTSY = maximum theoretical sugar yield.
17
18

19 **Figure 3:** Effect of changes to AFEX moisture content and residence time on
20 monomeric glucose and xylose yields following enzymatic hydrolysis. Base AFEX
21 conditions: moisture content (60% dwb), ammonia loading (1.0 g NH₃ g⁻¹ dry
22 biomass), temperature (90°C) and residence time (5 min). Yields are in terms of
23 untreated dry biomass. MC = moisture content, RT = residence time, Glu =
24 glucose, Xyl = xylose, MTSY = maximum theoretical sugar yield.
25
26

27 **Figure 4:** Interaction effect plot of key parameter interaction with corn stover
28 fraction and harvest period on monomeric glucose (A) and xylose (B) yields
29 following 72 hours of enzymatic hydrolysis. Yields are in terms of untreated dry
30 biomass. MC = moisture content, RT = residence time, DWB = dry weight basis, N
31 = no xylanase added, Y = xylanase added (10% of total cellulase protein).
32
33
34

35 **Figure 5:** Estimated dry matter distribution for late harvest corn stover and harvest
36 scenarios based on 70% and 30% dry weight corn stover harvests. Percentages
37 of individual fraction harvested are based on the total amount of each fraction
38 available.
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Tables

Table 1: Corn stover composition for early and late harvest stover fractions. Values with different superscripts in an individual column were statistically different using Tukey's pairwise comparison with $\alpha = 0.05$. [The 'other' column determined by difference from 100%]

Corn Stover Fraction Composition (% dry biomass)						
Corn Stover Fraction		Glucan	Xylan	Acid-Insoluble Lignin	Ash	Other
Early	Leaves	27.5 ^b ± 3.2	17.8 ^e ± 1.7	13.2 ^{bc} ± 0.7	7.3 ^a ± 0.13	34.2
	Stem	35.1 ^a ± 2.6	19.0 ^{de} ± 1.1	14.9 ^{bc} ± 0.2	3.4 ^c ± 0.10	27.6
Late	Leaves	35.3 ^a ± 1.2	21.8 ^{cd} ± 0.6	13.6 ^{bc} ± 1.7	6.0 ^b ± 0.25	23.3
	Stem	37.8 ^a ± 0.9	23.6 ^{bc} ± 0.4	16.9 ^b ± 0.5	2.4 ^d ± 0.08	19.3
	Husk	39.0 ^a ± 2.2	26.5 ^b ± 1.5	11.6 ^c ± 0.3	2.1 ^e ± 0.11	20.8
	Cob	27.5 ^b ± 1.1	32.3 ^a ± 1.3	25.8 ^a ± 2.6	1.1 ^f ± 0.02	13.3

Table 2: Analysis of Variance for Factors influencing Sugar Yields. *significant at $\alpha = 0.05$

Factor	P-value			
	24 hr Glucose	72 hr Glucose	24 hr Xylose	72 hr Xylose
Harvest Date	0.775	0.437	0.000*	0.000*
Corn Stover Fraction	0.000*	0.006*	0.526	0.528
Ammonia Loading	0.000*	0.000*	0.000*	0.000*
Temperature	0.082	0.161	0.000*	0.022*
Moisture Content	0.018*	0.002*	0.000*	0.000*
Residence Time	0.001*	0.000*	0.003*	0.000*
Xylanase Addition	0.001*	0.002*	0.000*	0.000*
Harvest x Ammonia	0.007*	0.001*	0.001*	0.002*
Harvest x Temperature	0.918	0.932	0.824	0.392
Harvest x Moisture	0.687	0.762	0.943	0.424
Harvest x Res. Time	0.829	0.719	0.377	0.317
Harvest x Xylanase	0.919	0.760	0.111	0.063
Fraction x Ammonia	0.288	0.080	0.416	0.152
Fraction x Temperature	0.746	0.684	0.588	0.400
Fraction x Moisture	0.278	0.075	0.163	0.109
Fraction x Res. Time	0.916	0.859	0.715	0.542
Fraction x Xylanase	0.711	0.300	0.008*	0.030*

Table 3: Optimized harvest scenarios based on desired sugar and AFEX ammonia loading

Harvest Scenario	A	B	C
Optimized Sugar	Glucose/Total	Xylose	Xylose
Ammonia Loading (g NH₃ g⁻¹ dry SHCS)	1.0, 1.5	1.0	1.5
Best Fraction	Husk	Husk	Cob
	Leaf	Stem	Husk
	Stem	Cob	Stem
Worst Fraction	Cob	Leaf	Leaf

Table 4: Estimated yields for 70% collection of selectively harvested corn stover (SHCS) following AFEX, enzymatic hydrolysis and fermentation.

		1.0 g NH ₃ g ⁻¹ dry SHCS		1.5 g NH ₃ g ⁻¹ dry SHCS	
		Harvest Scenario A	Harvest Scenario B	Harvest Scenario A	Harvest Scenario C
Sugar Yield <i>g sugar kg⁻¹ dry SHCS</i>	Glucose	273.7	254.2	331.5	310.8
	Xylose	150.0	153.1	195.7	206.8
	Total	423.6	407.3	527.2	517.5
Hectare Yields <i>t ha⁻¹</i>	Total Sugars	2.55	2.45	3.17	3.12
	Theoretical Ethanol	1.30	1.25	1.62	1.59

Table 5: Estimated yields for 30% collection of selectively harvested corn stover (SHCS) following AFEX, enzymatic hydrolysis and fermentation.

		1.0 g NH ₃ g ⁻¹ dry SHCS		1.5 g NH ₃ g ⁻¹ dry SHCS	
		Harvest Scenario A	Harvest Scenario B	Harvest Scenario A	Harvest Scenario C
Sugar Yield <i>g sugar kg⁻¹ dry SHCS</i>	Glucose	305.1	278.2	354.4	311.3
	Xylose	151.8	161.3	192.7	215.4
	Total	456.9	439.5	547.1	526.7
Hectare Yields <i>t ha⁻¹</i>	Total Sugars	1.18	1.13	1.41	1.36
	Theoretical Ethanol	0.60	0.58	0.72	0.69

1
2
3 Table 6: Enzymatic hydrolysis
4 xylanase loading in terms of xylan
5 content of each fraction

		Xylanase Loading
Corn Stover Fraction		<i>mg xylanase g⁻¹ xylan</i>
Early	Leaves	4.78
	Stem	5.73
Late	Leaves	5.03
	Stem	4.97
	Husk	4.57
	Cob	2.64

Figure 1
[Click here to download high resolution image](#)

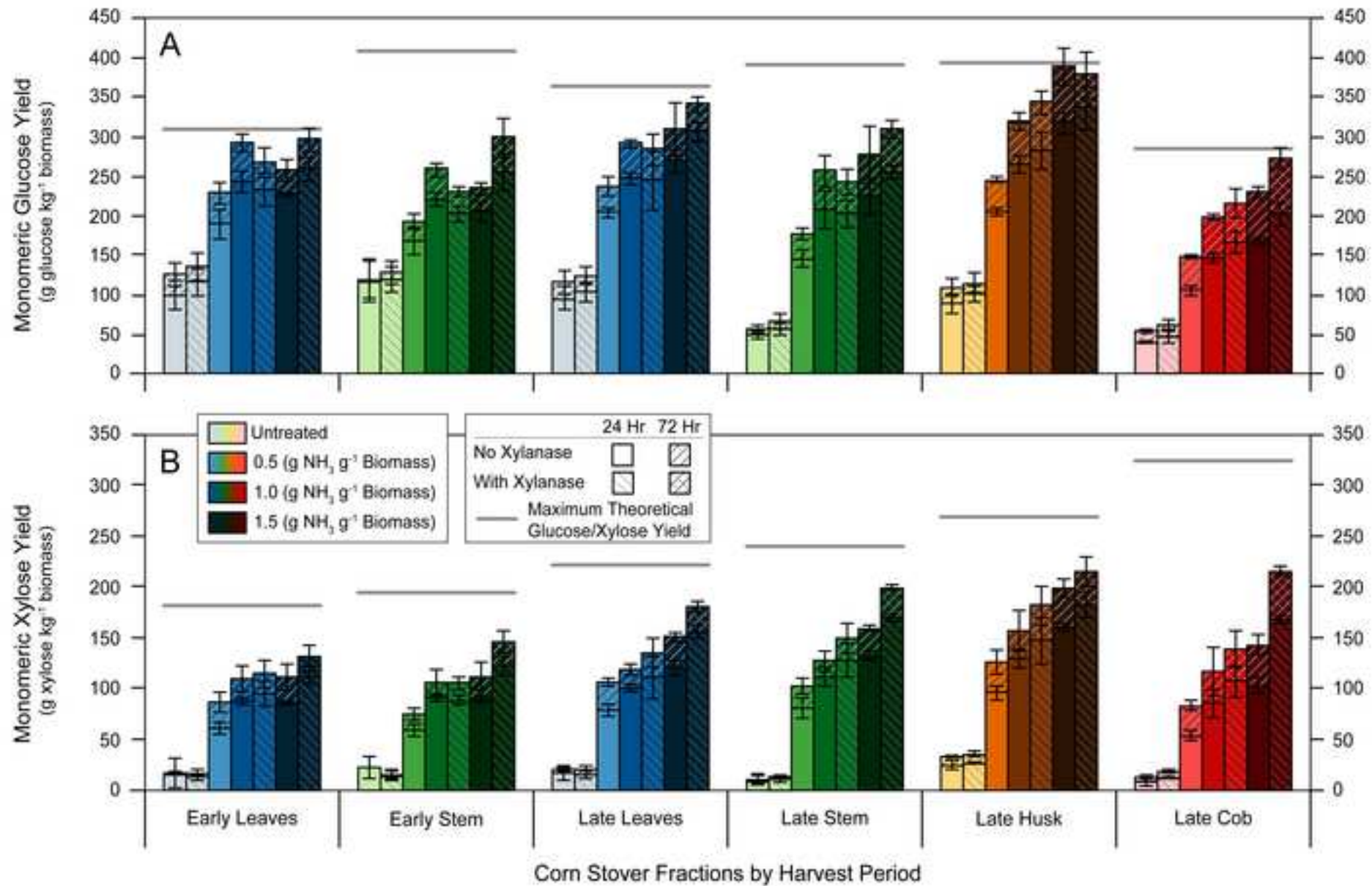


Figure 1 - grayscale
[Click here to download high resolution image](#)

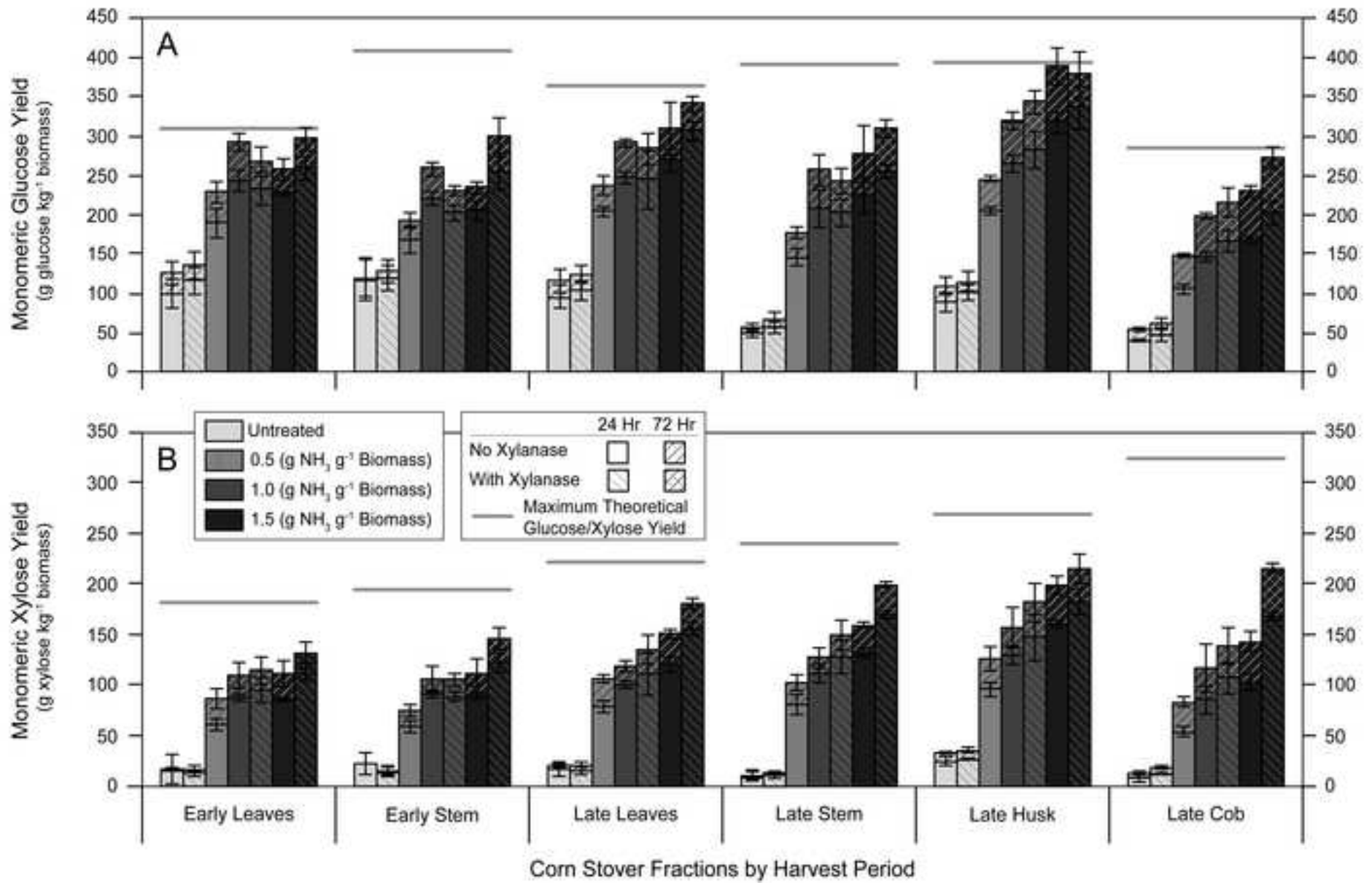


Figure 2
[Click here to download high resolution image](#)

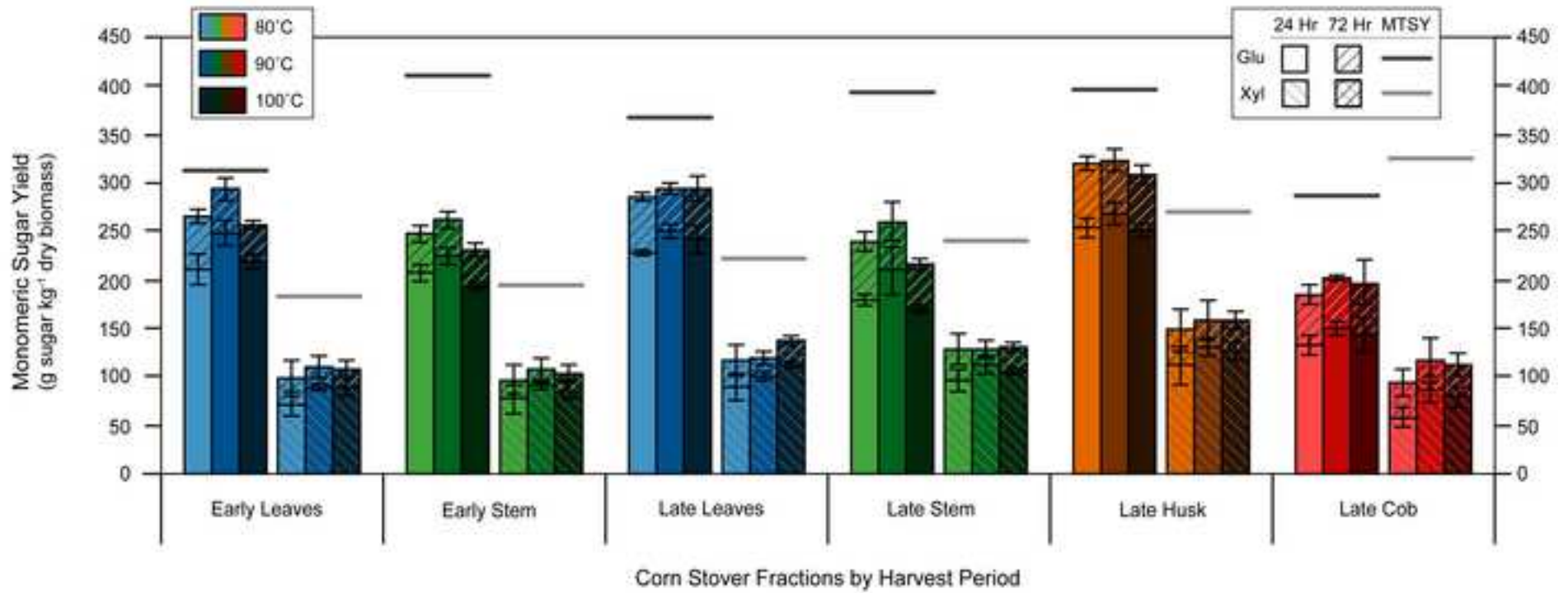


Figure 2 - grayscale
[Click here to download high resolution image](#)

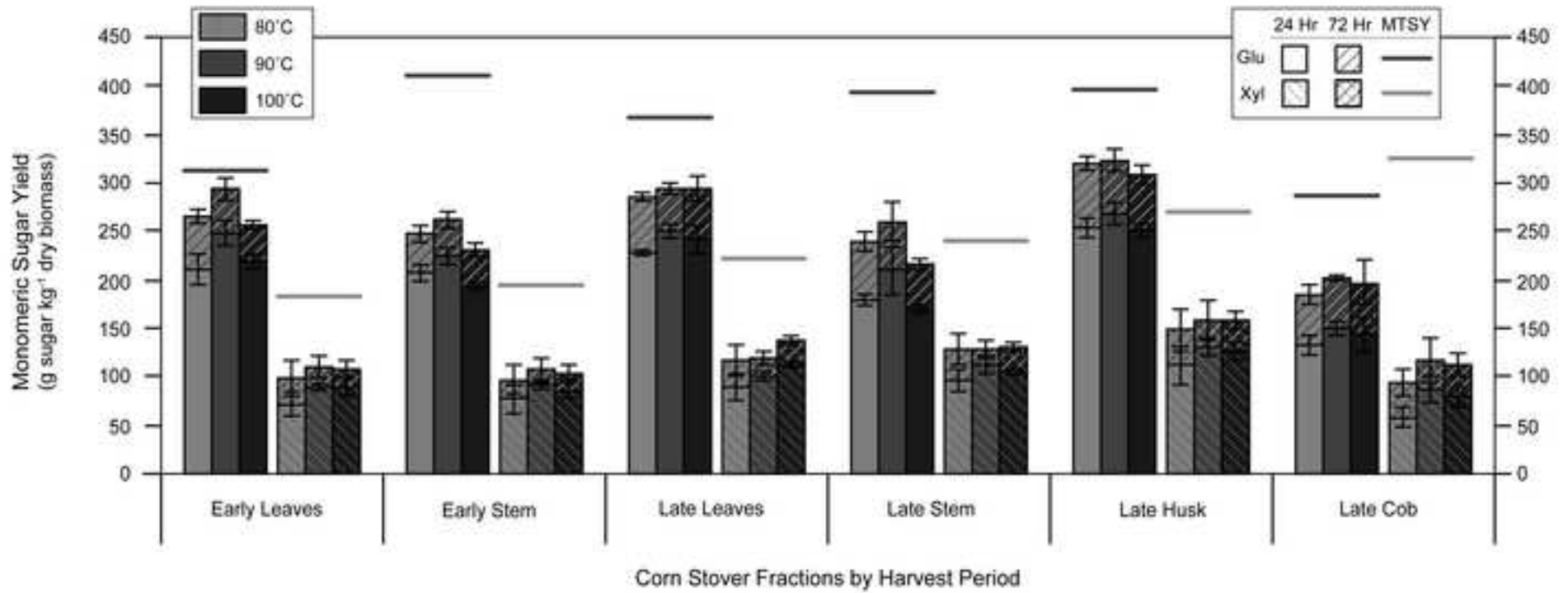


Figure 3
[Click here to download high resolution image](#)

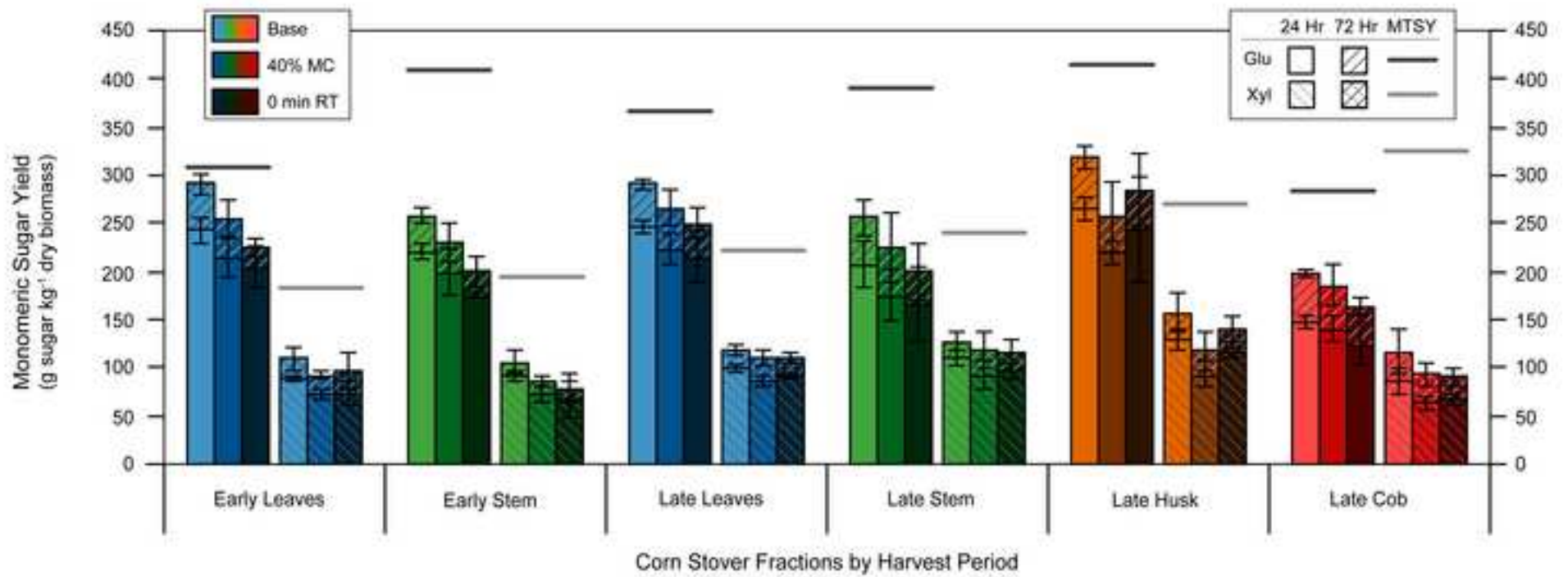


Figure 3 - grayscale
[Click here to download high resolution image](#)

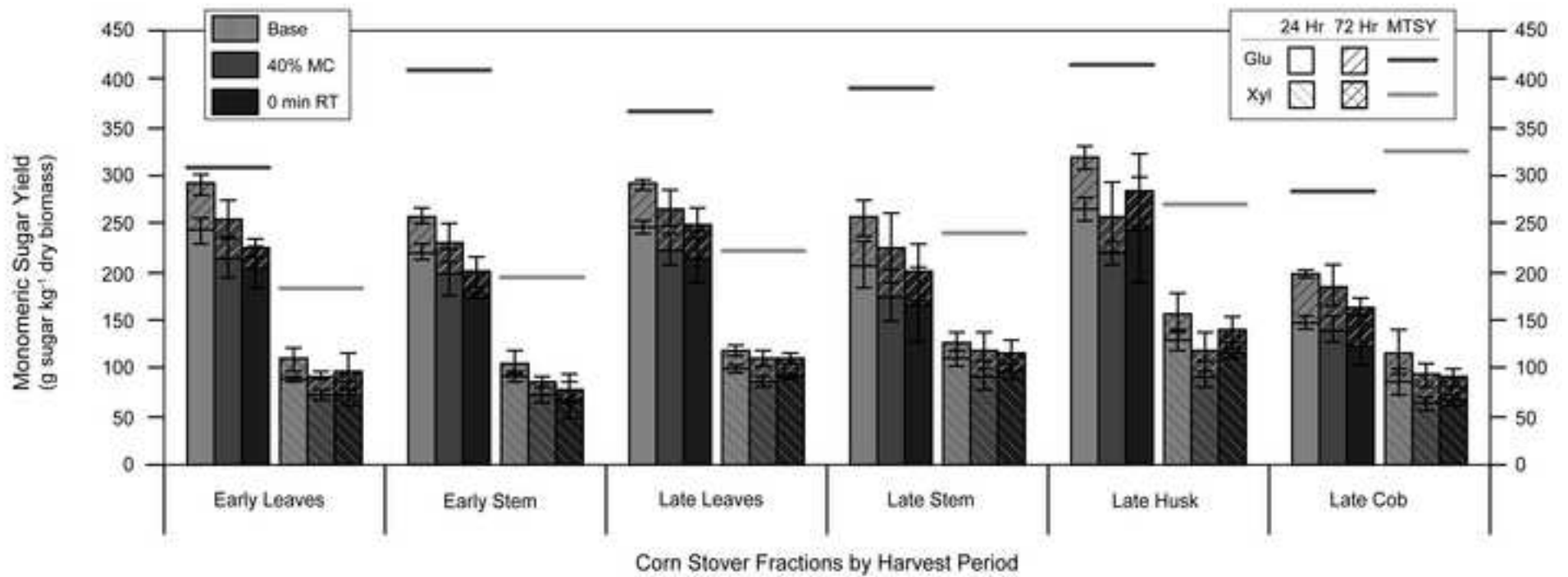


Figure 4
[Click here to download high resolution image](#)

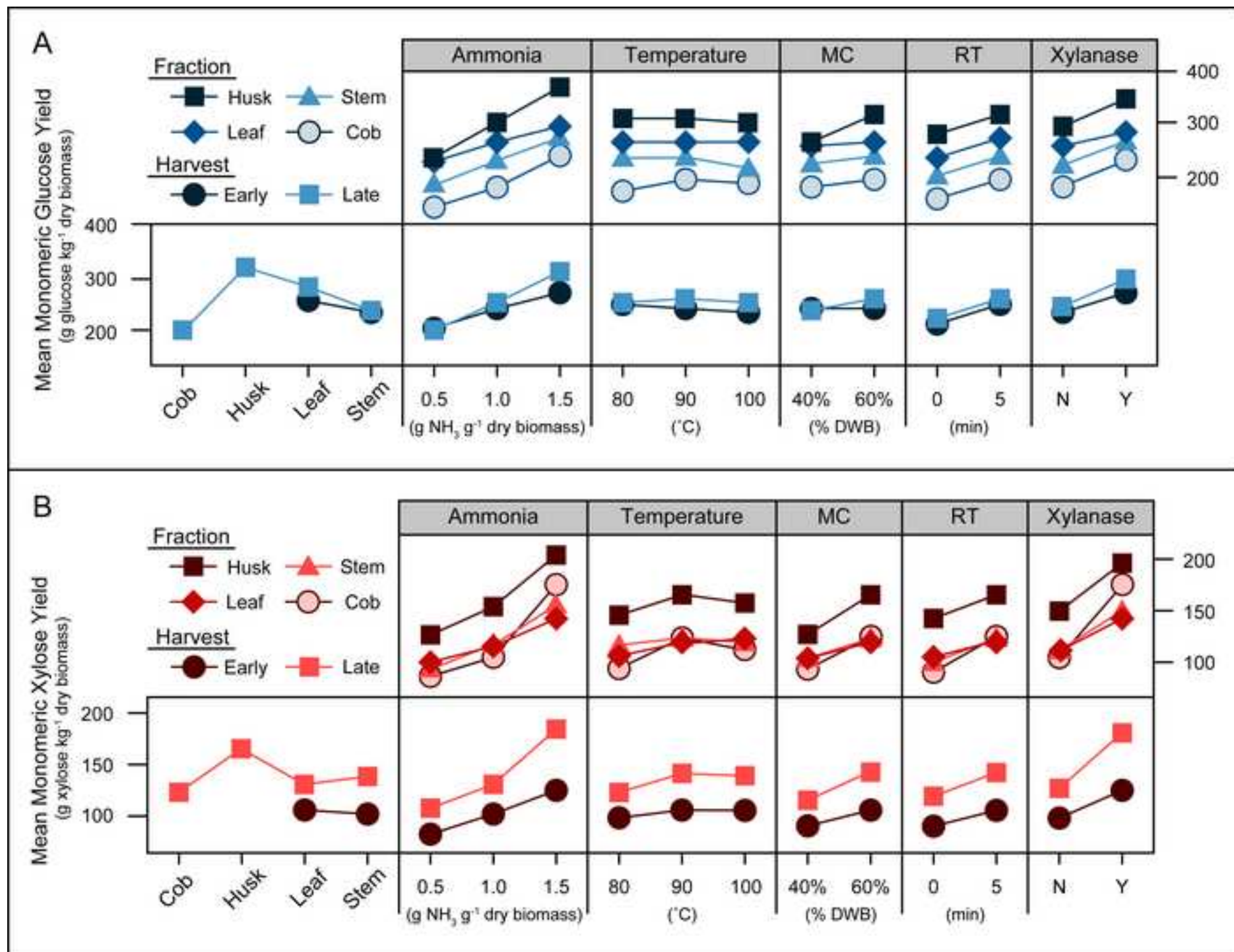


Figure 4 - grayscale
[Click here to download high resolution image](#)

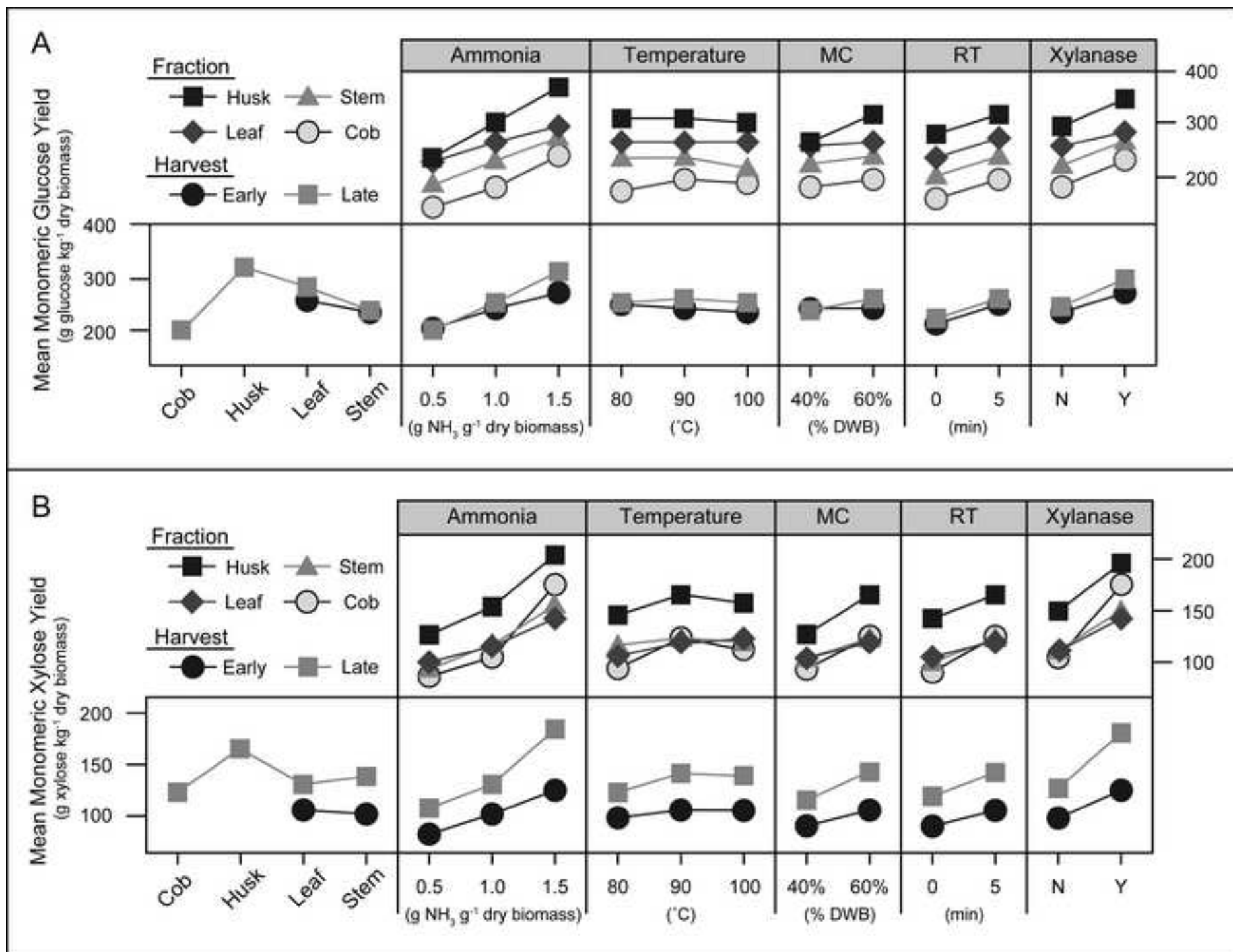


Figure 5
[Click here to download high resolution image](#)

Late Harvest Corn Stover
 Estimated Dry Matter Distribution

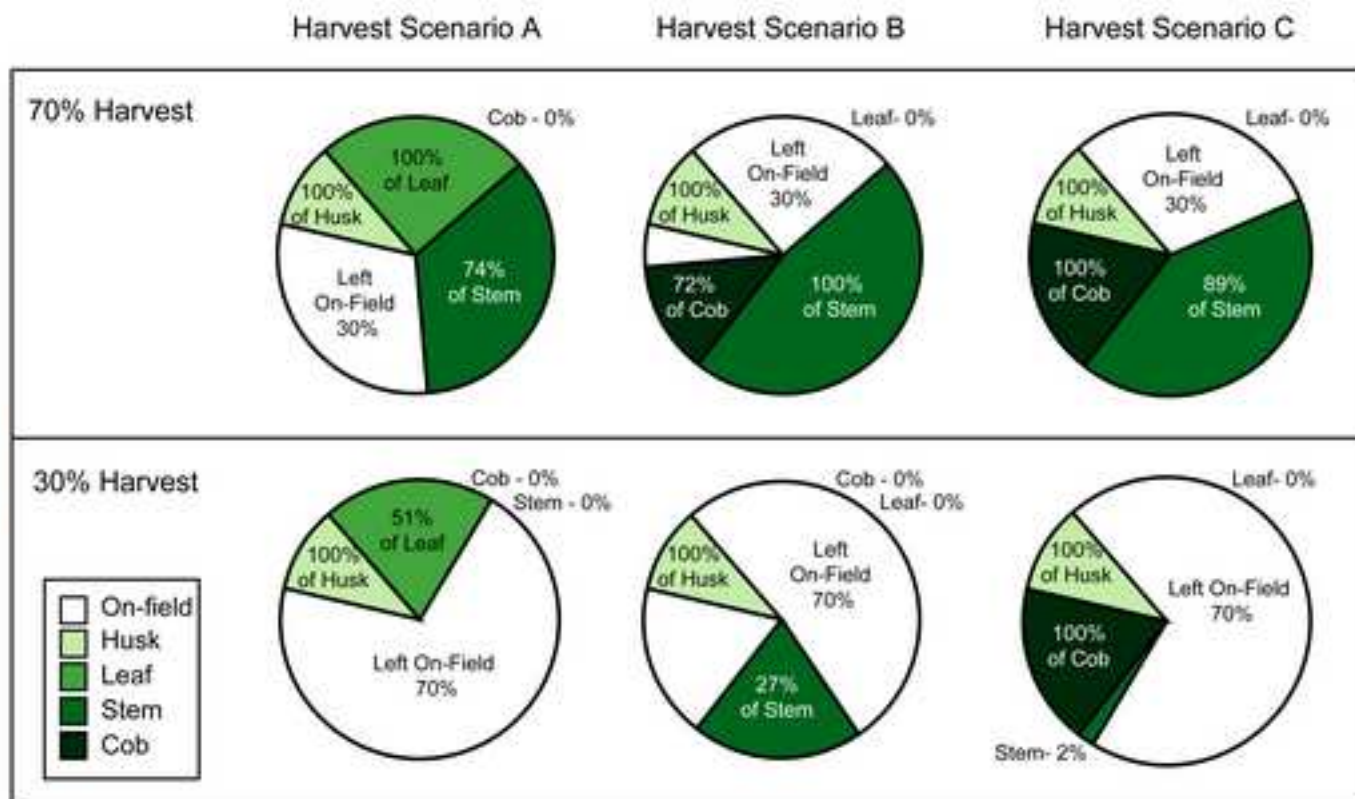
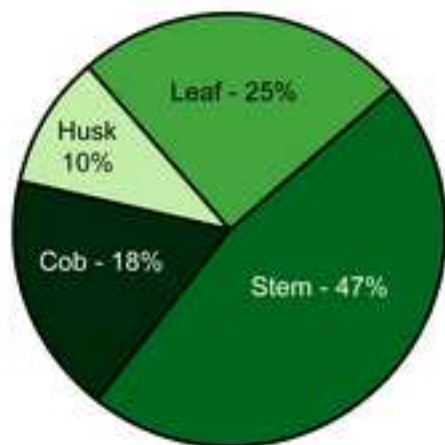


Figure 5 - grayscale
[Click here to download high resolution image](#)

Late Harvest Corn Stover
 Estimated Dry Matter Distribution

