By
JASON M. MORSON
A dissertation submitted to the
Graduate School - New Brunswick
Rutgers, The State University of New Jersey
In partial fulfillment of the requirements for the degree of
Doctor of Philosophy
Graduate Program in Ecology and Evolution
Written under the direction of
Dr. Daphne Munroe
And approved by
$\qquad$
$\qquad$
$\qquad$
$\qquad$
$\qquad$
New Brunswick, NJ
JANUARY 2018

ABSTRACT OF THE DISSERATION<br>UNDERSTANDING THE ECOLOGICAL PROCESSES THAT DRIVE PATTERNS IN FISHERY SELECTIVITY AND SURVEY CATCHABILITY By<br>JASON M. MORSON<br>Dissertation Director:<br>Dr. Daphne Munroe

Being able to confidently count organisms is fundamental to estimating the dynamics of populations and to making inferences about what influences those dynamics. However, an accurate census is difficult to achieve because an entire population is almost never fully detectable. The population size of aquatic organisms can be particularly problematic to estimate because the environment is not conducive to human observation, and, particularly in the ocean, the entire habitat of the population in question can never be fully sampled. When we attempt to count the numbers of fish or shellfish in a population, for example to help manage a fishery or to increase our understanding of how a population is responding to shifts in climate, this impediment almost always results in at least some portion of the population being unobserved and unquantified. Having some information about the size and composition of this unobserved demographic is fundamental to population ecology and is particularly
essential when the relative contribution of the unobserved demographic to the true population changes in space and time.

Advancements in fisheries population dynamics and stock assessment science have resulted in the generally accepted application of two parameters that aid in estimating the unobserved portion of the population from the observed one. The first, selectivity, defines the proportion of a given demographic group, available to, and retained by, survey or fishing gear once it comes into contact with it. The second, catchability, is defined as the proportion of the population caught by a single unit of fishing or survey effort. Despite the influence these two parameters have on our perception of what controls fish and shellfish population dynamics, we still don't fully understand the underlying processes that influence them for many managed fisheries.

Using fishery and population survey data from two mid-Atlantic fisheries, summer flounder and Eastern oysters, I use the following four chapters to 1.) identify patterns in selectivity and catchability and understand the underlying ecological processes that drive them, and 2.) propose how we might better utilize this information to assess and manage these and other fisheries.

In Chapter 1 I use summer flounder data collected from commercial and recreational landings and a stock assessment trawl survey to evaluate the selectivity of the survey and fishing gear for different demographic groups. Some interesting patterns were identified, particularly that selectivity for female summer flounder is higher in the recreational fishery than in both the commercial fishery or the stock assessment trawl survey. This pattern suggested a highly female-biased recreational catch and that male and female summer flounder separate in space and time.

In Chapter 2 I explored, given the size- and sex-specific selectivity patterns and catch composition identified in Chapter 1, whether management actions could be taken to achieve a more sex-balanced harvest in the recreational fishery. I evaluated whether a series of slot limits, size regulations that require landed fish be between some minimum and maximum size, have the potential to simultaneously reduce mortality on large, fecund, females while maintaining or reducing total fishing mortality.

The patterns in summer flounder fishery catch composition and selectivity and the prescribed management actions identified and discussed in Chapters 1 and 2 should contribute significantly to our understanding of the life history, particularly relative to sex-specific habitat use, of summer flounder, and will likely be relevant to the stock assessment and management of this commercially and recreationally important fishery going forward.

In Chapter 3 I used a set of field experiments to derive empirical estimates of catchability for a survey dredge used in the stock assessment of eastern oysters in Delaware Bay. I identified an along-bay gradient in catchability that appeared to be driven by changes in oyster density. This density-dependent catchability lead to catch-per-unit-effort of the survey dredge being hyperstable at low oyster density, making catch-per-unit-effort (CPUE) an unreliable proxy for abundance at low oyster density.

In Chapter 4 I asked the question, given that evidence from Chapter 3 suggests a fixed catchability coefficient is not appropriate for estimating true density from survey CPUE, how do three alternative models perform in estimating the true density in the sampled area. In the first model, I corrected CPUE by applying spatially-explicit catchability coefficients, as opposed to a constant, that account for the along-bay gradient in density.

In the second, CPUE was corrected for by estimating catchability in situ for each tow using a logistic model fit to catch composition and tow covariates. For the third model, CPUE data were ignored entirely and a model that accounted for the proportion of the sample composition that was made up of oysters was applied to estimate oyster density in situ for each tow. The simplest model, which ignored both catchability of the survey gear and CPUE, and relied only on an estimate of the portion of the catch that was made up of oysters, performed best in estimating the true density in the sampled area.

The density-dependence in catchability identified for an oyster survey dredge in Chapter 3 is an important finding because it adds to a growing body of literature that density-dependent catchability, a phenomenon traditionally attributed to an interaction between fish and fishermen behavior, may be a common problem in standardized stock assessment survey data as well. In addition, Chapter 4 strongly suggests that when catchability varies at fine spatial and temporal scales, raw catch components may more accurately reflect the true density in the sampled area than an index derived from catch-per-unit-effort and catchability. Both findings should have application to how reef growing populations of organisms, and oysters in particular, are assessed and managed going forward.

## ACKNOWLEDGEMENTS

First and foremost, I would like to thank my PhD advisor, Daphne Munroe, for her continued support and guidance over the last several years. I am extremely grateful for Daphne's willingness to let me pursue research avenues that were interesting to me because it created an encouraging and enjoyable learning environment. My committee, Ken Able, David Bushek, Olaf Jensen, and Pat Sullivan all made themselves readily available to answer questions and provide thoughtful input on my research whenever needed. I am thankful for the chance to have interacted with and learned from these incredible mentors.

I benefited a great deal from having had the opportunity to work with the friendly people at both the Haskin Shellfish Research Laboratory and the Rutgers University Marine Field Station. The mix of faculty, staff, and graduate students at both facilities allowed me to get feedback on my research from individuals with a variety of perspectives and backgrounds. I am thankful for the relationships I built at both places and look forward to continued collaboration and friendship with the people there in the future.

Finally, this entire endeavor would have been impossible without the support of my family. My wife, Crystal, provided constant love, support, and encouragement throughout this process, even when it meant she had to take on the bulk of the parenting duties. My two boys, Jason and Matthew, provided much needed distractions when times became stressful and were understanding when I needed to work. And my parents, John and Linda, and siblings, Ryan, Daniel, and Laura have throughout this
journey and throughout my entire life, inspired me to pursue my dreams with constant and unwavering encouragement.

## TABLE OF CONTENTS

ABSTRACT ..... ii
ACKNOWLEDGEMENTS ..... vi
LIST OF TABLES ..... ix
LIST OF ILLUSTRATIONS ..... xi
INTRODUCTION ..... 1
CHAPTER 1:
Estimating the sex composition of the summer flounder catch using fishery-independent data .....  8
CHAPTER 2 :
Evaluating the potential for a sex-balanced harvest approach in the recreational summer flounder Paralichthys dentatus fishery ..... 44
CHAPTER 3:
Density-dependent capture efficiency of a survey dredge and its influence on the stock assessment of Easter oysters Crassostrea virginica in Delaware Bay ..... 71
CHAPTER 4:
Alternative models to estimate density of Eastern oysters Crassostrea virginica using a hydraulic dredge ..... 97
CONCLUSIONS ..... 120

## LIST OF TABLES

Table 1.1. Total number of fish sampled for sex and length and the subset of those fish that were also aged by examining annuli in otoliths and scales (see Methods).

Table 1.2. Locations where samples were collected from the recreational and commercial Summer Flounder fisheries.

Table 1.3. Total number of fish sampled, by NMFS-NEFSC statistical area (see Figure 1.2), from the NMFS-NEFSC bottom trawl survey, the commercial fishery, and the recreational fishery.

Table 1.4. Akaike information criterion (AIC) for eight potential models describing the probability of a given age provided the length (null model) and the sex, region (north of $40^{\circ}$ latitude or south of $40^{\circ}$ latitude), and data source (data collected from the recreational fishery, the commercial fishery, or a fishery-independent trawl survey).

Table 2.1. Length and depth categories for sub-sampling the sex of discarded fish.
Table 2.2. Total number of summer flounder sampled by port, depth range, outcome (landed/discarded), and sex. A dash indicates a given cell was not sampled while a zero indicates there were zero fish for that cell.

Table 2.3. Observed (18-inch minimum size limit, top row) and simulated incremental slot limit performance metrics, including total number dead ( $\mathrm{N}_{\mathrm{d}}$ ), dead female biomass $\left(\mathrm{F}_{\mathrm{d}}\right)$, and ratio of dead discards to total dead ( $\mathrm{R}_{\mathrm{d}}$ ). In parentheses are the percent changes in a given performance metric relative to the observed (18-inch minimum size limit). Slot limits in bold are within $+/-10 \%$ of the observed total number dead.

Table 3.1. Oyster abundance and exploitation rate calculated using different catchability coefficients for each of the managed regions in 2015 (See Figure 3.1). The proportional change in abundance from the catchability coefficients using 2003 data only, to the application identified with this work as being most appropriate (1999, 2000, 2003, and 2013 data), is given in parentheses next to each abundance estimate in the lower half of the table. * The Low Mortality region was split out into separate sections (Arnolds / Upper Arnolds and Round Island) based on the results of analysis for spatial trends in capture efficiency (see Results section and Figure 3.4).

Table 3.2. Power function model parameters describing density-dependent capture efficiency.

Table 4.1. Coefficients for model parameters for average, logistic, and power models. In parentheses next to each coefficient is the p-value associated with that parameter. NS = not selected by backward stepwise AIC (see Methods).

Table 4.2. Model performance estimated using the mean squared error (MSE) for each modeling approach (average, logistic, and power) and each training-test data combination. The overall reported mean for each modeling approach is the mean of the MSE scores for each training-test combination.

## LIST OF ILLUSTRATIONS

Figure 1.1. Photographs showing a technique for determining the sex ( $A=$ Female; $B=$ Male) of a Summer Flounder without dissecting the entire fish. A small incision is made along the gut cavity and the gonads are pulled through the incision with a pair of forceps.

Figure 1.2. Map showing National Marine Fisheries Service, Northeast Fisheries Science Center statistical area units along the northeast USA and the western midAtlantic continental shelf (NEFSC 1997). The shapes around the statistical area number indicate the data source as NMFS-NEFSC bottom trawl survey (NFS), commercial fishery (CFS), or recreational fishery (RFS). Rectangles represent RFS only, diamonds represent CFS only, pentagons represent NFS only, triangles represent RFS and CFS, hexagons represent NFS and CFS, and circles represent RFS, CFS, and NFS.

Figure 1.3. Proportion female Summer Flounder at-age in the commercial fishery (blue dots) and the National Marine Fisheries Service, Northeast Fisheries Science Center (NMFS-NEFSC) bottom trawl survey (green dots). Lines represent the probability of landing a female estimated by logistic regression (see text). Commercial fishery data are compared to (A) raw NMFS-NEFSC bottom trawl survey data, (B) NMFS-NEFSC bottom trawl survey data that were left-truncated at 35 cm to simulate the minimum retention size in the commercial fishery, and (C) NMFS-NEFSC bottom trawl survey data that were left-truncated at 35 cm and assigned age based on age-length keys generated from commercial fishery data.

Figure 1.4. Proportion female Summer Flounder at-age in the recreational fishery (orange dots) and the National Marine Fisheries Service, Northeast Fisheries Science Center (NMFS-NEFSC) bottom trawl survey (green dots). Lines represent the probability of landing a female estimated by logistic regression (see text). Recreational fishery data are compared to (A) raw NMFS-NEFSC bottom trawl survey data, (B) NMFS-NEFSC bottom trawl survey data that were left-truncated at 46 cm to simulate the smallest minimum retention size in the recreational fishery, and (C) NMFS-NEFSC bottom trawl survey data that were left-truncated at 46 cm and assigned age based on age-length keys generated from recreational fishery data.

Figure 1.5. Proportion female Summer Flounder at-age in the recreational fishery (orange dots) and the National Marine Fisheries Service, Northeast Fisheries Science Center (NMFS-NEFSC) bottom trawl survey (red dots). Lines represent the probability of landing a female estimated by logistic regression (see text). Recreational fishery data are compared to (A) raw NMFS-NEFSC bottom trawl survey data, (B) NMFS-NEFSC bottom trawl survey data that were left-truncated at 53 cm to simulate the largest minimum retention size in the recreational fishery, and (C) NMFS-NEFSC bottom trawl survey data that were left-truncated at 53 cm and assigned age based on age-length keys generated from recreational fishery data.

Figure 1.6. Box and whisker plots of the mean length-at-age of Summer Flounder under different minimum retention size regulations. Plots are organized by region (north and south) and sex (male and female). Black horizontal lines are the mean, boxes are the interquartile ranges, black vertical lines are the $5^{\text {th }}$ and $95^{\text {th }}$ percentiles, and dots are outliers.

Figure 1.7. Proportion female Summer Flounder at-length in the commercial fishery (blue dots), the recreational fishery (orange dots), and the National Marine Fisheries Service, Northeast Fisheries Science Center (NMFS-NEFSC) bottom trawl survey (green dots). Lines represent the probability of landing a female estimated by logistic regression (see text). NMFS-NEFSC bottom trawl survey data are truncated according to the minimum retention size in the $(A)$ commercial fishery ( 35 cm ) and (B) recreational fishery ( 46 cm ).

Figure 2.1. Map of sampling locations.
Figure 2.2. Total number (A) and weight (B) of male and female Summer Flounder collected at each 1-inch length bin. Solid black line represents proportion female-atlength. Vertical dashed line represents the 18-inch minimum landing size in 2016.

Figure 2.3. Total number of male and female Summer Flounder collected at (A) Cape May, NJ, (B) Atlantic Highlands, NJ, (C) Captree, NY, (D) Montauk, NY, and (E) Pt. Judith, RI in each 1-inch length bin. Solid black line represents proportion female-at-length. Note differences in primary $y$-axis for each panel.

Figure 2.4. Total number of male and female Summer Flounder collected in (A) 0-25 feet (B) 25-50 feet (C) 50-75 feet, and (D) 75+ feet of water at each 1-inch length bin. Solid black line represents proportion female-at-length. Note differences in primary yaxis for each panel.

Figure 3.1. Map of sampling locations.
Figure 3.2. Length frequency (A) and probability density length distribution (B) for oysters collected at the same locations with a commercial oyster dredge and patent tongs.

Figure 3.3. Box and whiskers plot of mean capture efficiency of a commercial oyster dredge estimated in 2000, 2003, and 2013 at the same sampling locations. Bold horizontal lines represent the mean, boxes encompass the interquartile range, black whiskers extend to the $5^{\text {th }}$ and $95^{\text {th }}$ percentiles, and dots are outliers.

Figure 3.4. Box and whiskers plot of mean capture efficiency of a commercial oyster dredge at different oyster reef locations along the Delaware Bay, USA. Reef locations are organized on the $x$-axis from the lower bay (left side) to the upper bay (right side). Empty black boxes around the reef names represent the regional grouping of bedspecific catchability coefficients applied in the 2015 Delaware Bay oyster stock
assessment based on data collected from the 2003 experiments (Powell et al. 2007). Shaded, offset boxes, represent the regional groupings of bed-specific catchability coefficients identified as statistically appropriate with this work. Bold horizontal lines represent the mean, boxes encompass the interquartile range, black whiskers extend to the $5^{\text {th }}$ and $95^{\text {th }}$ percentiles, and dots are outliers.

Figure 3.5. Survey gear capture efficiency as a function of true oyster density. Error bars represent the standard deviation from 1,000 bootstrap simulations. Line indicates the best fit power model estimated by weighted nonlinear least squares. See methods section for details.

Figure 4.1. Map of sampling locations.
Figure 4.2. Capture efficiency as a function of two variables, total haul volume (left panels) and proportion of the sub-sample that was made up of cultch (right panels) for all three training-test pairs of data $(A=2003$ training data, $B=2013$ training data, $C=$ random training data).

Figure 4.3. Residuals (model-estimated density ed - true density $t d$ ) calculated from three approaches, a logistic model, a power model, and an average model, to estimating true density in the sampled area. See Methods for model descriptions.

Figure 4.4. The density of oysters in the sampled area as a function of the proportion of the sample that is made up oysters. (A) 2003 data were used to train and estimate power model parameters for the line while the points plotted are 2013, (B) 2013 data were used to train and estimate power model parameters while the points represent the 2003 data, and (C) a random draw of 24 of the 48 experimental observations (2003 and 2013 experimental data) were used to train and estimate parameters for the line while the points represent the remaining 24 data points.

## INTRODUCTION

While our ability to estimate the number of organisms in a population is central to applied ecology (Elphick 2008) and plays a major role in our understanding of biodiversity (Colwell and Coddington 1994) and species interactions (Faisal et al. 2010), identifying the most appropriate methodology used to count individuals, and the most appropriate data interpretations and transformations used to scale those counts to estimates of population size, continues to be a field of research rife with challenges (Jones 2011; Guthlin, et al. 2014; Maunder and Piner 2014). Some of the more common approaches to estimating abundance include stratified random sampling (Powel et al. 2008), mark-recapture methods (Grimm et al. 2014), depletion analyses (Young et al. 2004), and the application of complex models that account for the contribution of different demographic groups (Haddon 2011). Each method comes with a set of assumptions about how the true population relates to the sampled one, and attempts are made to account for these when drawing inferences about ecological phenomenon.

For population assessment scientists and natural resource managers and stakeholders, the necessity for reliable count data extends beyond the desire to advance theory and principles in ecology. When applied in a resource management context, unreliable, misused, or misinterpreted abundance data can result in ecological, social, and economic disasters. Marine fisheries management has a particularly storied, even if overstated, history of fisheries collapsing from what have been actual or perceived failures of population assessment scientists and/or the management system to
recognize important trends in the population dynamics of the species being managed (Hilborn and Hilborn 2012).

A major difficulty associated with counting and managing fisheries, as well as other mobile and inconspicuous organisms, was summarized famously by John Shepherd (1978) with, "Managing fisheries is hard; it's like managing a forest, in which trees are invisible and keep moving around." The fisheries literature is full of innovative attempts to overcome this reality (Hilborn and Walters 1992; Quinn and Deriso 1999). In addition, recent advancements in computing have made it possible to assess the fit of hundreds of non-linear model parameters simultaneously (Fournier et al. 2012) so that the evolution of population assessment might soon lead to models that mirror the complexity of a natural population. Yet, despite continued improvement to techniques for assessing population size and complex model fitting procedures, contemporary methods still cannot accurately depict key population parameters for the most recent years in a time series (Cadrin and Dickey-Collas 2014; Maunder and Piner 2014).

Selectivity and catchability are both stock assessment parameters that scale observations taken from a sample of a population to estimates of the true population size (Cadrin et al. 2016). Selectivity can be defined as the product of 1) the proportion of a given demographic group retained by survey or fishing gear once it comes into contact with it, also called contact selectivity, and 2) the proportion of the same demographic group available to the fishing or survey gear at the time and location it was fished, also called population selectivity (Maunder et al. 2014). Catchability is simply how much of the population is caught by a single unit of fishing or survey effort (Arreguin-Sanchez 1996). Although both are tightly linked with the life history and
behavior of the species being studied, experimental or field observation-based approaches to estimates are often neglected in favor of those derived inside the assessment model (Cadrin et al. 2016) where they are sometimes correlated with other model parameters (Arreguin-Sanchez 1996). Despite key stock assessment outputs, including total abundance and fishing mortality rate, being highly sensitive to misspecification of selectivity and catchability (Chen et al. 2003; Cardin et al. 2016), both remain poorly understood for most fisheries (Sampson and Scott 2011; Sampson 2013).

This work is aimed at understanding the driving forces influencing catchability and selectivity and intersects two fields, stock assessment science and natural history, that are now too often viewed as disparate. I present experimental and observational estimates of selectivity and catchability from two fisheries, summer flounder and eastern oysters. Two chapters are devoted to each fishery. In first chapter I endeavor to understand, through observation and experimentation, some of the underlying ecological processes that drive patterns in selectivity and catchability of fishing and survey gear. With a better understanding of these ecological processes, I then set out in the second chapter to evaluate how we might utilize this information to develop more appropriate tools and applications in fishery stock assessment and management.

I propose that one of the major complications with counting any organism is that our ability to do it well is directly dependent on how much we already know about the organism's life history (distribution, demography, growth rates, movement patterns). But such information can only be learned from properly designed population surveys, with unbiased interpretations of population data, so that a feedback loop forms
between the raw count data being used to model and assess the population and basic natural history information needed to collect the raw count data. The underlying processes (natural history) inform priors of the model parameters and generate more certain estimates of the size of the population, which in turn allow for stronger inference about the underlying processes, and so on. This is an admittedly trivial statement. To count, model, and manage a resource, of course one would need to understand the basic life history and ecology of that resource. However, financial and time constraints often lead to gross overgeneralizations of the underlying processes that make different species unique.

## Literature Cited - Introduction

Arreguin-Sanchez, F. 1996. Catchability: a key parameter for fish stock assessment. Reviews in Fish Biology and Fisheries 6: 221-242.

Cadrin, S. X. and M. Dickey-Collas. 2014. Stock assessment methods for sustainable fisheries. ICES Journal of Marine Science 72 (1): 1-6.

Cadrin, S. X., G. R. DeCelles, and D. Reid. 2016. Informing fishery assessment and management with field observations of selectivity and efficiency. Fisheries Research 184: 9-17.

Chen, Y. L. Chen, and K. I. Stergiou. 2003. Impacts of data quantity on fisheries stock assessment. Aquatic Sciences 65: 1-7.

Colwell, R. K. and J. A. Coddington. 1994. Estimating terrestrial biodiversity through extrapolation. Philosophical Transactions of The Royal Society B 345: 101-118.

Clark, W. G. 2014. Direct calculation of relative fishery and survey selectivities. Fisheries Research 158: 135-137.

Elphick, C. S. 2008. How you count counts: the importance of methods research in applied ecology. Journal of Applied Ecology 45: 1313-1320.

Faisal, A., F. Dondelinger, D. Husmeier, and C. M. Beale. 2010. Inferring species interaction networks from species abundance data: a comparative evaluation of various statistical and machine learning methods. Ecological Informatics 5(6): 451-464.

Fournier, D. A., H. J. Skaug, J. Ancheta, J. Ianelli, A. Magnusson, M. N. Maunder, A. Nielsen, and J. Silbert. AD model builder: using automatic differentiation for statistical inference of highly parameterized complex nonlinear models. Optimization Methods and Software 27(2): 233.249.

Glass, C. and J. Gunn. 2004. Influence of social behaviour and behavioural interactions in understanding temporal and spatial dynamics and their effect on availability and catchability. ICES Journal of Marine Science 61: 1236-1237.

Grimm, A., B. Gruber, and K. Henle. 2014. Reliability of different mark-recapture methods for population size estimation tested against reference population sizes constructed from field data. PLOS One 9(6): e98840.

Guthlin, D., I. Storch, and H. Kuchenhoff. 2014. Toward reliable estimates of abundance: comparing index methods to assess the abundance of a mammalian predator. PLOS One 9(4): e94537.

Haddon, M. 2011. Modeling and quantitative methods in fisheries. Taylor and Francis, New York, NY.

Hilborn, R. and U. Hilborn. Overfishing: what everyone needs to know. Oxford University Press, New York, NY.

Hilborn, R. and C. J. Walters. 1992. Quantitative fisheries stock assessment: choice, dynamics, and uncertainty. Chapman and Hall, New York, NY.

Holland, D. S. and G. E. Herrera. 2009. Uncertainty in the management of fisheries: contradictory implications and a new approach. Marine Resource Economics 24(3): 289-299.

Jones, J. P. G. 2011. Monitoring species abundance and distribution at the landscape scale. Journal of Applied Ecology 48(1): 9-13.

Manderson, J., L. Palamara, J. Kohut, and M. J. Oliver. 2011. Ocean observatory data are useful for regional habitat modeling of species with different vertical habitat preferences. Marine Ecology Press Series 438: 1-17.

Maunder, M. N., J. R. Sibert, A. Fonteneau, J. Hampton, P. Kleiber, and S. J. Harley. 2006. Interpreting catch per unit effort data to assess the status of individual stocks and communities. ICES Journal of Marine Science 63(8): 1373-1385.

Maunder, M. and K. R. Piner. 2014. Contemporary fisheries stock assessment: many issues still remain. ICES Journal of Marine Science 72(1): 7-18.

Maunder, M. N., P. R. Crone, J. L. Valero, and B. X. Semmens. 2014. Selectivity: theory, estimation, and application in fishery stock assessment models. Fisheries Research 158: 1-4.

Powell, E.N ., K.A. Ashton-Alcox, J.N. Kraeuter, S.E. Ford and D. Bushek. 2008. Long term trends in oyster population dynamics in Delaware Bay: Regime shifts and response to disease. J. Shellfish Res. 27:729-755.

Quinn, T. J. and R. B. Deriso. 1999. Quantitative fish dynamics. Oxford University Press, New York, NY.

Rose, G. A. and D. W. Kulka. 2011. Hyperaggregation of fish and fisheries: how catch-per-unit-effort increased as the northern cod (Gadus morhua) declined. Canadian Journal of Fisheries and Aquatic Sciences 56(S1): 118-127.

Sampson, D. B. and R. D. Scott. 2011. A spatial model for fishery age-selection at the population level. Canadian Journal of Fisheries and Aquatic Sciences 68(6): 1077-1086.

Sampson, D. B. 2013. Fishery selection and its relevance to stock assessment and fishery management. Fisheries Research 158: 5-14.

Shepherd, J. 1978. Thoughts and sayings. Retrieved from: jgshepherd.com/thoughts/
Young, I. A. G., G. J. Pierce, H. I. Daly, M. B. Santos, L. N. Key, N. Bailey, J. P. Robin, A. J. Bishop, G. Stowasser, M. Nyegaard, S. K. Cho, M. Rasero, and J. M. F. Pereira. Application of depletion methods to estimate stock size in the squid Loligo forbesi in Scottish waters (UK). Fisheries Research 69: 211-227.

## CHAPTER 1

# ESTIMATING THE SEX COMPOSITION OF THE SUMMER FLOUNDER CATCH USING FISHERY-INDEPENDENT DATA** 


#### Abstract

Models that account for sex-specific behavior and population dynamics are becoming more common in the stock assessment of sexually dimorphic fishes. However, such models can be data-intensive and require some knowledge or assumptions about the sex ratio of fishery landings. A recent stock assessment review of Summer Flounder Paralichthys dentatus identified the need to account for sex-specific fishing mortality in the assessment model; however, no data on the sex composition of the catch were available. Fishery-independent sex-specific information is collected annually for this species by the National Marine Fisheries Service, Northeast Fisheries Science Center on their ocean trawl survey. Sex-at-age from the survey could be applied to the fishery landings if the probability of landing a given sex at a given age is equivalent for fish collected by the survey and in the landings. To generate the first regionally comprehensive database on the sex ratio of Summer Flounder landings and to determine the efficacy of using survey sex-at-age keys to sex the landings, I recorded the sex composition of the commercial and recreational Summer Flounder catch (n= 31,912 ) in 2010 and 2011. When (1) survey length data were left-truncated to simulate the minimum retention sizes in the fisheries and (2) age-length keys generated from


[^0]fishery-dependent data were applied to length-frequency distributions from the survey to simulate the growth rates of fish landed in the catch, the sex-at-age pattern in the survey-derived data closely resembled the patterns in the catch. However, statistically significant differences in sex-at-age remained between the catch and the survey-derived data. I hypothesize that these differences are owed to differences in the spatiotemporal distribution of the sexes and the spatiotemporal distribution of the survey and fishing effort.

## Introduction

Recent evidence suggests that including sex structure in stock assessment models is important for sexually-dimorphic fishes. For example, if growth rates or natural mortality differ between male and female fish (Wang et al. 2005; Su et al. 2013), if sexspecific migratory behavior and fishing effort are not evenly distributed in space and time (Okamura et al. 2014), or if fishery selectivity is sex-specific (Myers et al. 2014), not including sex-structure in the assessment model can bias biological reference points. In light of these recent findings, sex-structured assessment models are becoming more common in describing the population dynamics of fishes that have sexually-dimorphic characteristics and/or behavior (Clark 2006; Wang et al. 2007; Fenske et al. 2011; Su et al. 2011; McGilliard et al. 2013).

Female Summer Flounder Paralichthys dentatus grow faster (Poole 1961; King et al. 2001) and mature at a larger size (Able and Kaiser 1994; Packer et al. 1999) than males. In addition, the sex ratio of young fish is skewed in favor of males, probably due to a complex interaction between temperature and the biochemistry of sex determination
(Luckenbach et al., 2009). However, as the cohort ages the balance in sex ratio shifts toward females (Smith and Daiber 1977; Bonzek et al. 2009), a pattern that suggests a higher natural mortality rate for the males (Maunder and Wong 2011). Sexually${ }^{1}$ dimorphic life history characteristics of Summer Flounder suggest that a sexstructured assessment model would be appropriate for this species (Jones et al. 2013; NEFSC 2013), yet the most recent assessment applied a single-sex statistical catch-atage model (NEFSC 2013).

Sex-structured assessment models can be difficult to implement because they are data-intensive, requiring either some prior knowledge about the sex composition of the catch or that assumptions be made about the sex-specific selectivity curve and its stability in space and time (Methot and Wetzel 2013; Myers et al. 2014). Sex-specific information is not collected from Summer Flounder landings as part of the annual assessment process because, like other flatfishes, there are no external characteristics that can be used to identify the sex and Summer Flounder cannot be dissected by port agents and still retain their market value. However, on the National Marine Fisheries Service, Northeast Fisheries Science Center (NMFS-NEFSC) bottom trawl survey, conducted twice annually in the spring and fall, sex-specific data are collected using fish dissection (NEFSC 2013).

When fishery-independent survey data are sex-specific and the probability of a given sex at a given length or age are similar between the survey and the catch, it may be possible to apply a sex-at-length or sex-at-age key developed from survey data directly to length or age data from the catch (Clark 2004). However, the sex composition of the
catch is influenced by a combination of the differences in the size-, age-, and sex-specific retention selectivity imposed by the minimum retention size in each fishery (Kendall and Quinn 2013), the size-, age-, and sex-specific vulnerability of fish resulting from their interaction with (Ryer 2008; Somerton et al. 2011), and/or their availability to (Okamura 2014; Sampson 2014), the fishing gears, and the spatial-, temporal-, and sexspecific growth rates of the fish being retained by the different sampling gears (Poole 1961; Planes et al. 1999; Wilderbuer and Turnock, 2009). For instance, different minimum retention size limits are implemented in the recreational and commercial Summer Flounder fisheries (NEFSC 2013) and some limited evidence suggests that the availability/vulnerability of the sexes at a given length to the fishing effort may vary seasonally and between the recreational fishery and the NMFS-NEFSC survey (Morson et al. 2012). These fishery-specific processes can result in differences in the sexcomposition within the catch and between the catch and the survey, so an assumption of equal probabilities of sex-at-length or sex-at-age between catch and survey data may be incorrect.

In this chapter I examine trends in the sex composition of summer flounder landed in the recreational and commercial fisheries. Given these trends, I test the efficacy of applying NMFS-NEFSC survey-derived sex-at-age keys to catch data so that the stock assessment of Summer Flounder might implement a sex-specific model. Finally, I discuss some of the underlying ecological processes that may be driving the observed patterns in selectivity.

## Methods

## Data Collection

Summer Flounder Paralichthys dentatus were collected along the northeast and midAtlantic continental shelf of the USA in 2010 and 2011 from a fishery-independent survey (NFS) and from commercial (CFS) and recreational hook-and-line (RFS) fishery landings (Table 2.1). For all fish, total length, sex, and catch location were recorded. For a subset of the fish sampled, otoliths and/or scales were removed for aging (Table 2.1). Otoliths were removed from carcasses collected from the RFS. Scales were collected for specimens up to 60 cm and otoliths for fish greater than 60 cm from the CFS and the NFS. Otoliths were processed by selecting the right side otolith and centeraligned in a merchandise tag, embedded in black polyester resin, and then crosssectioned through the core with a low-speed sectioning saw. After processing, otoliths were examined using a Leica MZ-6 dissecting microscope with transmitted light at between 8 and 60 times magnification. Annuli from otoliths were identified as narrow opaque bands and were counted along the ventral side of the sulcal groove. Scales were processed by selecting five or six samples from each fish and impressing them on acetate sheets. Impressions were examined using a microfiche reader with 20 and 29mm lenses. Annuli were identified in scales by using established protocols described in Pentilla and Dery (1988).

The National Marine Fisheries Service, Northeast Fisheries Science Center (NMFSNEFSC) provided NFS data for Summer Flounder sampled from the spring and fall bottom trawl survey cruises at predetermined sampling strata at depths ranging from 27 m-365 m from Cape Hatteras, North Carolina, USA to Nova Scotia, Canada. A
detailed description of the bottom trawl survey design can be found in Azarovitz (1981) and NEFSC (1988), but briefly between 350 and 400 stations are sampled each year with a bottom trawl that has a 1.5 cm stretch mesh liner and rollers for sampling hard substrate. Tows are conducted for 30 minutes at each station at a tow speed of 3.5 knots. CFS and RFS data were collected by sampling fish landed at marinas, packing houses, and fishing docks from North Carolina to Massachusetts (Table 2.2).

Sampling of the CFS occurred year-round wherever the fishery was operational. The minimum mesh size in the CFS was a 14 cm diamond mesh or 15 cm square mesh applied throughout the net body, extension, and codend. For each visit to a commercial dock, samples from up to 100 fish were collected haphazardly in each market category (medium, large, and jumbo) available from a given fishing trip. The market category is determined by each dock or packing house, but generally, small fish ranged from 35 cm -45 cm , medium fish from $45 \mathrm{~cm}-55 \mathrm{~cm}$, and jumbo fish were greater than 55 cm . At these sizes, all fish sampled were sexually mature. Summer Flounder cannot be sexed using external characteristics; so, to avoid a reduction in market value, a developed a minimally-invasive technique for determining sex (Figure 2.1). A one-inch incision was made on the pigmented side of the fish in an area halfway between the anterior end of the anal fin and the center of the pectoral fin. Using forceps, the gonads were pulled out through this incision. Orange eggs or empty ovaries and the white testis tissue were used to differentiate female and male fish, respectively. Both male and female fish were easily identified even when sampling did not occur during the spawning season.

RFS ports were sampled once per week during the recreational fishing season (MaySeptember) to collect racks (fileted carcasses) of all Summer Flounder caught that day
on all participating boats. Once Summer Flounder were fileted, the sex organs remained intact and visible on the rack. Boat captains and crew saved fish racks in a bin and those racks were collected upon arrival at a participating dock. In order to increase the number of fish available for collection, freezers were placed at each dock. Bags and waterproof tags were provided to the boat captains and crew and were available near the freezers so that samples could be accurately labeled with the date and location of the catch. Participating boat captains were asked to deposit all fish racks from the day's catch in these tagged bags and place the bags in the freezers. Freezers were emptied by scientists upon arrival at a dock to collect fresh racks. Morson et al. (2012) concluded that the change in Summer Flounder length from freezing was negligible. To ensure a representative sample of harvested Summer Flounder sex, length, and age, all fish caught on a fishing trip were sampled without regard to size. Sex was determined by macroscopic investigation of exposed gonad on filleted fish carcasses. Over ninety-nine percent of fish collected had reproductive organs intact and readily visible to the naked eye.

For all fish sampled from the RFS and CFS, a record was kept of the NMFS-NEFSC statistical area where the fish were caught (Figure 2.2 and Table 2.3). The CFS and RFS operate in different statistical areas so that there was not complete overlap between the two fisheries relative to fishing location. On the NFS, data were collected along stratified sampling strata. To make spatially-comparable comparisons between the NFS and the RFS/CFS, latitude and longitude from each NFS tow were used to assign NFS samples to NMFS-NEFSC statistical areas.

## Data analysis

Age was determined for 23,020 of the 36,745 Summer Flounder sampled from the RFS, CFS, and NFS (Table 2.1). To utilize the full set of data to determine how well sex-at-age collected on the NFS described sex-at-age in the RFS and CFS, it was necessary to generate appropriately partitioned age-length keys (ALKs) and apply those keys to fish not aged empirically. Age-length data were partitioned by sex, region, and data source (SFS/CFS/RFS). Region was defined as north, which included all fish caught or collected north of $40^{\circ}$ latitude; or south, which included all fish caught or collected south of $40^{\circ}$ latitude. To compare ALKs, the age distribution at a given length was modeled using multinomial logistic regression (Gerritsen et al. 2006). The multinomial log-linear models were fit via neural networks using the function multinom() available in the R package nnet (Venables and Ripley 2002). This approach provided a simple and robust method for identifying significant differences between ALKs generated from data partitioned by sex, region, and data source. The best-fit model identified the most appropriate stratification of the data and ALKs were generated based on that stratification. ALKs were then applied to all fish not aged empirically based on a probability of each age given the length of each fish using the ageKey function in the R package FSA (Isermann and Knight 2005; Ogle 2014).

I used logistic regression to evaluate the efficacy of applying a sex-at-age key developed from NFS data to CFS and RFS age data. The probability of landing a female at given age was modeled as the dependent variable and the data source (NFS, CFS, and RFS) was modeled as the independent variable (Wilson and Hardy 2002; Morson et al. 2012). Significant differences between the NFS and both fishery data sources suggested
a sex-at-age key developed from raw NFS data would not be appropriate in describing the sex composition from either fishery. I hypothesized that the size-, age-, and sexspecific retention selectivity imposed by the minimum retention size (MRS) of 35 cm in the CFS and of between 46 cm and 53 cm in the RFS, as well as the differences in growth rates between fish collected from the three data sources, could explain the variability in sex-at-age among the datasets. Therefore, I simulated, independently for the CFS and RFS, the effects of these fishery-specific processes on the fishery-independent survey data and re-analyzed the resultant sex ratio-at-age.

First, to simulate the length-specific retention selectivity of the RFS and CFS, I lefttruncated the NFS data at the MRS of each fishery. This approach provided a NFSderived measure of sex-at-age comparable to the left-truncated sex composition in each fishery-dependent set of data. A 35-cm MRS was implemented coast-wide in the CFS during the sample-collection period, so NFS and CFS data were left-truncated at 35 cm for the CFS comparison. The recreational fishery MRS varied from $46 \mathrm{~cm} \mathrm{-53} \mathrm{~cm}$ depending on the landing year and state. To avoid too many pair-wise comparisons using too little data, I left-truncated the NFS and RFS data at 46 cm and at 53 cm in two separate comparisons that together encompass the full range of potential comparisons.

To simulate the observed growth rates in the CFS and RFS, sex- and region-specific ALKs generated from CFS and RFS data were applied to the left-truncated sex- and region-specific NFS length data. This allowed us to assign age to the left-truncated distribution of lengths in the NFS data as if the NFS fish grew at the same rates as in each respective fishery.

Finally, as a proxy for evaluating the length-specific vulnerability of the sexes to the fisheries, I compared the left-truncated NFS-derived sex ratio-at-length to the measured sex-at-length from the CFS and RFS. Since the NFS utilizes a smaller mesh size than the regulation size in the CFS and the RFS-MRS is set higher than that in the CFS, all lefttruncated NFS data in these comparisons come from fish that are fully recruited to the survey. A logistic regression model was fit to estimate the probability of landing a female at a given length for each pairwise comparison.

All statistical analyses were computed in R v3.0.3 (R Core Team 2014). Model selection followed Akaike Information Criterion (AIC). A model score that was two AIC units lower was accepted as evidence for better fit among competing models (Arnold 2010).

## Results

Summer Flounder Paralichthys dentatus from 27 different NMFS-NEFSC statistical areas were sampled on the NMFS-NEFSC bottom trawl survey (NFS) and from the commercial (CFS) and recreational (RFS) fishery port sampling programs (Table 2.3). The samples collected from the CFS were distributed inshore and offshore across similar statistical areas as those collected on the NFS, but the RFS samples were limited to statistical areas located inshore (Figure 2.2 and Table 2.3).

The probability that a fish of a given length was a given age depended on its sex, region, and data source (NFS/CFS/RFS; Table 2.4). Sex was the most important effect in the model, followed by data source, and region.

The sex ratio-at-age in the NFS was significantly different than the CFS ( $\mathrm{p}<0.0001$ ) and the RFS ( $\mathrm{p}<0.0001$ ) when comparisons included all NFS data (Figure 2.3a and Figure 2.4a). A higher proportion of the catch-at-age is female in both fisheries (Figure 2.3a, 4a, and 5a). However, fish landed under a minimum retention size (MRS) had a higher mean length-at-age than fish collected by the NFS. The higher the MRS imposed on the landed fish, the higher the mean length-at-age (Figure 2.6). This effect was apparent for all ages containing the MRS within their length frequency. When NFS data were left-truncated to simulate the MRS in the CFS and RFS to account for this, the sex ratio-at-age pattern more closely approximated the pattern in the data collected directly from the fisheries (Figure 2.3b, 4b, and 5b). The proportions-at-age in the survey-derived data shifted dramatically toward female fish at younger ages (Figures 3b, 4b, and 5b). Applying, separately, sex- and region-specific age-length keys (ALK) generated from CFS and RFS data to left-truncated sex- and region-specific NFS length data resulted in a sex ratio-at-age pattern that further resembled the pattern of the data collected directly from the CFS and RFS (Figures 3c, 4c, and 5c). The shift in sex ratio-at-age suggests that the differences in growth between the fish landed in the fisheries and the fish collected on the survey is sex-specific. Females landed in both fisheries had grown faster than female fish collected from the NFS, and male fish landed in both fisheries had grown slower than male fish collected from the NFS. Thus, applying fishery-specific ALKs resulted in a shift in the sex ratio of younger fish further toward female and a shift in the sex ratio of older fish toward male (Figures 3b and 3c; Figure $2.4 b, 4 c, 5 b$, and $5 c$ ). For example, in the CFS comparison (Figure $2.3 b$ and $3 c$ ), one-year-old survey-derived sex-at-age increased from approximately 40\% female (Figure
2.3b) to approximately $65 \%$ female (Figure 2.3 c ) whereas six-year-old survey-derived sex-at-age decreased from approximately 75\% female (Figure 2.3b) to approximately 50\% female (Figure 2.3c). Even so, statistically significant differences remained between the CFS $(\mathrm{p}<0.0001)$ and RFS $(\mathrm{p}<0.0001)$ data and the NFS-derived data that were manipulated to simulate the fishery-specific MRS and growth processes within catches from the fisheries.

Given that accounting for the length-specific retention selectivity and the differences in growth rates between the fish landed in the fisheries and those collected on the survey did not remove all of the variability in sex-at-age between the survey and catch data, I attempted to estimate the difference in availability of the sexes to the survey and the fisheries. The length-specific vulnerability of the sexes to the gear, modeled as the probability of a fish being female at a given length, was similar, but statistically different ( $\mathrm{p}=0.0037$ ) for the left-truncated NFS data and CFS data comparison (Figure 2.7a). The probability of landing a female at a given length is slightly higher in the CFS than in the left-truncated NFS (Figure 2.7a). In addition, the proportion female-at-length increased in both the CFS and the left-truncated NFS at similar rates until each reached the smallest MRS in the recreational fishery ( $\sim 45 \mathrm{~cm}$ ). At this point the female proportion-at-length remained unchanged or increased only slightly from one centimeterincrement to the next until reaching approximately the $50-\mathrm{cm}$ length increment when the rate for both datasets returned to a logistic form (Figure 2.7). The probability of landing a female at a given length was much higher in the RFS than for the lefttruncated NFS data ( $\mathrm{p}<0.0001$; Figure 2.7b). In fact when a fish is landed in the recreational fishery at the smallest MRS ( 46 cm ), there is an $80 \%$ chance that the fish
will be female and this probability rapidly approaches $100 \%$ with increasing size (Figure 2.7b).

## Discussion

To determine whether the sex composition of Summer Flounder Paralichthys dentatus from a fishery-independent trawl survey could be used to estimate the sex composition of the Summer Flounder catch without collecting sex-specific data directly from the fisheries, I simulated fishery-specific processes on the survey data, including the minimum retention size (MRS) in each fishery and the growth rates of fish landed in both fisheries, and compared those survey-derived estimates to the actual sex composition in the catch.

Fish landed in the fisheries had grown at different rates than those collected from the survey, and at different rates from one another, suggesting that each gear is sampling a different part of the population. Furthermore, this effect was sex-specific; female fish landed in the fisheries had grown faster than those collected on the survey, while male fish collected on the survey had grown faster than male fish landed in the fisheries. This suggests that fisheries are selectively targeting faster-growing females and slowergrowing males. Stari et al. (2010) offered that sampling location or gear selectivity could explain differences in age-length keys (ALK) for North Sea Haddock Melanogrammus aeglefinus collected from different gears. I propose the differences in ALKs found here could result from the spatial sex-specific segregation of Summer Flounder previously suggested (Morson et al. 2012), the influence of region on growth rates previously identified (Kraus and Musick 2001) and further confirmed with this
work, and the spatial and temporal distribution of fishing effort; though the influence of age-specific gear selectivity cannot be ruled out. I discuss spatiotemporal distribution of fish and fishing effort in more detail below.

The MRS in each fishery, operating on the sex-specific growth rates (Poole 1961; King et al. 2001), forces a higher fraction female-at-age in the catch than would be expected in a non-culled, fishery-independent sample. I was able to largely correct for this by left-truncating the survey data to mimic the MRS in each fishery and this transformation of the survey data recovered a sex-at-age pattern more closely resembling the one measured directly from the catch. Not surprisingly, when I applied ALKs developed from fishery-dependent data to left-truncated survey data, the result was a sex-at-age pattern in the survey-derived data that even more closely resembled the one measured directly from the catch.

This step-wise approach to simulating fishery-specific effects on fishery-independent data recovered the pattern of sex-at-age in the fisheries and it may therefore have application to other fisheries where biological data from the catch are limited and a desire exists to estimate sex-specific parameters in an assessment. However, these manipulations did not recover the exact proportions-at-age measured directly from the catch. I hypothesize that this is due to some difference in the spatiotemporal distributions in stock structure and fishing effort, such that the availability of the sexes to the survey and the fishing fleets was not equivalent. As evidence, I show that sex-atlength, used as a proxy for the availability of fish on the bottom to the fishing gears, varied slightly between the commercial fishery and the survey, and dramatically between the recreational fishery and the survey. The recreational fishery occurs
inshore from late spring to early fall, the commercial fishery occurs year-round inshore and offshore, and the NMFS-NEFSC survey occurs offshore once in March and once in September. As no overlap exists in space or time between the recreational fishery and the survey, the expectation is that if spatial and temporal structure to the sex composition exists in the population, then differences in the spatiotemporal availability of the sexes to the survey and recreational fishery will be reflected in the catch. This was observed, manifested in the large difference in sex ratio-at-length between the recreational catch and survey data.

As the commercial fishery overlaps both in time and space with the survey, the expectation is that sex-specific availability of fish to the commercial fishery and survey will be similar. Such a finding also was observed, manifested in the fairly close match in sex-at-length between the commercial catch and the survey data. This is the first evidence, to my knowledge, of sex-specific segregation of Summer Flounder in space and time.

Spatial and temporal segregation of fishes by sex is a common phenomenon and occurs in other sexually-dimorphic flatfishes (Morgan and Trippel 1996; Swain 1997; Swain and Morin 1997; Gorchinksy 1998; Richards et al. 2008; Gerritsen 2010; Sahin and Gunes 2010; Loher 2012; Loher and Hobden 2012). Explanations for sex-specific segregation include temperature (Swain 1997) and/or depth (Swain and Morin 1997) selection, sex-specific life history strategies that promote sexually-dimorphic characteristics (Gerritsen et al. 2010), and sex-specific foraging intensity and the spatial and temporal distribution of the forage base (Swain and Morin 1996). Any of these postulations could be advanced to explain the sex-specific distribution of Summer

Flounder in space and time. The importance, however, of estimating and understanding the sex-specific spatial and temporal distribution of Summer Flounder goes beyond these interesting ecological implications. Okamura et. al (2014) showed that biological reference points are highly sensitive to assumptions about effort allocation in space and time when seasonal migrations of Offshore Lingcod Ophiodon elongatus are sex-specific. A similar analysis for Summer Flounder is beyond the scope of this paper, but, minimally, the application of sex-aggregated ALKs to landings data should result in highly biased estimates of the catch-at-age given that female Summer Flounder grow much faster than males and that the recreational fishery lands female fish almost exclusively. Accounting for the complex spatiotemporal dynamics in the distribution of sex composition and fishing effort will undoubtedly complicate future stock assessments of Summer Flounder.

Given the importance of having an accurate estimate of the sex composition of the catch and the failure of my method to recover an exact match by applying sex-specific processes to survey data, because of what appear to be complex, time- and spacevarying sex-specific fishery selectivities, it makes sense to explore other options for collecting sex-composition data directly from fishery landings. In the recreational fishery, this would be easy to incorporate into any port-sampling program that already collects length-frequency data from the catch. I show that even when Summer Flounder are fileted at sea, the gonad remains intact and readily visible to the naked eye. An observer measuring the length of a fish can determine the sex macroscopically with no additional time or cost. In the commercial fishery, collecting sex-specific information may prove more difficult because fish are not fileted at sea to expose the gonads and
fish are typically sold whole to market, which precludes full dissection of fish to determine the sex. I introduced a method for determining sex using a small incision in the gut cavity. Although this may still be too invasive for some fishing ports or fish sellers or buyers, I encountered no resistance on the part of the many commercial docks that I sampled. The Pacific Halibut Commission used microsatellite-based genetic techniques to measure the sex composition of the catch (Loher et al. 2012). This reduced the influence of spatial and temporal variability in survey and fishing effort when deriving sex composition of the catch using survey data because it provided a direct measure of the sex composition of the catch. A cost-benefit analysis of purchasing fish and dissecting them vs. the development of microsatellite techniques may shed some light on the most cost- and time-effective way forward for determining the sex composition of the commercial catch. No matter how the data are obtained though, if a program is instituted to collect sex composition data directly from the catch it would need to mirror the spatial and temporal distribution of the fishing effort since it appears male and female fish disaggregate in time and space.

The trends observed in the sex, length, and age of the catch may interact with fishing regulations and induce changes in population structure. The mean length-at-age of individuals in the Summer Flounder population is declining over time with no accompanying change in the length-weight relationship or condition index (NEFSC 2013) that one might expect from density-dependent influences on individual fish growth (Rose et al. 2001). Given the dynamic nature of climate change in the northwestern Atlantic Ocean (Scavia et al. 2002), and its influence on the fish stocks of this region (Rothschild and Jiao 2012; Sparrevohn et al. 2013), a simple
environmentally-determined change in growth rate in the Summer Flounder stock cannot be discounted. Nonetheless, I show a trend of increasing mean length-at-age with increasing MRS in the fishery landings. The MRS being implemented is selecting, at a given age, the fastest growing fish while leaving behind the slower growing fish to reproduce another year. A growing literature addresses the potential evolutionary effects of size-selective fishing (e.g., Conover and Munch 2002; Law 2007). Varying opinions exist concerning the time-scale and fishing pressure needed to induce evolutionary change as opposed to simple phenotypic change in a fish population (e.g., Anderson and Brander 2009; Enberg et al. 2009); however the possibility cannot be ruled out that the size-selective fishing pressure on the Summer Flounder population could be causing a shift in the genetic structure of the stock favoring slower growth and resulting in a decrease in the mean length-at-age over time.

Additionally, the proportion female-at-age in the Summer Flounder population is in decline (NEFSC 2013). If the MRS is selecting the faster growing fish in the population at a given age, those fish have to be disproportionately female because of the difference in growth rates between the sexes. My sampling found that the sex-at-age relationships from the recreational catch, and to a lesser extent from the commercial catch, are skewed toward female fish in comparison to the sex-at-age expected from survey data. Simulation of the effects of the MRS shows that this result accrues in part from the implementation of the MRS in each fishery. For example, the proportion of fish that are female increases with increasing length in the survey, as well as in the fisheries. But, interestingly, in both the commercial fishery data and survey data, at around the 5 cm increment that encompasses the range of MRS implemented in the recreational fishery
( $45 \mathrm{~cm}-50 \mathrm{~cm}$ ), the sex ratio remains relatively unchanged, when the expectation is that it should increase as it does at each length increment before and after the 45 cm 50 cm increment. This suggests the MRS implemented in the recreational fishery may have altered the population sex ratio within this range of sizes. One potential explanation, therefore, for the trend of decreasing proportion female-at-age in the population over the last decade (NEFSC 2013) could be the increases in MRS used to rebuild the stock over that same time period and the associated increases in femalespecific fishing mortality. Sex-specific differences in fishery selectivity happen in other sexually-dimorphic flatfishes regulated by an MRS (Briggs 1965; Lohre and Hobden 2012). In fact, the sex composition estimates for commercial Pacific halibut Hippoglossus stenolepis landings indicate that males may never reach mean sizes associated with full fishery selectivity-at-age (Clark and Hare 2006).

If current management strategies are driving changes in the structure of the Summer Flounder stock, it is important for managers to consider the potential adverse effects of those changes. Size-selective fishing can alter the sex ratio of a population (Kendall and Quinn 2013) and influence the reproductive potential of the stock (Rowe and Hutchings 2003; Claereboudt et al. 2004; Rijnsdorp et al. 2010) and genetic traits selected against by implementing a MRS as a management strategy may be difficult to recover (Enberg et al. 2009).

## Acknowledgements

I acknowledge all of the participating fishing docks, for-hire vessel captains and mates that participated in this project. I thank the NMFS-NEFSC Ageing Center, especially Eric Robillard, Integrated Statistics, Mark Luckenbach, Erin Reilly, Patti McGrath, and Roger Mann of the Virginia Institute of Marine Science, Joe Cimino and Joe Grist of the Virginia Marine Resources Commission, Chris Batsavage of the North Carolina Division of Marine Fisheries, Jacqueline Wilson, Joe Costanzo, and Emerson G. Hasbourck of the Cornell Cooperative Marine Program, and the many technicians at the Haskin Shellfish Research Laboratory, Rutgers University. Special thanks to the Partnership for Marine

Fisheries Science for providing two years of funding for this project through NOAA NMFS Award NA10NMF4720017 and NA10NMF4720402 and to the New Jersey Sea Grant College Program (NJSG-15-892).

## Literature Cited - Chapter 1

Able, K. W., and S. C. Kaiser. 1994. Synthesis of summer flounder habitat parameters. National Oceanic and Atmospheric Administration Coastal Ocean Office, Decision Analysis Series 1, Silver Spring, Maryland.

Anderson, K. H., and K. Brander. 2009. Expected rate of fisheries-induced evolution is slow. Proceedings of the National Academy of Sciences of the United States of America 106: 11657-11660.

Arnold, T. W. 2010. Uninformative parameters and model selection using Akaike's information criterion. The Journal of Wildlife Management 74: 1175-1178.

Azarovitz, T. R. 1981. A brief historical review of the Woods Hole laboratory trawl survey time series. Canadian Special Publication of Fisheries and Aquatic Sciences 58: 62-67.

Bonzek, C. F., J. Gartland, J. D. Lange Jr., and R. J. Latour. 2009. Northeast Area Monitoring and Assessment Program (NEAMAP) Near Shore Trawl Survey Final Report.

Atlantic States Marine Fisheries Commission, Washington, D.C.
Briggs, P. T. 1965. The sport fisheries for winter flounder in several bays of Long Island. New York Fish and Game Journal 12: 48-70.

Claereboudt, M. R., J. L. Mcllwain, H. S. Al-Oufi, and A. A. Ambu-Ali. 2004. Relationships between fishing gear, size frequency and reproductive patterns for the kingfish (Scomberomorus commerson, Lacepede) fishery in the Gulf of Oman. Pages 56-67 in A. I. L. Payne, C. M. O’Brien, and S. I. Rogers, editors. Management of shared fish stocks. Blackwell Publishing, Oxford.

Clark, W. G. 2004. A method of estimating the sex composition of the commercial landings from setline survey data. In Report of Assessment and Research Activities 2003. Edited by L. Sadorus. International Pacific Halibut Commission, Seattle, WA. pp. 111-162.

Clark, W. G. 2006. Assessment and management of Pacific halibut: data, methods, and policy. International Pacific Halibut Commission Scientific Report 83. International Pacific Halibut Commission, Seattle, WA.

Conover, D. O., and S. B. Munch. 2002. Sustaining fishery yields over evolutionary time scales. Science 297: 94-96.

Enberg, K., C. Jorgensen, E. S. Dunlop, M. Heino, and U. Dieckmann. 2009. Implications of fisheries-induced evolution for stock rebuilding and recovery. Evolutionary Applications 2: 394-414.

Fenske, K. H., M. J. Wilberg, D. H. Secor, and M. C. Fabrizio. 2011. An age- and sexstructured assessment model for American eels (Anguilla rostrata) in the Potomac River, MD. Canadian Journal of Fisheries and Aquatic Science 68: 1024-1037.

Gerritsen, H. D., D. McGrath, and C. Lordan. 2006. A simple method for comparing agelength keys reveals significant regional differences within a single stock of haddock (Melanogrammus aeglefinus). ICES Journal of Marine Science 63: 1096-1100.

Gerritsen, H. D., D. McGrath, C. Lordan, and X. Harlay. 2010. Differences in habitat selection of male and female megrim (Lepidorhombus whiffiagonis, Walbaum) to the west of Ireland. A result of differences in life-history strategies between the sexes? Journal of Sea Research 64: 487-493.

Gorchinsky, K. V. 1998. Spatial and temporal variations in Greenland halibut (Reinhardtius hippoglossoides) sex ratio-at-age in NAFO Divisions 0B, 2GH, 3K and 3LM. North Atlantic Fisheries Organization Scientific Council Studies 31: 73-78.

Isermann, D. A., and C. T. Knight. 2005. A computer program for age length keys incorporating age assignment to individual fish. North American Journal of Fisheries Management 25: 1153-1160.

Jones, C. M., R. Cook, J. Simmonds, and H. Sparholt. 2013. Summary Report of the 57th Northeast Regional Stock Assessment Review Committee (SARC 57). Northeast Fisheries Science Center, Woods Hole, MA. 47pp.

Kendall, N. W., and T. P. Quinn. 2013. Size-selective fishing affects sex ratios and the opportunity for sexual selection in Alaskan sockeye salmon Oncorhynchus nerka. Oikos 122: 411-420.

King, N. J., G. C. Nardi, and C. J. Jones. 2001. Sex-linked growth divergence of summer flounder from a commercial farm: are males worth the effort? Journal of Applied Aquaculture 11:77-78.

Kraus, R. T., and J. A. Musick. 2001. A brief interpretation of summer flounder, Paralichthys dentatus, movements and stock structure with tagging data on juveniles. Marine Fisheries Review 63(3): 1-6.

Law, R. 2007. Fisheries-induced evolution: present status and future directions. Marine Ecology Progress Series 335: 271-277.

Loher, T., and J. Hobden. 2012. Length and sex effects on the spatial structure of catches of Pacific halibut (Hippoglossus) on longline gear. Fishery Bulletin 110: 46-51.

Loher, T., M. Woods, I. Jimenez-Hidalgo, and L. Hauser. 2016. Variance in age-specific sex composition of Pacific halibut catches, and comparison of statistical and genetic methods for reconstructing sex ratios. Journal of Sea Research 107: 90-99.

Luckenbach, J. A., R. J. Borski, H. V. Daniels, and J. Godwin. 2009. Sex determination in flatfishes: mechanisms and environmental influences. Seminars in Cell \& Developmental Biology 20: 256-263.

Maunder, M. N., and R. A. Wong. 2011. Approaches for estimating natural mortality: application to summer flounder (Paralichthys dentatus) in the U.S. mid-Atlantic. Fisheries Research 111:92-99.

McGilliard, C. R., W. Palsson, W. Stockhausen, and J. Ianelli. 2013. Assessment of the deepwater flatfish stock in the Gulf of Alaska. In: Stock Assessment and Fishery Evaluation Report for the Groundfish Resources of the Gulf of Alaska. pp. 403-536.

Methot, R. D., Jr., and C. R. Wetzel. 2013. Stock synthesis: a biological and statistical framework for fish stock assessment and fishery management. Fisheries Research 142: 86-99.

Morgan, M. J., and E. A. Trippel. 1996. Skewed sex ratios in spawning shoals of Atlantic cod (Gadus morhua). ICES Journal of Marine Science 53: 820-826.

Morson, J. M., E. A. Bochenek, E. N. Powell, and J. E. Gius. 2012. Sex-at-length of summer flounder landed in the New Jersey recreational party boat fishery. North American Journal of Fisheries Management 32: 1201-1210.

Myers, R. A., M. W. Smith, J. M. Hoenig, N. Kmeicik, M. A. Luehring, M. T. Drake, P. J. Schmalz, and G. G. Sass. 2014. Size and sex-specific capture and harvest selectivity of walleyes from tagging studies. Transactions of the American Fisheries Society 143: 438-450.

Northeast Fisheries Science Center (NEFSC). 1988. An evaluation of the bottom trawl survey program of the Northeast Fisheries Science Center. National Oceanic and Atmospheric Administration Technical Memorandum. NMFS-F/NEC-52, 81 pp.

Northeast Fisheries Science Center (NEFSC). 1997. Report to the 24th Northeast Regional Stock Assessment Workshop (24th SAW): Stock Assessment Review Committee (SARC) Consensus Summary of Assessments. Northeast Fisheries Science Center Reference Document 97-12, 291 pp.

Northeast Fisheries Science Center (NEFSC). 2013. 57th Northeast Regional Stock Assessment Workshop (57th SAW) Assessment Report. U.S. Department of Commerce, Northeast Fisheries Science Center Reference Document 13-16. 967pp.

Ogle, D. H. 2014. FSA: Fisheries Stock Analysis. R package version 0.4.16.
Okamura, H., M. K. McAllister, M. Ichinokawa, L. Yamanaka, and K. Holt. 2014. Evaluation of the sensitivity of biological reference points to the spatio-temporal distribution of fishing effort when seasonal migrations are sex-specific. Fisheries Research 158: 116-123.

Packer, D. B., S. J. Griesbach, P. L. Berrien, C. A. Zetlin, D. L. Johnson, and W. W. Morse. 1999. Essential fish habitat source document: summer flounder, Paralichthys dentatus, life history and habitat characteristics. National Oceanic and Atmospheric Administration Technical Memorandum NMFS-NE-151. 98pp.

Pentilla, J. and L. M. Dery. 1988. Age determination methods for northwest Atlantic species. National Oceanic and Atmospheric Administration Technical Report. NMFS Volume 72, 135 pp.

Planes, S., E. Macpherson, F. Biagi, A. Garcia-Rubies, J. Harmelin, M. Harmelin-Vivien, J-Y. Jouvenel, L. Tunesi, L. Vigliola, and R. Galzin. 1999. Spatio-temporal variability in growth of juvenile sparid fishes from the Mediterranean littoral zone. Journal of the Marine Biological Association of the United Kingdom 79: 137-143.

Poole, J. C. 1961. Age and growth of the fluke in Great South Bay and their significance to the sport fishery. New York Fish and Game Journal 8: 1-18.

R Core Team. 2014. R: A Language and Environment for Statistical Computing: R Foundation for Statistical Computing, Vienna, Austria.

Rijnsdorp, A. D., D. J. G. van Damme, and P. R. Witthames. 2010. Implications of fisheries-induced changes in stock structure and reproductive potential for stock recovery of a sex-dimorphic species, North Sea plaice. ICES Journal of Marine Science 67: 1931-1938.

Rose, K. A., J. H. Cowan Jr., K. O. Winemiller, R. A. Myers, and R. Hilborn. 2001. Compensatory density dependence in fish populations: importance, controversy, understanding and prognosis. Fish and Fisheries 2: 293-327.

Rothschild, B. J., and Y. Jiao. 2012. Characterizing variation in northwest Atlantic fishstock abundance. ICES Journal of Marine Science 69: 913-922

Rowe, S., and J. A. Hutchings. 2003. Mating systems and the conservation of commercially exploited marine fish. Trends in Ecology and Evolution 18: 567-572.

Ryer, C. H. 2008. A review of flatfish behavior relative to trawls. Fisheries Research 90: 138-146.

Sahin, T., and E. Gunes. 2010. Seasonal variation in length, weight, and sex distribution of flounder (Platichtys flesus luscus) in the South-Eastern Black Sea. Journal of Fisheries Sciences 4: 238-245.

Sampson, D. B. 2014. Fishery selection and its relevance to stock assessment and fishery management. Fisheries Research 158: 5-14.

Scavia, D., J. C. Field, D. F. Boesch, R. W. Buddemeier, V. Burkett, D. R. Cayan, M. Fogarty, M. A. Harwell, R. W. Howarth, C. Mason, D. J. Reed, T. C. Royer, A. H. Sallenger, and J. G. Titus. 2002. Climate change impacts on U.S. coastal and marine ecosystems. Estuaries 25: 149-164.

Smith, R.W., and F. C. Daiber. 1977. Biology of summer flounder, Paralichthys dentatus, in Delaware Bay. U. S. National Marine Fisheries Service Fishery Bulletin 75:823-830.

Somerton, D. A., K. Williams, P. G. von Szalay, and C. S. Rose. 2011. Using acoustics to estimate the fish-length selectivity of trawl mesh. ICES Journal of Marine Science 68: 1558-1565.

Stari. T., K. F. Preedy, E. McKenzie, W. S. C. Gurney, M. R. Heath, P. A. Kunzlik, and D. C. Speirs. 2010. Smooth age length keys: observations and implications for data collection on North Sea haddock. Fisheries Research 105: 2-12.

Swain, D. P. 1997. Sex-specific temperature distribution of American plaice (Hippoglossoides platessoides) and its relation to age and abundance. Canadian Journal of Fisheries and Aquatic Sciences 54: 1077-1087.

Swain, D. P., and R. Morin. 1996. Relationship between geographic distribution and abundance of the American plaice (Hippoglossoides platessoides) in the southern Gulf of St Lawrence. Canadian Journal of Fisheries and Aquatic Sciences 53: 106-119.

Swain, D. P., and R. Morin. 1997. Effects of age, sex and abundance on the bathymetric pattern of American plaice in the southern Gulf of St Lawrence. Journal of Fish Biology 50: 181-200.

Su, N-J., C-L. Sun, A. E. Punt, S-Z. Yeh, and G. DiNardo. 2011. Evaluation of a spatially sex-specific assessment method incorporating a habitat preference model for blue marlin (Makaira nigricans) in the Pacific Ocean. Fisheries Oceanography 20:415-433.

Su, N-J., C-L. Sun, A. E. Punt, S-Z. Yeh, W-C. Chaing, Y-J. Chang, and H-Y. Chang. 2013. Effects of sexual dimorphism on population parameters and exploitation ratios of blue marlin (Makaira nigricans) in the northwest Pacific Ocean. Aquatic Living Resources 26: 19-24.

Venables, W. N., and B. D. Ripley. 2002. Modern applied statistics with S, 4th Edition. Springer, New York.

Wang, S-P., C-L. Sun, A. E. Punt, and S-Z. Yeh. 2005. Evaluation of a sex-specific agestructured assessment method for the swordfish, Xiphias gladius, in the North Pacific Ocean. Fisheries Research 73: 79-97.

Wang, S-P., C-L. Sun, A. E. Punt, and S-Z. Yeh. 2007. Application of a sex-specific agestructured assessment method for the swordfish, Xiphias gladius, in the North Pacific Ocean. Fisheries Research 84: 282-300.

Wilderbuer, T. K., and B. J. Turnock. 2009. Sex-specific natural mortality of arrowtooth flounder in Alaska: implications of a skewed sex ratio on exploitation and management. North American Journal of Fisheries Management 29: 306-322

Wilson, K., and I. Hardy. 2002. Statistical analysis of sex ratios: an introduction. Pages 48-92 in Ian C. W., editor. Sex Ratios: Concepts and Research Methods. Cambridge University Press, Cambridge, UK.

## TABLES - CHAPTER 1

Table 1.1. Total number of fish sampled for sex and length and the subset of those fish that were also aged by examining annuli in otoliths and scales (see Methods).

| Survey | Sex and length | Age |
| :--- | :--- | :--- |
|  |  |  |
| NMFS-NEFSC bottom trawl survey |  |  |
| (NFS) | 4,828 | 3,935 |
| Commercial fishery survey (CFS) | 18,685 | 11,809 |
| Recreational fishery survey (RFS) | 13,232 | 7,276 |
|  |  |  |
| Total | 36,745 | 23,020 |

Table 1.2. Locations where samples were collected from the recreational and commercial Summer Flounder fisheries.

| Recreational Fishery |  | Commercial Fishery |  |
| :---: | :---: | :---: | :---: |
| State | Port | State | Port |
| VA | Virginia Beach | NC | Wanchese |
|  | Hampton | VA | Hampton |
|  | Buckroe |  | Newport News |
|  | James River | NJ | Cape May |
|  | Capeville |  | Barnegat Light |
|  | Wachapreague |  |  |
|  | Yorktown |  |  |
|  | Cape Charles |  |  |
|  | Mathews |  | Point Pleasant |
| MD | Ocean City | NY | Point Lookout |
| DE | Lewes |  | Hampton Bays |
| NJ | Cape May |  | Mattituck |
|  | Fortescue |  | East Hampton |
|  | Barnegat Light |  | Montauk |
|  | Point Pleasant | CT | Stonington |
|  | Atlantic Highlands | RI | Point Judith |
| NY | Freeport | MA | Westport |
|  | Huntington |  | New Bedford |
|  | Captree |  | Hyannis |
|  | Port Jefferson |  |  |
|  | Moriches |  |  |
|  | Riverhead |  |  |
|  | Hampton Bays |  |  |
|  | Mattituck |  |  |
|  | Greenport |  |  |
|  | East Hampton |  |  |
|  | Montauk |  |  |
| CT | Niantic |  |  |
| RI | Point Judith |  |  |
| MA | New Bedford |  |  |
|  | Hyannis |  |  |

Table 1.3. Total number of fish sampled, by NMFS-NEFSC statistical area (see Figure 1.2), from the NMFS-NEFSC bottom trawl survey, the commercial fishery, and the recreational fishery.

| NMFS-NEFSC <br> Statistical <br> area | NMFS-NEFSC <br> bottom trawl <br> survey (NFS) | Commercial <br> fishery (CFS) | Recreational <br> fishery (RFS) |
| :--- | :--- | :--- | :--- |
| 201 |  |  | 930 |
| 393 | 12 |  | 419 |
| 514 | 3 | 40 |  |
| 521 | 14 | 120 |  |
| 522 | 71 | 401 |  |
| 525 | 230 | 157 |  |
| 526 | 741 | 1,596 | 1,278 |
| 537 | 5 | 454 | 1,065 |
| 538 | 270 | 2,558 |  |
| 539 | 27 | 39 | 1,078 |
| 561 | 7 | 2,028 | 3,454 |
| 562 | 658 | 842 | 839 |
| 611 | 112 | 3,985 | 45 |
| 612 | 262 | 687 | 40 |
| 613 | 441 | 3,343 | 4,080 |
| 614 | 421 | 432 |  |
| 615 | 289 | 508 |  |
| 616 | 18 |  | 176 |
| 621 | 178 | 488 |  |
| 622 | 493 | 141 |  |
| 623 | 114 |  |  |
| 625 | 293 |  |  |
| 626 | 44 |  |  |
| 631 |  |  |  |
| 632 |  |  |  |
| 635 |  |  |  |
|  |  |  |  |

Table 1.4. Akaike information criterion (AIC) for eight potential models describing the probability of a given age provided the length (null model) and the sex, region (north of $40^{\circ}$ latitude or south of $40^{\circ}$ latitude), and data source (data collected from the recreational fishery, the commercial fishery, or a fishery-independent trawl survey).

| Model | DF | AIC | Delta AIC | Residual <br> deviance |
| :--- | :--- | :--- | :--- | :--- |
| Sex, region, data source | 65 | 59836 | 0 |  |
| Sex, region | 56 | 60226 | 390 | 69668 |
| Sex, data source | 70 | 60268 | 432 | 60114 |
| Sex | 42 | 60605 | 769 | 60521 |
| Region, data source | 70 | 65590 | 5754 | 65450 |
| Data source | 56 | 65949 | 6113 | 65837 |
| Region | 42 | 66242 | 6406 | 66158 |
|  | 28 | 66550 | 6714 | 66494 |

## FIGURES - CHAPTER 1



Figure 1.1. Photographs showing a technique for determining the sex ( $A=$ Female; $B=$ Male) of a Summer Flounder without dissecting the entire fish. A small incision is made along the gut cavity and the gonads are pulled through the incision with a pair of forceps.


Figure 1.2. Map showing National Marine Fisheries Service, Northeast Fisheries Science Center statistical area units along the northeast USA and the western midAtlantic continental shelf (NEFSC 1997). The shapes around the statistical area number indicate the data source as NMFS-NEFSC bottom trawl survey (NFS), commercial fishery (CFS), or recreational fishery (RFS). Rectangles represent RFS only, diamonds represent CFS only, pentagons represent NFS only, triangles represent RFS and CFS, hexagons represent NFS and CFS, and circles represent RFS, CFS, and NFS.


Figure 1.3. Proportion female Summer Flounder at-age in the commercial fishery (blue dots) and the National Marine Fisheries Service, Northeast Fisheries Science Center (NMFS-NEFSC) bottom trawl survey (green dots). Lines represent the probability of landing a female estimated by logistic regression (see text). Commercial fishery data are compared to (A) raw NMFS-NEFSC bottom trawl survey data, (B) NMFS-NEFSC bottom trawl survey data that were left-truncated at 35 cm to simulate the minimum retention size in the commercial fishery, and (C) NMFS-NEFSC bottom trawl survey data that were left-truncated at 35 cm and assigned age based on age-length keys generated from commercial fishery data.


Figure 1.4. Proportion female Summer Flounder at-age in the recreational fishery (orange dots) and the National Marine Fisheries Service, Northeast Fisheries Science Center (NMFS-NEFSC) bottom trawl survey (green dots). Lines represent the probability of landing a female estimated by logistic regression (see text). Recreational fishery data are compared to (A) raw NMFS-NEFSC bottom trawl survey data, (B) NMFS-NEFSC bottom trawl survey data that were left-truncated at 46 cm to simulate the smallest minimum retention size in the recreational fishery, and (C) NMFS-NEFSC bottom trawl survey data that were left-truncated at 46 cm and assigned age based on age-length keys generated from recreational fishery data.


Figure 1.5. Proportion female Summer Flounder at-age in the recreational fishery (orange dots) and the National Marine Fisheries Service, Northeast Fisheries Science Center (NMFS-NEFSC) bottom trawl survey (red dots). Lines represent the probability of landing a female estimated by logistic regression (see text). Recreational fishery data are compared to (A) raw NMFS-NEFSC bottom trawl survey data, (B) NMFS-NEFSC bottom trawl survey data that were left-truncated at 53 cm to simulate the largest minimum retention size in the recreational fishery, and (C) NMFS-NEFSC bottom trawl survey data that were left-truncated at 53 cm and assigned age based on age-length keys generated from recreational fishery data.


Figure 1.6. Box and whisker plots of the mean length-at-age of Summer Flounder under different minimum retention size regulations. Plots are organized by region (north and south) and sex (male and female). Black horizontal lines are the mean, boxes are the interquartile ranges, black vertical lines are the $5^{\text {th }}$ and $95^{\text {th }}$ percentiles, and dots are outliers.


Figure 1.7. Proportion female Summer Flounder at-length in the commercial fishery (blue dots), the recreational fishery (orange dots), and the National Marine Fisheries Service, Northeast Fisheries Science Center (NMFS-NEFSC) bottom trawl survey (green dots). Lines represent the probability of landing a female estimated by logistic regression (see text). NMFS-NEFSC bottom trawl survey data are truncated according to the minimum retention size in the (A) commercial fishery ( 35 cm ) and (B) recreational fishery ( 46 cm ).

## CHAPTER 2

# EVALUATING THE POTENTIAL FOR A SEX-BALANCED HARVEST APPROACH IN THE RECREATIONAL SUMMER FLOUNDER PARALICHTHYS DENTATUS FISHERY** 


#### Abstract

Summer Flounder (Paralichthys dentatus) support important recreational and commercial fisheries along the northeast and mid-Atlantic coasts of the USA. In the recreational sector, management efforts to constrain harvest below the maximum allowable catch have typically involved increasing the minimum landing size; however, females grow faster than males. Thus, reliance on increased minimum size limits as a management strategy has resulted in approximately $90 \%$ of the recent recreational landings being large, female fish. I evaluated the potential for slot limits to produce a sex-balanced harvest in the recreational Summer Flounder fishery. To estimate the size- and sex-specific vulnerability I sampled the landed and discarded fish ( $\mathrm{n}=3,290$ ) caught by recreational anglers on select party boats from New Jersey to Rhode Island during the 2016 recreational fishing season. I then examined the performance of a wide array of slot limits to estimate which would have promoted a more sex-balanced harvest while maintaining a fixed fishing mortality given the observed catch composition. I demonstrate that slot limits applied to the recreational Summer Flounder fishery have the potential to simultaneously meet multiple management objectives, including conservation of female biomass while maintaining a fixed fishing


[^1]mortality; however, no single slot limit performed best at all sampling locations. Results should therefore be viewed as optimal given the observed catch composition for the year, fishing mode, and locations that were observed and further evaluation of interannual, spatial, and fishing mode variability is warranted.

## Introduction

Targeted fishing of certain species within an ecosystem or a particular demographic within a population is known as selective fishing, an activity that may lead to detrimental imbalances (Law 2000; Bundy et al. 2005; Daan et al. 2005; Jorgensen et al. 2007; Anderson et al. 2008). Distributing a moderate fishing mortality across the widest possible range of species, stocks, and demographics in an ecosystem in proportion to their natural productivity could reduce the negative effects of selective fishing on biodiversity and population productivity (Zhou et al. 2010; Garcia et al. 2012). This 'balanced harvest' approach has gained traction as ecosystem-based strategies to fisheries management become more popular, but the purported necessity and benefits of a blanket balanced harvest approach continue to be debated (Jacobsen et al. 2013; Froese et al. 2015; Breen et al. 2016; Anderson et al. 2016; Froese et al. 2016; Kolding et al. 2016). There is, however, strong evidence that at least sex-selective harvesting can have a negative impact on reproductive rates and stock productivity, as well as alter sex ratio and life history (Clark and Tait 1982; Orensanz et al. 1998; Alonzo and Mangel 2004; Hamilton et al. 2007; Hutchings and Rowe 2008). When feasible, fishing regulations that promote exploitation of male and female fish in equal
proportions are therefore often preferred over those that result in a female-biased harvest.

Summer Flounder, Paralichthys dentatus, support important recreational and commercial fisheries along the northeast and mid-Atlantic coasts of the USA. In 2015, an estimated 12.5 million summer flounder were caught in the recreational fishery alone (Terceiro 2016). In this sector, management efforts to constrain harvest below the maximum allowable catch have typically involved increasing the minimum landing size; however, Summer Flounder are sexually dimorphic. Females grow faster and mature at a larger size than males (Poole 1961; Morse 1981; Packer et al. 1999). Thus, reliance on increased minimum size limits as a management strategy has resulted in approximately $90 \%$ of the recent recreational landings being female fish (Morson et al. 2012, 2015). Furthermore, the female fish that are targeted are also the largest, and therefore potentially the most fecund fish in the population (Morse 1981; Berkeley et al. 2004; Birkeland and Dayton 2005; Hixon et al. 2013; Shelton et al. 2015; Stige et al. 2017).

Slots limits offer an alternative management approach to traditional minimum size limits in that they restrict landings to some intermediate range of sizes while large and small fish are released (Gwinn et al. 2013). In the Summer Flounder recreational fishery, it was previously demonstrated that slot limits have the potential to increase landings in numbers under a fixed fishing exploitation rate by weight (Bochenek et al. 2010; Powell et al. 2010). If smaller males are vulnerable to recreational fishing effort, a similar approach that redirects some fraction of the fishing mortality toward smallersized fish may also balance the fishing mortality with respect to sex. However, the only
information available on the sex composition of the Summer Flounder recreational catch comes from fish that were landed and are therefore larger than the minimum size limit (Morson et al. 2012, 2015). Without an estimate of the sex composition for fish below the minimum landing size, it is not possible to evaluate the sex-specific outcomes of such alternative management options.

Given the highly sex-biased catch composition identified in Chapter 1 for the recreational Summer Flounder fishery, in this chapter I evaluate the potential for slot limits to produce a more sex-balanced harvest. To estimate the full size- and sexspecific vulnerability I sampled additional landings, as well as all discarded fish, caught by recreational anglers on select party boats from New Jersey to Rhode Island during the 2016 recreational fishing season. I then examined the performance of a wide array of slot limits to estimate which would have promoted a more sex-balanced harvest while maintaining a fixed fishing mortality given the observed catch composition.

## Materials and Methods

## Field program

Data collection focused on three states: New Jersey, New York, and Rhode Island; and one fishing mode: for-hire mode. In 2016, these states accounted for $82 \%$ of the total catch by state; however, the for-hire fishing mode accounted for only 5\% of the total catch by mode. While the for-hire mode accounted for only a small fraction of the total catch, the private mode, which accounted for $89 \%$, operated across a similar spatial scale. In $2016,34 \%$ and $31 \%$ of the total catch in the for-hire and private modes, respectively, came from open ocean waters less than or equal to three miles from the
coast. In the same year, $42 \%$ and $54 \%$ of the total catch in the for-hire and private modes, respectively, came from inland waters (personal communication from the National Marine Fisheries Service, Fisheries Statistics Division, 5/25/17). Given the two fishing modes access the same areas, data collection focused on the for-hire mode. The larger, for-hire vessels had space for up to 75 anglers which significantly increased the sample size potential on any one sampling trip.

Data collection spanned one entire recreational fishing season from May 23, 2016 through September 16, 2016. Fish were collected bi-weekly aboard participating fishing vessels from Cape May (8 trips) and Atlantic Highlands (8 trips), NJ, Captree (9 trips) and Montauk (10 trips), NY, and Point Judith, RI (6 trips) (Figure 2.1). The total length of every Summer Flounder caught, whether landed or discarded, on each trip was measured. In addition, the sex for all landed Summer Flounder was recorded. Since Summer Flounder must be dissected to determine sex, ten discarded fish were also sacrificed within predetermined fish length and water depth bins on each sampling trip (Table 2.1). A sex ratio, by depth bin, length bin, and trip, was applied to any unsexed discarded fish to assign sex to the entire discarded portion of the catch (Table 2.1).

## Analysis

To estimate the effect of different slot limits on three performance metrics, the total number of dead fish (landings + dead discards), the biomass of dead females, and the ratio of dead discards to total number dead, I simulated outcomes from twenty-one potential slot limits. The smallest lower size limit evaluated was 14 inches, the minimum size limit in the commercial fishery, and the largest upper size limit evaluated
was 21 inches, the largest minimum size ever implemented in the recreational Summer Flounder fishery. The width of the slots limits varied from a minimum width of two inches to a maximum width of seven inches.

A slot limit can be defined several ways, but here I am referring to a regulation where only fish within a minimum and maximum size limit could be kept while all fish that are greater than the maximum size or less than the minimum size must be discarded. Since I did not record catch-rate-per-angler, and could therefore not evaluate alternative bag limits within a given slot limit, all fish between the minimum retention size and maximum retention size of a slot limit were assumed to be kept (no bag limit). Each slot-limit was imposed on the observed catch and catch composition data overall, as well as by location and depth category, assuming effort was fixed at what was observed. Finally, $10 \%$ mortality was applied to all discarded fish in conformity with the recreational discard mortality rate currently applied in the Summer Flounder stock assessment (Terceiro 2016).

To convert observed individual fish lengths (in inches) to estimated individual fish weights (in pounds) I applied sex-specific parameters from Morse (1981) for the equation,

$$
w(l)=\alpha l^{\beta}
$$

The total number dead, $\mathrm{N}_{\mathrm{d}}$ for each simulated slot limit s , was calculated as:

$$
N_{d_{s}}=N_{L_{s}}+\left(0.10 * N_{D_{s}}\right)
$$

where $N_{L}$ is the total number of fish that were landed and $N_{D}$ is the total number of fish that were discarded. The biomass of dead females, $F_{d}$ was calculated as:

$$
F_{d_{s}}=\left[\sum_{i=1}^{n} W_{i_{L_{f_{s}}}}+\left(0.10 \times \sum_{i=1}^{n} w_{i_{D_{f_{s}}}}\right)\right]
$$

where $w$ is the weight in pounds of fish $i$ and $f$ denotes female fish. Finally, the ratio of dead discards to total dead was calculated as:
$R_{d_{s}}=\frac{\left(0.10 * N_{D_{s}}\right)}{N_{d_{s}}}$

The influence of each slot limit on these metrics was evaluated separately using the observed catch composition at each sampling location. For each slot limit, I calculated the proportional change from the observed metric at an 18-inch minimum retention size to the calculated metric given the slot limit. Slot limits that produced a $10 \%$ or less change, whether negative or positive, in the total number of dead fish, were deemed suitable alternatives in that they would not have resulted in a significant change in fishing mortality given total catch and total effort was fixed at what was observed. In other words, only slot limits that kept fishing exploitation rate near constant were viewed as potential alternatives to the 18 -inch minimum retention size.

## Results

## Observed catch composition

Under an 18-inch minimum size restriction, I observed a total catch of 3,290 Summer Flounder by recreational anglers on forty-one directed party boat trips (Table 2; Figure 2). Of the total catch, 2,645 ( $80 \%$ ) were discarded, however the proportion of the catch that was discarded varied by region and by depth category (Table 2). The discard proportion was lower in deeper water and in more northern ports (Table 2) where smaller fish occurred less frequently (Figures 3, 4). The proportion female increased with size resulting in $87 \%(n=558)$ of the landings and $38 \%(n=1,011)$ of the discards being female overall (Figure 2). However, the sex ratio at length varied by location and by depth category. At a given length, the sex ratio was more heavily skewed toward female fish in shallower water and in more southern ports (Table 2; Figures 3, 4).

## Simulated catch composition

Slot limits that kept the total number dead at or near that observed under the 18inch minimum size limit, herein referred to as "suitable", were all narrow, ranging from two to four inches wide. Of the suitable slot limits, most contained 18 inches within the slot limit and only once did the bottom of the slot limit fall below 16 inches, when a 15to 17 -inch slot limit was suitable in Atlantic Highlands, NJ. All suitable slot limits reduced the total biomass of dead females, one of which, referred to herein as "optimal", produced the greatest reduction in total biomass of dead females. Few suitable slot limits had a significant impact on the proportion of the total dead made up of dead discards (Table 3).

No single optimal slot limit minimized dead female biomass at every location.
Variation in length frequency and sex ratio at length observed across the different
sampling locations produced varying suitable and optimal slot limits at each sampling location (Figure 3; Table 3). In Cape May, New Jersey a 17- to 19 -inch slot would have been optimal, reducing dead female biomass by $31 \%$ (from 262.37 lb to 181.59 lb ), while in the same state, in Atlantic Highlands, a 15- to 17-inch slot would have been optimal, producing a $58 \%$ reduction in dead female biomass (from 537.85 lb to 226.89 lb). Similarly, in New York, the catch composition from Captree would have generated a $21 \%$ reduction in dead female biomass (from 181.1 lb to 143.93 lb ) at an 18 - to 20 -inch optimal slot limit, while in Montauk, the optimal slot limit was 16 to 18 inches, which would have produced a $55 \%$ reduction (from 533.27 lb to 238.04 lb ). Finally, in Pt. Judith, Rhode Island, among slot limits deemed suitable, a three-inch slot from 17 to 20 inches would have been optimal, generating a 55\% reduction in dead female biomass (from 222.94 lb to 100.48 lb ). At some sampling locations, the potential to reduce dead female biomass came primarily from males being more accessible at lower sizes, for example in Atlantic Highlands, NJ (Figure 3b) and in Montauk, NY (Figure 3d), while in other sampling locations it came primarily from the protection of larger females, for example in Cape May, NJ (Figure 3a) and in Captree, NY (Figure 3c) and less from accessing smaller males at lower sizes.

## Discussion

The overall sex composition and the spatial and depth-dependent trends in sex ratio of large fish matched what has been previously reported for the recreational Summer Flounder fishery (Morson et al. 2012, 2015). Furthermore, the overall discard rate, $80 \%$, is similar to the $79 \%$ discard rate estimated for the entire for-hire fishing mode in

2016 (personal communication from the National Marine Fisheries Service, Fisheries Statistics Division, 5/25/17). The proportion female in the larger size classes is higher than that observed in the commercial catch or on the NMFS-NEFSC trawl survey (Morson et al. 2015). However, while the observed sex ratio pattern for the larger fish suggests large males do not come inshore and are therefore not available to the recreational fishery when and where it takes place, I show here that smaller-sized males are both available inshore and show up in the recreational catch at smaller sizes. This is an important finding for two reasons. First, it suggests that any sex-specific movement or habitat use that produces such a highly skewed sex ratio in the landings must also be size-specific. That is, while large male fish may remain offshore where they are less likely to be accessible to the recreational fishery, smaller males do move inshore in the spring/summer where they are available to the recreational fishing effort. Sex- and size-dependent separation in space and time has been well documented in other sexually dimorphic flatfishes (Swain 1997; Swain and Morin 1997; Sahin and Gunes 2010; Loher and Hobden 2012), so the occurrence of this behavior in Summer Flounder is not surprising and ecological theory for intraspecific partitioning of resources along a life history is well established (Schoener 1968). Second, the availability of male fish at lower sizes enables changes in size regulations to influence the sex composition of the catch. This second point is especially important given how much of the fishing mortality is being directed at the female portion of the stock in this fishery under the current minimum size restrictions and the desire to evaluate alternative management options that could promote a more sex-balanced harvest (Morson et al. 2012, 2015).

Slot limits have been demonstrated to produce higher harvest numbers, maintain natural age structure in the population, reduce discard mortality, positively influence recruitment potential, and conserve biomass across a variety of fish life histories (Birkeland and Dayton 2005; Powell et al. 2010; Koehn and Todd 2012; Law et al. 2012; Gwinn et al. 2013; Sanchez-Hernandez et al. 2016). Here I add additional support to a growing body of literature that suggests slot limits can simultaneously achieve a number of desired management goals and demonstrate that, for fish with sexuallydimorphic growth, slot limits have the potential to distribute sex-biased fishing exploitation more evenly across both sexes. In the Summer Flounder recreational fishery, there is an obvious trade-off available to managers where the catch of large, heavy, female fish could be replaced by a similar number of smaller, lighter, male and female fish.

One important consideration in the evaluation of any slot limit relative to a minimum size limit is whether the lifetime spawner reproductive potential is negatively affected. Since nearly all of the lower ends of the slot limits prescribed here as optimal are only an inch or two smaller than the current 18-inch size limit, both the current 18-inch size limit and the prescribed optimal slot limits would allow fish the opportunity to spawn multiple times before recruiting to the fishery. Nevertheless, to fully evaluate the longterm impacts of varying slot limits on stock productivity, a spawning stock biomass per recruit analysis would be necessary (Haddon 2011). A more appropriate method for evaluating viable slot limit options and potential outcomes therefore is a management strategy evaluation that links annual management decisions on slot limits with a stock assessment model and includes annual, seasonal, and spatial dynamics in the
population, the catch composition given the prescribed slot, and the fishing effort (Punt et al. 2014). Such an evaluation is beyond the scope of this work, however the sex composition data now exist for the full range of sizes available to the recreational fishery so the development of such a model is an obvious next step.

Several important limitations are inherent in the data and analysis I present herein. First, my approach assumes effort would have remained constant under alternative management scenarios. If, for example, a given slot limit would encourage more anglers to participate in the fishery than participated in it under an 18-inch minimum size limit, the total catch under any alternative scenario may have increased, resulting in a higher total catch, and mortality, relative to the observed. Second, this work focused on the catch of the for-hire mode and did not sample the private or shore-based modes. It is possible the catch size and catch length and sex composition vary across these other fishing modes, which would alter the observed catch as well as the simulated outcomes of different slot limits. Finally, the observed catch is only representative of the catch in the year in which it was collected. Having observed a catch composition that appears to support a specific slot limit this year does not guarantee the same measure would be appropriate in any other year. For this reason, slot limits as a general strategy have the potential to be highly successful in achieving multiple management goals, particularly relative to a sex-balanced harvest in the recreational Summer Flounder fishery, but should be viewed as optimal given the observed catch composition for the year, locations, and mode I sampled and further evaluation of interannual, spatial, and mode-specific dynamics is warranted.

In conclusion, the availability of male fish to the recreational fishing effort at smaller sizes, identified here for the first time, suggests the conservation of large, female fish is achievable in this fishery with prescribed management actions. I demonstrate a few viable options that would have achieved either a more sex-balanced harvest or a reduction in dead female biomass under the catch conditions in the mode, location, and time I observed. However, a more robust, spatially- and temporally-dynamic management strategy evaluation could be used to estimate how alternative applications, including slot limits and trophy limits, would perform for this fishery in any given year so that multiple management goals, including limiting mortality of large females, increasing angler satisfaction, and balancing harvest sex ratio under a fixed harvest rate might be achieved simultaneously.

## Acknowledgements

I thank the captains and crews of the F/V Porgy IV, Bonanza II, Big Mohawk, Fishermen, Laura Lee, Lazy Bones, and Gail Frances for permitting us to collect samples aboard their fishing vessels. I am grateful to Sarah Borsetti, Emerson Hasbrouck, Scott CuratoWagemann, Tara Froelich, and Kristin Gerbino for helping with data collection. Eleanor Bochenek assisted with coordinating sampling trips, for which I am thankful. Helpful comments were provided by Rich Wong and two anonymous reviewers. Partial funding for this work was provided by the National Science Foundation Industry/University Cooperative Research Center SCeMFiS (Science Center for Marine Fisheries) through membership fees under the direction of the Industry Advisory Board (IAB). SCeMFiS administrative support is provided by NSF award no. 1266057. Additional funding
provided by the Save the Summer Flounder Fishery Fund, and the Jersey Coast Anglers Association.

## Literature Cited - Chapter 2

Alonzo, S. H., and M. Mangel. 2004. The effects of size-selective fisheries on the stock dynamics of and limitation in sex-changing fish. Fishery Bulletin 102 (1): 1-13.

Anderson, C. N. K., Hsieh, C., S. A. Sandin, R. Hewitt, A. Hollowed, J. Beddington, R. M. May, and G. Sugihara. 2008. Why fishing magnifies fluctuations in fish abundance. Nature 452: 835-839.

Anderson, K. H., J. L. Blanchard, E. A. Fulton, H. Gislason, N. S. Jacobsen, and T. van Kooten. 2016. Assumptions behind size-based ecosystem models are realistic. ICES Journal of Marine Science 73: 1651-1655.

Berkeley, S. A., C. Chapman, and S. M. Sogard. 2004. Maternal age as a determinant of larval growth and survival in a marine fish, Sebastes melanops. Ecology 85 (5): 12581264.

Birkeland, C. and P. K. Dayton. 2005. The importance in fishery management of leaving the big ones. Trends in Ecology and Evolution 20 (7): 356-358.

Bochenek, E. A., E. N. Powell, J. DePersenaire, and S. E. King. 2010. Evaluating catch, effort, and bag limits on directed trips in the recreational Summer Flounder party boat fishery. Marine and Coastal Fisheries 2: 412-423.

Breen, M., N. Graham, M. Pol, P. He, D. Reid, and P. Suuronen. 2016. Selective fishing and balanced harvesting. Fisheries Research 184: 2-8.

Bundy, A., P. Fanning, and K. C. T. Zwanenburg. 2005. Balancing exploitation and conservation of the eastern Scotian Shelf ecosystem: application of a 4D ecosystem exploitation index. ICES Journal of Marine Science 62 (3): 503-510.

Clark, C. W. and D. E. Tait. 1982. Sex-selective harvesting of wildlife populations. Ecological Modeling 14(3-4) 251-260.

Daan, N., H. Gislason, J. G. Pope, and J. C. Rice. 2005. Changes in the North Sea fish community: evidence of indirect effects of fishing? ICES Journal of Marine Science 62: 177-188.

Froese, R., C. Walters, D. Pauly, H. Winker, O. L. F. Weyl, N. Demirel, A. C. Tsikliras, and S. Holt. 2015. A critique of the balanced harvesting approach to fishing. ICES Journal of Marine Science 73: 1640-1650.

Froese, R., C. Walters, D. Pauly, H. Winker, O. F. Weyl, N. Demirel, A. C. Tsikliras, and S. Holt. 2016. Reply to Anderson et al. (2016) "Assumptions behind size-based ecosystem models are realistic." ICES Journal of Marine Science 73(6): 1656-1658.

Garcia, S. E., J. Kolding, J. Rice, M. J. Rochet, S. Zhou, T. Arimoto, J. E. Beyer, L. Borges, A. Bundy, D. Dunn, E. A. Fulton, M. Hall, M. Heino, R. Law, M. Makino, A. Rijnsdorp, F. Simard, and A. Smith. 2012. Reconsidering the consequences of selective fisheries. Science 335: 1045-1047.

Gwinn, D. C., M. S. Allen, F. D. Johnston, P. Brown, C. R. Todd, and R. Arlinghaus. 2013. Rethinking length-based fisheries regulations: the value of protecting old and large fish with harvest slots. Fish and Fisheries 16 (2): 259-281.

Haddon, M. 2011. Modeling and quantitative methods in fisheries. Taylor and Francis, New York, NY.

Hamilton, S. L., J. E. Caselle, J. D. Standish, D. M. Schroeder, M. S. Love, J. A. RosalesCasian, and O. Sosa-Nishizaki. 2007. Size-selective harvesting alters life histories of a temperate sex-changing fish. Ecological Applications 17 (8): 2268-2280.

Hutchings, J. A. and S. Rowe. 2008. Consequences of sexual selection for fisheriesinduced evolution: an exploratory analysis. Evolutionary Applications 1: 129-136.

Hixon, M. A., D. W. Johnson, and S. M. Sogard. 2013. BOFFFFs: on the importance of conserving old-growth age structure in fishery populations. ICES Journal of Marine Science 71 (8): 2171-2185.

Jacobsen, N. S., H. Gislason, and K. H. Anderson. 2013. Consequences of balanced harvesting of fish communities. Proceedings of the Royal Society B 281 (1775): 1-9.

Jorgensen, C., K. Enberg, E. S. Dunlop, R. Arlinghaus, D. S. Boukal, K. Brander, B. Ernande, A. Gardmark, F. Johnston, S. Matsumura, H. Pardoe, K. Raab, A. Silva, A. Vainikka, U. Dieckmann, M. Heino, and A. D. Rijnsdorp. 2007. Ecology: managing evolving fish stock. Science 318: 1247-1248.

Kolding, J., S. M. Garcia, S. Zhour, and Mikko Heino. 2016. Balanced harvest: utopia, failure, or functional strategy? ICES Journal of Marine Science
doi:10.1093/icesjms/fsw060
Koehn, J. D. and C. R. Todd. 2012. Balancing conservation and recreational fishery objectives for a threatened fish species, the Murray cod, Maccullochella peelii. Fisheries Management and Ecology 19 (5): 410-425.

Law, R. 2000. Fishing, selection, and phenotypic evolution. ICES Journal of Marine Science 57 (3): 659-668.

Law, R., M. J. Plank, and J. Kolding. 2012. On balanced exploitation of marine ecosystems: results from dynamic size spectra. ICES Journal of Marine Science 69 (4): 602-614.

Loher, T. and J. Hobden. 2012. Length and sex effects on the spatial structure of catches of Pacific Halibut (Hippoglossus stenolpis) on longline gear. U.S. National Marine Fisheries Service Fishery Bulletin 110:46-51.

Morse, W. W. 1981. Reproduction of the Summer Flounder Paralichthys dentatus (L.). Journal of Fish Biology 19 (2): 189-203.

Morson, J. M., E. A. Bochenek, E. N. Powell, and J. E. Gius. 2012. Sex at length of Summer Flounder landed in the New Jersey recreational party boat fishery. North American Journal of Fisheries Management 32 (6): 1201-1210.

Morson, J. M., E. A. Bochenek, E. N. Powell, E. C. Hasbrouck, J. E. Gius, C. F. Cotton, K, Gerbino, and T. Froelich. 2015. Estimating the sex composition of the Summer Flounder catch using fishery-independent data. Marine and Coastal Fisheries 7 (1): 393-408.

Orensanz, J. M., J. Armstrong, D. Armstrong, and R. Hilborn. 1998. Crustacean resources are vulnerable to serial depletion - the multifaceted decline of crab and shrimp fisheries in the Greater Gulf of Alaska. Reviews in Fish Biology and Fisheries 8: 117176.

Packer, D. B., S. J. Griesbach, P. L. Berrien, C. A. Zetlin, D. L. Johnson, and W. W. Morse. 1999. Essential fish habitat source document: Summer Flounder, Paralichthys dentatus, life history and habitat characteristics. NOAA Technical Memorandum NMFS-NE-151.

Poole, J. C. 1961. Age and growth of the fluke in Great South Bay and their significance to the sport fishery. New York Fish and Game Journal 8: 1-18.

Powell, E. N., E. A. Bochenek, and J. DePersenaire. 2010. Evaluation of bag-and-sizelimit options in the management of Summer Flounder Paralichthys dentatus. Fisheries Research 105: 215-227.

Punt, A., D. S. Butterworth, C. L. de Moor, J. A. A. De Oliveira, and M. Haddon. 2014. Management strategy evaluation: best practices. Fish and Fisheries: 17 (2): 303-334.

Sahin, T. and E. Gunes. 2010. Seasonal variation in length, weight, and sex distribution of flounder (Paralichthys flesus luscus) in the southeastern Black Sea. Journal of Fisheries Sciences 4: 238-245.

Sanchez-Hernandez, J., S. L. Shaw, F. Cobo, and M. S. Allen. 2016. Influence of a minimum-length limit regulation on wild brown trout: an example of recruitment and growth overfishing. North American Journal of Fisheries Management 36 (5): 10241035.

Schoener, T. W. 1968. Resource partitioning in ecological communities. Science 185: 27 39.

Shelton, A. O., J. A. Hutchings, R. S. Waples, D. M. Keith, H. Resit Akcakaya, and N. K. Dulvy. 2015. Maternal age effects on Atlantic cod recruitment and implications for future population trajectories. ICES Journal of Marine Science 72(6): 1769-1778.

Stige, L. C., N. A. Yaragina, O. Langangen, B. Bogstad, N. C. Stenseth, and G. Ottersen. 2017. Effect of a fish stock's demographic structure on offspring survival and sensitivity to climate. Proceedings of the National Academy of Sciences 114 (6): 13471352.

Swain, D. P. 1997. Sex-specific temperature distribution of American Plaice (Hippoglossoides platessoides) and its relation to age and abundance. Canadian Journal of Fisheries and Aquatic Sciences 54: 1077-1087.

Swain, D. P. and R. Morin. 1997. Effects of age, sex and abundance on the bathymetric pattern of the American Plaice (Hippoglossoides platessoides) in the southern gulf of St. Lawrence. Canadian Journal of Fisheries and Aquatic Sciences 53: 106-119.

Terceiro, M. 2016. Stock assessment of Summer Flounder for 2016. Northeast Fisheries Science Center, Reference Document 16-15. Woods Hole, MA.

Zhou, S., A. D. M. Smith, A. E. Punt, A. J. Richardson, M. Gibbs, E. A. Fulton, S. Pascoe, C. Bulman, P. Bayliss, and K. Sainsbury. 2010. Ecosystem-based fisheries management requires a change to the selective fishing philosophy. Proceedings of the National Academy of Sciences 107 (21): 9485-9489.

## TABLES - CHAPTER 2

Table 2.1. Length and depth categories for sub-sampling the sex of discarded fish.

| Length bins (inches) | Depth bins (feet) |
| :---: | :---: |
| $8.0-9.9$ | $0-25.4$ |
| $10.0-11.9$ | $25.5-50.4$ |
| $12.0-13.9$ | $50.5-75.4$ |
| $14.0-15.9$ | $75.5+$ |
| $16.0-17.9$ |  |

Table 2.2. Total number of summer flounder sampled by port, depth range, outcome (landed/discarded), and sex. A dash indicates a given cell was not sampled while a zero indicates there were zero fish for that cell.

| State | Port | Depth <br> Range (ft) | Outcome | Female | Male |
| :---: | :---: | :---: | :---: | :---: | :---: |
| New Jersey |  |  |  |  |  |
|  | Cape May |  |  |  |  |
|  |  | 0-25 |  |  |  |
|  |  |  | Landed | - | - |
|  |  |  | Discarded | - | - |
|  |  | 25-50 |  |  |  |
|  |  |  | Landed | - | - |
|  |  |  | Discarded | - | - |
|  |  | 50-75 |  |  |  |
|  |  |  | Landed | 60 | 4 |
|  |  |  | Discarded | 144 | 229 |
|  |  | 75+ |  |  |  |
|  |  |  | Landed | 28 | 0 |
|  |  |  | Discarded | 37 | 138 |
|  | Atlantic Highlands |  |  |  |  |
|  |  | 0-25 |  |  |  |
|  |  |  | Landed | 0 | 0 |
|  |  |  | Discarded | 9 | 24 |
|  |  | 25-50 |  |  |  |
|  |  |  | Landed | 166 | 16 |
|  |  |  | Discarded | 150 | 209 |
|  |  | 50-75 |  |  |  |
|  |  |  | Landed | 8 | 2 |
|  |  |  | Discarded | 75 | 118 |
|  |  | 75+ |  |  |  |
|  |  |  | Landed | - | - |
|  |  |  | Discarded | - | - |
| New York |  |  |  |  |  |
|  | Captree |  |  |  |  |
|  |  | 0-25 |  |  |  |
|  |  |  | Landed | 42 | 2 |
|  |  |  | Discarded | 163 | 169 |
|  |  | 25-50 |  |  |  |
|  |  |  | Landed | 13 | 0 |
|  |  |  | Discarded | 53 | 74 |
|  |  | 50-75 |  |  |  |
|  |  |  | Landed | 8 | 5 |
|  |  |  | Discarded | 70 | 85 |



Table 2.3. Observed ( 18 -inch minimum size limit, top row) and simulated incremental slot limit performance metrics, including total number dead ( $\mathrm{N}_{\mathrm{d}}$ ), dead female biomass $\left(\mathrm{F}_{\mathrm{d}}\right)$, and ratio of dead discards to total dead ( $\mathrm{R}_{\mathrm{d}}$ ). In parentheses are the percent changes in a given performance metric relative to the observed (18-inch minimum size limit). Slot limits in bold are within $+/-10 \%$ of the observed total number dead.

| State | Port | Bottom of Slot Limit | Top of Slot Limit | Total Number Dead | Dead Female Biomass | Ratio of Dead Discards to Total Dead |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| New Jersey |  |  |  |  |  |  |
|  |  |  |  |  |  |  |
|  | Cape May | 18.00 | - | 147 | 262.37 | 0.37 |
|  |  | 14.00 | 15.99 | 321 (118\%) | 117.35 (-55\%) | 0.11 (-70\%) |
|  |  | 14.00 | 16.99 | 411 (180\%) | 187.01 (-29\%) | 0.06 (-84\%) |
|  |  | 14.00 | 17.99 | 447 (204\%) | 244.6 (-7\%) | 0.05 (-86\%) |
|  |  | 14.00 | 18.99 | 487 (231\%) | 321.34 (22\%) | 0.03 (-92\%) |
|  |  | 14.00 | 19.99 | 506 (244\%) | 369.87 (41\%) | 0.03 (-92\%) |
|  |  | 14.00 | 20.99 | 519 (253\%) | 409.91 (56\%) | 0.03 (-92\%) |
|  |  | 15.00 | 16.99 | 273 (86\%) | 153.98 (-41\%) | 0.15 (-59\%) |
|  |  | 15.00 | 17.99 | 310 (111\%) | 211.57 (-19\%) | 0.12 (-68\%) |
|  |  | 15.00 | 18.99 | 349 (137\%) | 288.31 (10\%) | 0.09 (-76\%) |
|  |  | 15.00 | 19.99 | 368 (150\%) | 336.84 (28\%) | 0.08 (-78\%) |
|  |  | 15.00 | 20.99 | 382 (160\%) | 376.88 (44\%) | 0.08 (-78\%) |
|  |  | 16.00 | 17.99 | 190 (29\%) | 174.52 (-33\%) | 0.26 (-30\%) |
|  |  | 16.00 | 18.99 | 230 (56\%) | 251.26 (-4\%) | 0.2 (-46\%) |
|  |  | 16.00 | 19.99 | 249 (69\%) | 299.79 (14\%) | 0.18 (-51\%) |
|  |  | 16.00 | 20.99 | 262 (78\%) | 339.83 (30\%) | 0.16 (-57\%) |
|  |  | 17.00 | 18.99 | 141 (-4\%) | 181.59 (-31\%) | 0.4 (8\%) |
|  |  | 17.00 | 19.99 | 159 (8\%) | 230.13 (-12\%) | 0.34 (-8\%) |
|  |  | 17.00 | 20.99 | 173 (18\%) | 270.17 (3\%) | 0.3 (-19\%) |
|  |  | 18.00 | 19.99 | 123 (-16\%) | 172.54 (-34\%) | 0.47 (27\%) |
|  |  | 18.00 | 20.99 | 136 (-7\%) | 212.58 (-19\%) | 0.41 (11\%) |
|  |  | 19.00 | 20.99 | 96 (-35\%) | 135.84 (-48\%) | 0.63 (70\%) |
|  | Atlantic |  |  |  |  |  |
|  | Highlands | 18.00 | - | 251 | 537.85 | 0.23 |
|  |  | 14.00 | 15.99 | 301 (20\%) | 154.23 (-71\%) | 0.18 (-22\%) |
|  |  | 14.00 | 16.99 | 411 (64\%) | 259.93 (-52\%) | 0.1 (-57\%) |
|  |  | 14.00 | 17.99 | 466 (86\%) | 327.79 (-39\%) | 0.07 (-70\%) |
|  |  | 14.00 | 18.99 | 545 (117\%) | 461.88 (-14\%) | 0.05 (-78\%) |
|  |  | 14.00 | 19.99 | 575 (129\%) | 539.13 (0\%) | 0.04 (-83\%) |
|  |  | 14.00 | 20.99 | 602 (140\%) | 616.99 (15\%) | 0.03 (-87\%) |
|  |  | 15.00 | 16.99 | 277 (10\%) | 226.89 (-58\%) | 0.2 (-13\%) |
|  |  | 15.00 | 17.99 | 332 (32\%) | 294.75 (-45\%) | 0.15 (-35\%) |
|  |  | 15.00 | 18.99 | 411 (64\%) | 428.84 (-20\%) | 0.1 (-57\%) |


|  | 15.00 | 19.99 | 441 (76\%) | 506.09 (-6\%) | 0.08 (-65\%) |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  | 15.00 | 20.99 | 467 (86\%) | 583.95 (9\%) | 0.07 (-70\%) |
|  | 16.00 | 17.99 | 242 (-4\%) | 254.37 (-53\%) | 0.25 (9\%) |
|  | 16.00 | 18.99 | 322 (28\%) | 388.46 (-28\%) | 0.16 (-30\%) |
|  | 16.00 | 19.99 | 352 (40\%) | 465.71 (-13\%) | 0.13 (-43\%) |
|  | 16.00 | 20.99 | 378 (51\%) | 543.57 (1\%) | 0.12 (-48\%) |
|  | 17.00 | 18.99 | 212 (-16\%) | 282.76 (-47\%) | 0.3 (30\%) |
|  | 17.00 | 19.99 | 242 (-4\%) | 360 (-33\%) | 0.25 (9\%) |
|  | 17.00 | 20.99 | 269 (7\%) | 437.86 (-19\%) | 0.21 (-9\%) |
|  | 18.00 | 19.99 | 188 (-25\%) | 292.14 (-46\%) | 0.35 (52\%) |
|  | 18.00 | 20.99 | 214 (-15\%) | 370 (-31\%) | 0.29 (26\%) |
|  | 19.00 | 20.99 | 134 (-47\%) | 235.91 (-56\%) | 0.53 (130\%) |
| New York |  |  |  |  |  |
| Captree | 18.00 | - | 132 | 181.1 | 0.47 |
|  | 14.00 | 15.99 | 285 (116\%) | 162.17 (-10\%) | 0.16 (-66\%) |
|  | 14.00 | 16.99 | 366 (177\%) | 243.48 (34\%) | 0.1 (-79\%) |
|  | 14.00 | 17.99 | 393 (198\%) | 283.12 (56\%) | 0.08 (-83\%) |
|  | 14.00 | 18.99 | 436 (230\%) | 357.99 (98\%) | 0.06 (-87\%) |
|  | 14.00 | 19.99 | 445 (237\%) | 380.35 (110\%) | 0.06 (-87\%) |
|  | 14.00 | 20.99 | 454 (244\%) | 406.74 (125\%) | 0.06 (-87\%) |
|  | 15.00 | 16.99 | 213 (61\%) | 175.18 (-3\%) | 0.25 (-47\%) |
|  | 15.00 | 17.99 | 240 (82\%) | 214.82 (19\%) | 0.21 (-55\%) |
|  | 15.00 | 18.99 | 283 (114\%) | 289.7 (60\%) | 0.16 (-66\%) |
|  | 15.00 | 19.99 | 292 (121\%) | 312.05 (72\%) | 0.15 (-68\%) |
|  | 15.00 | 20.99 | 301 (128\%) | 338.45 (87\%) | 0.14 (-70\%) |
|  | 16.00 | 17.99 | 177 (34\%) | 167.65 (-7\%) | 0.32 (-32\%) |
|  | 16.00 | 18.99 | 219 (66\%) | 242.53 (34\%) | 0.24 (-49\%) |
|  | 16.00 | 19.99 | 228 (73\%) | 264.88 (46\%) | 0.22 (-53\%) |
|  | 16.00 | 20.99 | 237 (80\%) | 291.28 (61\%) | 0.21 (-55\%) |
|  | 17.00 | 18.99 | 138 (5\%) | 161.22 (-11\%) | 0.44 (-6\%) |
|  | 17.00 | 19.99 | 147 (11\%) | 183.57 (1\%) | 0.41 (-13\%) |
|  | 17.00 | 20.99 | 156 (18\%) | 209.97 (16\%) | 0.38 (-19\%) |
|  | 18.00 | 19.99 | 120 (-9\%) | 143.93 (-21\%) | 0.52 (11\%) |
|  | 18.00 | 20.99 | 129 (-2\%) | 170.33 (-6\%) | 0.48 (2\%) |
|  | 19.00 | 20.99 | 87 (-34\%) | 95.45 (-47\%) | 0.77 (64\%) |
| Montauk | 18.00 | - | 261 | 533.27 | 0.28 |
|  | 14.00 | 15.99 | 404 (55\%) | 207.19 (-61\%) | 0.14 (-50\%) |
|  | 14.00 | 16.99 | 529 (103\%) | 312.22 (-41\%) | 0.08 (-71\%) |
|  | 14.00 | 17.99 | 573 (120\%) | 361.73 (-32\%) | 0.07 (-75\%) |
|  | 14.00 | 18.99 | 627 (140\%) | 448.41 (-16\%) | 0.05 (-82\%) |
|  | 14.00 | 19.99 | 656 (151\%) | 511.83 (-4\%) | 0.04 (-86\%) |
|  | 14.00 | 20.99 | 699 (168\%) | 635.37 (19\%) | 0.03 (-89\%) |
|  | 15.00 | 16.99 | 326 (25\%) | 249.51 (-53\%) | 0.2 (-29\%) |



## FIGURES - CHAPTER 2



Figure 2.1. Map of sampling locations.


Figure 2.2. Total number (A) and weight (B) of male and female Summer Flounder collected at each 1-inch length bin. Solid black line represents proportion female-atlength. Vertical dashed line represents the 18-inch minimum landing size in 2016.


Figure 2.3. Total number of male and female Summer Flounder collected at (A) Cape May, NJ, (B) Atlantic Highlands, NJ, (C) Captree, NY, (D) Montauk, NY, and (E) Pt. Judith, RI in each 1-inch length bin. Solid black line represents proportion female-at-length. Note differences in primary $y$-axis for each panel.


Figure 2.4. Total number of male and female Summer Flounder collected in (A) 0-25 feet (B) 25-50 feet (C) 50-75 feet, and (D) 75+ feet of water at each 1-inch length bin. Solid black line represents proportion female-at-length. Note differences in primary yaxis for each panel

## CHAPTER 3

# DENSITY-DEPENDENT CAPTURE EFFICIENCY OF A SURVEY DREDGE AND ITS INFLUENCE ON THE STOCK ASSESSMENT OF EASTERN OYSTERS CRASSOSTREA VIRGINICA IN DELAWARE BAY 


#### Abstract

A reliable measure of gear capture efficiency is required to calculate unbiased estimates of population size and fishing mortality from survey data in a stock assessment. However, capture efficiency can vary spatially and temporally due to changes in abundance, stock area, the environment, and the sampling gear itself. Therefore, periodic reassessment of this parameter is necessary to ensure that the catchability coefficients being applied accurately reflect the capture efficiency of the survey sampling gear, especially when catchability is being estimated outside of the stock assessment model. Using data from field experiments conducted in 1999, 2000, 2003, and 2013, I evaluated spatial and temporal variability in capture efficiency for a commercial dredge used to conduct a fishery-independent survey of the eastern oyster (Crassostrea virginia) population in Delaware Bay, USA. A spatial gradient in capture efficiency was detected, but no temporal trend. Capture efficiency was a function of the density of oysters in the sampled area. To my knowledge this is the first time densitydependent capture efficiency has been identified for a sessile invertebrate stock survey. Since density dependence in capture efficiency leads to hyperstable catch-per-uniteffort, caution is advised when deriving oyster abundance from dredge survey catch-


per-effort data, especially at low oyster density and when high spatial resolution estimates of survey dredge capture efficiency are not available.

## Introduction

Stock assessment of oyster populations is typically done using data collected from surveys that employ one of three types of sampling gear. Diver quadrat sampling and patent tongs are nearly $100 \%$ efficient for collecting oysters within the area sampled (Chai et al. 1993; Mann et al. 2004). However, patent tongs can be destructive to the reef and can only sample a small area per unit of effort; therefore, some have advocated against their long-term use for surveying natural resources (Rothschild et. al 1994). Diver sampling suffers from the same limitation in sample size and typically also suffers from limited sample numbers due to the time required per sample. Alternatively, a commercial dredge can be used to sample a large area, providing a high sample density at relatively low cost, but is also inefficient (Powell et al. 2002; Mann et al. 2004; Powell et al. 2007; Marenghi et al. 2017). When the sampling gear captures less than $100 \%$ of the individuals in the sampled area, it is important to estimate the efficiency of the gear and to account for this when converting survey catch-per-unit-effort (CPUE) data to an index of population size (Hilborn and Walters 1992; Arreguin-Sanchez 1996).

The capture efficiency of survey gear, and the catchability coefficient derived from capture efficiency, can be estimated within the stock assessment model (Fournier et al. 1998) or can be explicitly defined, either as a fixed value (Powell et al. 2007), or by some functional relationship (Wilberg et al. 2010). Field and experimental observations from depletion experiments (Lasta and Iribarne 1997; Gedamke et al.

2004; Rago et al. 2006; Hennen et al. 2012; Wilberg et al. 2013) and simultaneous sampling of survey gear and gear that is 100\% efficient (Powell et al. 2002; Mann et al. 2004; Powell et al. 2007; Singh et al. 2014) have been used to estimate catchability of shellfish survey dredges directly. Often the capture efficiency will vary spatially (Powell et al. 2002, 2007; Doray et al. 2010; Marenghi 2017) and temporally (Walters and Martell 2004; Powell et al. 2007), likely due to changes in abundance, stock area, the environment, and the sampling gear (Wilberg et al. 2010). For this reason, periodic reassessment of survey dredge capture efficiency has been advised (Powell et al. 2007; Marenghi et al. 2017).

The dredge survey used in the annual stock assessment for the eastern oyster Crassostrea virginica in Delaware Bay dates back to 1953 and provides one of the longest continuous abundance records of any shellfish population (Powell et al. 2008). Field experiments conducted in 1999, 2000 and 2003 identified spatial and temporal trends in the capture efficiency of the dredge used for this survey (Powell et al. 2002, 2007). To account for these, time- and location-specific catchability coefficients have been applied in the stock assessment to estimate an index of abundance from survey CPUE (Ashton-Alcox et al. 2015). However, Powell et al. (2007) suggest that capture efficiency of this survey gear may change rapidly, over only a few years at any one location. If the catchability coefficients currently being applied in the Delaware Bay oyster stock assessment no longer accurately reflect the capture efficiency of the sampling gear, this will lead to biased estimates of population size and exploitation rate (Pope and Shepherd 1985; Wilberg and Bence 2006) with the magnitude of the bias linked directly to the magnitude of the error in the estimate of capture efficiency used.

In this chapter I use data from a set of field experiments to update estimates of capture efficiency for the survey dredge used to estimate abundance in the stock assessment of the Delaware Bay eastern oyster population. I evaluate whether the time- and location-specific coefficients calculated from earlier experiments (1999, 2000, and 2003), which have been applied in the stock assessment, are still appropriate given new data collected during an experiment in 2013.

## Methods

## Field sampling

Capture efficiency was estimated at twelve oyster reef locations in Delaware Bay in 1999, 2000, 2003, and 2013 (Figure 3.1). At each location, three measurements of capture efficiency were collected within a $0.2^{\prime}$ latitude by $0.2^{\prime}$ longitude grid (approximately 25 acres). For the 1999-2003 experiments, measurements consisted of paired dredge tows and diver transect samples. Paten tongs and divers are equally efficient at sampling oysters (Chai et al. 1992). Since patent tongs were more readily available at the time, measurements from the 2013 experiment consisted of paired dredge tows and patent tong transect samples. For each replicate, a one-minute tow was conducted by the $F / V$ Howard $W$. Sockwell using a standard 1.27 -m commercial dredge and twelve haphazardly placed $0.25 \mathrm{~m}^{2}$ diver quadrats or six haphazardly placed $0.89 \mathrm{~m}^{2}$ patent tong samples were taken by a second boat in a transect parallel to but not in the dredge path. The time and GPS coordinates were recorded when the dredge tow began and ended. The total volume caught in each tow was measured as 37-L bushels in a pre-calibrated hopper. If the dredge was full after the 1-minute tow,
the tow time was reduced to 45 s so that the total number of bushels of material collected in the swept area could be determined. A single bushel was collected from each tow as a representative sub-sample of the catch and the remaining catch was discarded overboard. The entire tong or diver quadrat sample, minus the sediment, were retained for analysis. Both dredge and tong or diver samples were sorted into oysters, cultch (oyster shell), and boxes (dead oysters with the hinge still intact).

Volumes and weights of each component of the catch were recorded and all oysters and boxes in each sample were counted and measured.

## Data analyses

## Deriving capture efficiency

The swept area, $s a$, for each dredge tow, in square meters, was calculated as
(1) $\quad s a=d^{*} w$
where $d$ is the distance towed in meters determined by tallying the distances between GPS coordinates every 10 seconds during the tow, and $w$ is the dredge width in meters. The apparent density, $a d$, of oysters on the bottom for a given tow was calculated as
(2) $a d=\left(o^{*} b\right) / s a$
where $o$ is the total number of oysters present in the bushel sample from a given tow and $b$ is the total number of bushels caught in that tow. The correct density, $c d$, of oysters on the bottom for a given tow was calculated as

$$
\begin{equation*}
c d=\left(\sum^{n=6} o d\right) /(n * t a) \tag{3}
\end{equation*}
$$

where od is the total number of oysters collected by patent tong or diver sample taken along a given tow path, $n$ is the total number of patent tong or diver samples taken along that same tow path, and $t a$ is the area sampled by the patent tong or diver quadrat. Oyster dredge capture efficiency, $e$, was calculated as
(4) $e=a d / c d$.

Preliminary evaluation of $e$ suggested that at values of $c d$ below 25 oysters $/ \mathrm{m}^{2}, e$ was commonly estimated to be greater than 1 and sometimes greater than 2 . Since the tong/diver samples cover a much smaller spatial area than the dredge, the low abundance at these locations likely lead to tong/diver undersampling bias (Powell et al. 2017) and therefore an overestimate of capture efficiency. Therefore, I did not include any capture efficiency estimates in subsequent analyses that were derived from $c d$ observations below 25 oysters $/ \mathrm{m}^{2}$.

Finally, catchability, $q$, was calculated as
(5) $q=1 / e$.

## Evaluating size-specific retention probability

I evaluated the size-dependent retention probability of the survey dredge using a logistic model that was fit to selectivity at a given length $s(1)$ calculated as
(6) $s(l)=D_{l, d} /\left(D_{l, t}+D_{l, d}\right)$,
where $\mathrm{D}_{l, d}$ was the experiment-wide density of oysters at a given length collected in the dredge and $\mathrm{D}_{l, t}$ was the experiment-wide density of oysters at that same length
collected in the tong/diver samples. Given the results of this analysis (no size selectivity), I used the full range of sizes for oysters in all further analyses.

## Evaluating spatial and temporal variation in capture efficiency

Logistic regression was applied to assess spatial and temporal effects on the capture efficiency of the survey dredge. To evaluate temporal variability, the dataset was truncated to include only locations that were sampled in all years. Given the results of this analysis (no temporal variability), the decision was made to include data from all years in subsequent spatial analyses.

Spatial analyses identified three statistically significant groups of oyster reef locations relative to capture efficiency. Additionally, these groups were different from those identified in the 2003 experiment (Powell et al. 2007) which have been applied in the stock assessment since 2005 (Ashton-Alcox et al. 2016). To evaluate the impact of changing the capture efficiency of the survey gear, and the catchability coefficients derived from capture efficiency, on the stock assessment, mean catchability coefficients were derived separately from capture efficiency based on analyses conducted by Powell et al. (2007) and on analyses conducted with this work. Powell et al. (2007) analyzed capture efficiency separately for different-sized oysters and while they found no change over time in capture efficiency for all oysters, a significant effect was detected for market-sized oysters. Therefore, time-specific catchability coefficients have been applied in the stock assessment and the most recent part of time series uses regional mean catchability coefficients derived from data collected during the 2003 experiment only to convert CPUE to abundance. However, since I detected no size selectivity of the dredge, and no temporal trend in capture efficiency, I derived regional mean
catchability coefficients from the full set of experimental data (1999, 2000, 2003, and 2013) and calculated regional means based on the statistically significant reef groupings from this work. The proportional change in abundance or exploitation rate $P$ from the previously applied catchability coefficients for the stock assessment (Powell et al. 2007; Ashton-Alcox 2015) to the application identified as most appropriate with this work was calculated as
(7) $\quad P=\left[\left(\delta_{\text {new }}-\delta_{\text {old }}\right) / \delta_{\text {old }}\right] * 100$
where $\delta$ represents abundance or exploitation rate, new represents the application of catchability coefficients identified with this work as most appropriate and old represents the application of catchability coefficients that have been used in the stock assessment (Ashton-Alcox 2015), based on previous analyses by Powell et al. (2007).

Estimating uncertainty in capture efficiency and modeling densitydependence

Uncertainty in the estimate of $e$ can be defined by the variation in intra-tong/diver quadrat density used to estimate $c d$ for a given sample. For each estimate of $e$, six tong density measurements or twelve quadrat density measurements were drawn at random with replacement from the set of observed densities and $c d$ and $e$ were recalculated using these random draws. This was repeated 1,000 times in a bootstrap simulation and the variance, $\sigma^{2}$, of the simulated distribution of $e$ was used subsequently as a measure of uncertainty.

A power function is commonly used to describe density-dependence in catchability for many fisheries (Wilberg et al. 2010) and has been used to describe densitydependence in survey capture efficiency for trawl surveys in at least one instance
(Kotwicki et al. 2014). Therefore, to determine whether $e$ of the survey dredge was density-dependent here, I modeled $e$ as a power function of $c d$

$$
\text { (8) } \quad e \sim \alpha^{*} c d^{-\phi},
$$

where $\alpha$ and $\phi$ are model parameters, using weighted non-linear least squares regression. The weights $w t$ for each observation of $e$ were the reciprocal the $\sigma^{2}$
(9) $\quad w t=1 / \sigma^{2}$
where $\sigma^{2}$ was estimated by bootstrap simulation as described above.

## Results

The patent tongs and diver samples, assumed to have collected $100 \%$ of the oysters in the sampled area, had the same size frequency distribution as the commercial oyster dredge (Figure 2). Hence, oyster length was not a significant ( $\mathrm{p}=0.82$ ) predictor of retention probability and all subsequent analyses were conducted without separating oysters by size class or correcting for size-specific retention probability.

When data were truncated to include only experiments done at reefs sampled in all three years, year was not a significant predictor of capture efficiency ( $p=0.29$ ), and neither was the interaction between year and reef $(p=0.44)$, but reef was significant ( $p$ $=0.003$ ). Capture efficiency was nearly constant across all three years sampled (Figure $3)$.

For the full set of data, reef $(p=0.004)$ remained a significant predictor of capture efficiency. Efficiency is more variable, and higher in the lower bay, decreases and becomes less variable in the middle section of the bay, and increases in the upper bay (Figure 4). Reefs formed statistically significant groups relative to capture efficiency
and these groups were different from those estimated by Powell et al. (2007), who used data collected during the set of experiments conducted in 2003 only.

When updated location-specific catchability coefficients that included data from all experiments (1999, 2000, 2003, and 2013) were applied to calculate 2015 abundance and exploitation rate, the proportional change varied with region (Table 1). For instance, on the Very Low Mortality region and on the Round Island reef in the Low Mortality region, the estimated abundance of oysters decreased by 67\%; however, on the Shell Rock region, the estimated abundance of oysters increased by $166 \%$ and the estimated exploitation rate decreased by 62\%. Overall, the 2015 estimated abundance decreased by $9 \%$ and the exploitation rate increased by $11 \%$.

Uncertainty in estimates of capture efficiency increased with decreasing oyster density (Figure 5). Oyster density significantly influenced the capture efficiency of the survey dredge. A two-parameter power function described the relationship between capture efficiency and oyster density (Table 2). Capture efficiency declined with increasing oyster density but at a slower rate at oyster densities greater than approximately 100 oysters/m² (Figure 5).

## Discussion

## Density-dependent capture efficiency

Density-dependent capture efficiency or catchability, and the hyperstability/hyperdepletion of CPUE that results from density-dependence, are typically thought to be processes that make fishery catch data disproportionate to abundance because of the interaction between organism and fisher behavior (Hilborn
and Walters 1992). CPUE data collected from fishery-independent surveys, however, were traditionally thought to provide reliable indices of abundance because survey design is standardized (Godo 1994). More recently, though, evidence for densitydependent catchability in fishery-independent bottom-trawl surveys has been described as well (Godo et al. 1999; Kotwicki et al. 2013, 2014). To my knowledge this is the first time density-dependent capture efficiency has been identified and modeled for a sessile invertebrate stock survey. For bottom-trawl fishery-independent surveys, density-dependence likely results from fish reacting, such as schooling or gear avoidance behaviors at high density, to the survey gear (Godo et al. 1999; Hoffman et al. 2009). Oysters are sessile and immobile, so density-dependent capture efficiency is not manifested in behavioral responses to survey gear. For oysters, density dependence may be explained by their gregarious setting behavior and the three-dimensional structure of reefs that become consolidated and cemented at high density. When oyster density is low, the dredge is likely to capture most oysters in the dredge path, but as density increases, the vertical relief of the reef, cementation, and consolidation likely increase as well making it more difficult for the dredge to capture $100 \%$ of the oysters in the dredge path.

Since the efficiency of the survey gear evaluated here is density-dependent, one must caution against biased interpretation of survey data due to the likelihood of hyperstability in survey CPUE. Hyperstability describes a situation where CPUE remains stable while true density or abundance declines (Hilborn and Walters 1992). Density-dependence in capture efficiency and catchability have been shown to cause hyperstability in CPUE for fisheries and for fishery-independent surveys (Hilborn and

Walters 1992; Rose and Kulka 1999; Godo et al. 1999; Wilberg et al. 2010; Ward et al. 2013; Kotwicki et al. 2013, 2014). As true density or abundance declines, if the efficiency of a fishing fleet or of a survey gear increases, CPUE will remain stable. For the oyster population in Delaware Bay this means increases or declines in oyster density are likely to be difficult to detect, especially at low densities, because capture efficiency increases rapidly with declining density. Interpreting temporal trends from survey CPUE may be particularly problematic for reefs in the lower bay where heavy disease pressure keeps oyster density low.

Interestingly, above 100 oysters $/ \mathrm{m}^{2}$ the capture efficiency declines at a much lower rate. Since density-dependent capture efficiency of survey gear has not been modeled for other sessile invertebrates it is difficult to speculate on the broader ecological significance of this inflection point. However, it is possible that much of the change in density above 100 oysters $/ \mathrm{m}^{2}$ comes from recruitment of small oysters that, due to limitations on space and resources within a square meter of habitat, do not survive to larger sizes. If this is the case, then these oysters would not contribute substantially to increasing vertical relief of the reef, cementation, and consolidation, which I suggest might be influencing the capture efficiency of the survey gear. As similar information becomes available for oyster reefs in different regions with different resource availability, it will be informative to compare how parameters of the power model I used to model capture efficiency vary.

Size-selectivity and spatiotemporal variability in capture efficiency
In many cases, the size of the organism being surveyed provides an accurate estimate of capture probability because survey and fishing gears are designed to retain
individuals above a certain size threshold (Millar 1992). The lack of any size selectivity here, however, is in agreement with previous work that evaluated capture efficiency separately for different sized oysters (Powell et al. 2007; Marenghi et al. 2017). Sizedependent retention should be rare because oysters grow in aggregations with the larvae cementing onto existing oyster substrate.

The probability that a given clump of oysters will be retained by the dredge is therefore a function of the size of the entire clump and not of the size of any one oyster in that clump.

No temporal change in capture efficiency was detected, but reefs formed statistically significant regional groups within Delaware Bay along the salinity gradient in the lower, middle, and upper bay. The same was true when an earlier subset of these data were evaluated (Powell et al. 2007), though the specific reefs in each group changed slightly. Since I have identified a strong density-dependence in capture efficiency, this suggests that density at each of the sampled locations has changed relatively little from 1999, when the first experiment was conducted, to 2013, when the last experiment was conducted. In fact, relative to the long-term time series, the oyster population in Delaware Bay, NJ has been stable over this time period (Ashton-Alcox 2016). While spatial and temporal variability in survey gear capture efficiency can create biased estimates of abundance and exploitation rate (Pope and Shepherd 1985; Wilberg and Bence 2006), progress continues to be made on innovative ways to deal with both (Wilberg et al. 2010; Cadrin et al. 2016), and the region-specific mean catchability coefficients being applied in the stock assessment of Delaware Bay oysters appear to adequately account for the spatial variation in oyster density and catchability.

However, as has been recommended by others (Powell et al. 2007; Marenghi et al. 2017), periodic re-evalution of spatial patterns in density is advised to detect and correct for any temporal changes in survey gear catchability. Alternatively, coupled acoustic methods have the potential to accurately map different forms of oyster habitat (Allen et al. 2005; Legare and Mace 2017). Advancement in these and other acoustic methods may allow oyster density estimation without the need to correct for capture efficiency or catchability of dredge survey gears.

## Stock assessment implications for updating catchability coefficients

The catchability coefficients derived from the 2003 experimental data using statistically significant regional group means (Powell et al. 2007), herein referred to as old- $q$, differed from those derived from the present analysis, herein referred to as newq. Given that the exploitation rate is fixed at the number of oysters harvested divided by the estimated population size at that time, the only variables that can change from updating catchability coefficients from old-q to new-q are the estimate of what the population size is and/or the estimate of how heavily the population is being exploited. For instance, on the Shell Rock region if the total number of oysters harvested was 5 million, it remains 5 million whether the exploitation rate (Table 1) was calculated to be 0.069 at a population size of 78 million oysters or 0.026 at a population size of 208 million oysters. Therefore, the management implications for updating to new- $q$ values may appear inconsequential because the harvest remains fixed. However, there is inherent value in knowing, with as much certainty as possible, both the actual abundance of the population on which the fishery is prosecuted, and how heavily that population is being exploited. This is because sustainable rates of exploitation for $C$.
virginica populations in one system, e.g. Delaware Bay, can influence how a fishery is managed in another system. In the case of Shell Rock in particular, a 7\% exploitation rate would appear suitable to sustain the population there based on old- $q$ values, however, the exploitation rate there is much closer to $2.5 \%$, based on new- $q$ values. Given that recent simulation modeling suggests oyster populations can sustain exploitation rates of approximately $2 \%$, and that this is robust across different systems and fisheries (Powell et al. in press), the 7\% exploitation rate was likely artificially high and an artifact of a misspecified catchability coefficient. Hence, as has been identified in other fisheries (Pope and Shepherd 1985; Wilberg and Bence 2006), inaccurate catchability coefficients can lead to biased estimates of abundance and exploitation rate in oyster fisheries.

Applying new-q values to 2015 stock assessment survey data resulted in a 9\% reduction in total abundance and an $11 \%$ increase in exploitation rate. However, it is important to recognize that the change in catchability is not unidirectional for all regions evaluated; whether the effect of the new- $q$ values results in an increase or a decrease in total stock size depends on the relative contribution of any one region to the total stock size in any given year. For years and regions where catchability is now estimated to be higher and the region makes up a larger relative portion of the total stock, the result will be an overall increase in stockwide abundance, and associated decrease in exploitation rate, not an increase.

## Acknowledgements

I thank Russell Babb, Jason Hearon, and Craig Tomlin of the New Jersey Division of Fish and Wildlife Bureau of Shellfisheries and Jim Wesson, Vernon Rowe, and Kyle Jones of the Virginia Marine Resources Commission for vessel support and field assistance. I am also grateful to the staff at the Haskin Shellfish Research Laboratory for helping in the field and processing samples in the lab. Funding for this work was provided by the Army Corp of Engineers.

## Literature Cited - Chapter 3

Allen, Y. C., C. A. Wilson, H. H. Roberts, and J. Supan. 2005. High resolution mapping and classification of oyster habitats in nearshore Louisiana using sidescan sonar. Estuaries 28 (3): 4350446.

Arreguin-Sanchez, F. 1996. Catchability: a key parameter for fish stock assessment. Reviews in Fish Biology and Fisheries 6 (2): 221-242.

Ashton-Alcox, K., D. Bushek, J. Gius, J. Morson, and D. Munroe. 2015. Report of the 2014 Stock Assessment Workshop (14th SAW) for the New Jersey Delaware Bay Oyster Beds. Haskin Shellfish Research Laboratory, Port Norris, NJ. 112pp.

Ashton-Alcox, K., D. Bushek, J. Gius, J. Morson, and D. Munroe. 2016. Report of the 2015 Stock Assessment Workshop (14th SAW) for the New Jersey Delaware Bay Oyster Beds. Haskin Shellfish Research Laboratory, Port Norris, NJ. 158pp.

Cadrin, S. X., G. R. DeCelles, and D. Reid. 2016. Informing fishery assessment and management with field observations of selectivity and efficiency. Fisheries Research 184: 9-17

Chai, A., M. Homer, C. Tsai, and P. Goulletquer. 1992. Evaluation of oyster sampling efficiency of patent tongs and an oyster dredge. North American Journal of Fisheries Management 12: 825-832.

Doray, M., S. Mahevas, and V. M. Trenkel. 2010. Estimating gear efficiency in a combined acoustic and trawl survey, with reference to the spatial distribution of demersal fish. ICES Journal of Marine Science 67: 668-676.

Ellis, N. and Y.-G. Wang. 2007. Effects of fish density distribution and effort distribution on catchability. ICES Journal of Marine Science 64: 178-191.

Gedamke, T, W. D. DuPaul, and J. M. Hoenig. 2004. A spatially explicit open-ocean DeLury analysis to estimate gear efficiency in the dredge fishery for sea scallop Placopecten magellanicus. North American Journal of Fisheries Management 24(2): 335-351.

Godo, O. R. 1994. Factors affecting reliability of groundfish abundance estimates from bottom trawl surveys. In Marine fish behavior in capture and abundance estimation, pp. 45-68. Eds. A. Gerno and S. Olsen. Fishing News Books, Oxford.

Godo, O. R., S. J. Walsh, and A. Engas. 1999. Investigating density-dependent catchability in bottom-trawl surveys. ICES Journal of Marine Science 56: 292-298.

Hennen, D., L. Jacobson, and J. Tang. 2012. Accuracy of the patch model used to estimate density and capture efficiency in depletion experiments for sessile invertebrates and fish. ICES Journal of Marine Science 69:240-249.

Hilborn, R. and C. J. Walters. 1992. Quantitative fisheries stock assessment: choice, dynamics, and uncertainty. Chapman and Hall, New York, NY.

Hoffman, J. C., C. F. Boznek, and R. J. Latour. 2009. Estimation of bottom trawl catch efficiency for two demersal fishes, Atlantic croaker and white perch, in Chesapeake Bay. Marine and Coastal Fisheries: Dynamics, Management, and Ecosystem Science 1: 255269.

Kotwicki, S., A. De Robertis, J. N. Ianelli, A. E. Punt, and J. K. Horne. 2013. Combining bottom trawl and acoustic data to model acoustic dead zone correction and bottom trawl efficiency parameters for semi-pelagic species. Canadian Journal of Fisheries and Aquatic Sciences 70: 208-219.

Kotwicki, S., J. N. Ianelli, and A. E. Punt. 2014. Correcting density-dependent effects in abundance estimates from bottom-trawl surveys. ICES Journal of Marine Science 71(5): 1107-1116.

Latsa, M. L. and O. O. Iribarne. 1997. Southwestern Atlantic scallop (Zygochlamys patagonica) fishery: assessment of gear efficiency through a depletion experiment. Journal of Shellfish Research 16: 59-62.

Legare, B. and C. Mace. 2017. Mapping and classifying eastern oyster (Crassostrea virginica) habitat in Copano Bay, Texas, by coupling acoustic technologies. Journal of Coastal Research 33(2): 286-294.

Mann, R., M. Southworth, J. M. Harding, and J. Wesson. 2004. A comparison of dredge and patent tongs for estimation of oyster populations. Journal of Shellfish Research 23(2):287-390.

Marenghi, F., K. A. Alcox, R. Wong, B. Reynolds, and G. Ozbay. 2017. Dredge efficiency on natural oyster grounds in Delaware Bay and its application in monitoring the Eastern oyster (Crassostrea virginica) stock in Delaware Bay, USA. Fisheries Research 186: 292-300.

Millar, R. B. 1992. Estimating the size-selectivity of fishing gear by conditioning on the total catch. Journal of the American Statistical Association 87: 962-968.

Pope, J. G. and J. G. Shepherd. 1985. A comparison of the performance of various methods for tuning VPAs using effort data. ICES Journal of Marine Science 42 (2): 129151.

Powell, E. N., K. A. Ashton-Alcox, J. A. Dobarro, M. Cummings, and S. E. Banta. 2002. The inherent efficiency of oyster dredges in survey mode. Journal of Shellfish Research 21: 691-695.

Powell, E. N., K. A. Ashton-Alcox, and J. N. Kraeuter. 2007. Reevaluation of eastern oyster dredge efficiency in survey mode: application in stock assessment. North American Journal of Fisheries Management 27: 492-511.

Powell, E. N ., K.A. Ashton-Alcox, J.N. Kraeuter, S.E. Ford and D. Bushek. 2008. Long term trends in oyster population dynamics in Delaware Bay: Regime shifts and response to disease. Journal of Shellfish Research 27: 729-755.

Powell, E. N., R. Mann, K. A. Ashton-Alcox, K. M. Kuykendall, and M. Chase Long. 2017. Can we estimate molluscan abundance and biomass on the continental shelf? Estuaries, Coastal, and Shelf Sciences 198: 213-224.

Powell, E. N., E. E. Hofmann, and J. M. Klinck. In press. Oysters, sustainability, management models, and the world of reference points. Journal of Shellfish Research.

Rago, P.J., J.R. Weinberg, and C. Weidman. 2006. A spatial model to estimate gear efficiency and animal density from depletion experiments. Canadian Journal of Fisheries and Aquatic Science 63: 2377-2388.

Rose, G. A. and D. W. Kulka. 1999. Hyperaggregation of fish and fisheries: how catch-per-unit effort increased as the northern cod (Gadus morhua) declined. Canadian Journal of Fisheries and Aquatic Sciences 56: 118-127.

Rothschild, B. J., J. S. Ault, P. Goulletquer, and M. Heral. 1994. Decline in Chesapeake Bay oyster poplation: a century of habitat destruction and overfishing. Marine Ecology Press Series 111: 29-39.

Sing, W., E. B. Ornolfsdottir, and G. Stefansson. 2014. A small-scale comparison of Iceland scallop size distributions obtained from a camera based autonomous underwater vehicle and dredge survey. Plos One 9(10): 1-10.

Walters, C. J. and S. J. D. Martell. 2004. Fisheries ecology and management. Princeton University Press.

Ward, H. G. M., P. J. Askey, and J. R. Post. 2013. A mechanistic understanding of hyperstability in catch per unit effort and density-dependent catchability in a multistock recreational fishery. Canadian Journal of Fisheries and Aquatic Sciences 70 (10): 1542-1550.

Wilberg, M. W. and J. R. Bence. 2006. Performance of time-varying catchability estimators in statistical catch-at-age analysis. Canadian Journal of Fisheries and Aquatic Sciences 63: 2275-2285.

Wilberg. M. W., J. T. Thorson, B. C. Linton, and J. Berkson. 2010. Incorporating timevarying catchability into population dynamic stock assessment models. Reviews in Fisheries Science 18 (1): 7-24.

Wilberg, M. W., J. M. Robinson, S. A. M. Rains, J. L. Humphrey, and R. N. Lipcius. 2013. Effects of location errors on estimates of dredge catchability. Fisheries Research 148: 1-8.

## TABLES - CHAPTER 3

Table 3.1. Oyster abundance and exploitation rate calculated using different catchability coefficients for each of the managed regions in 2015 (See Figure 3.1). The proportional change in abundance from the catchability coefficients using 2003 data only, to the application identified with this work as being most appropriate (1999, 2000, 2003, and 2013 data), is given in parentheses next to each abundance estimate in the lower half of the table. * The Low Mortality region was split out into separate sections (Arnolds / Upper Arnolds and Round Island) based on the results of analysis for spatial trends in capture efficiency (see Results section and Figure 3.4).

|  | Experimental Data Used To |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
|  | Calculate Catchability |  | Exploitation | Catchability |
|  | Coefficient | Oyster Abundance | Rate | Coefficients |
| All | 2003 | 1,701,095,858 | 0.018 | - |
| Very Low Mortality | 2003 | 485,588,439 | 0 | 7.3 |
| Low Mortality* - Round Island | 2003 | 57,480,614 | 0 | 7.3 |
| Low Mortality* - Arnolds/Upper Arnolds | 2003 | 182,871,268 | 0.025 | 7.3 |
| Medium Mortality Transplant | 2003 | 412,269,327 | 0.024 | 7.3 |
| Medium Mortality Market | 2003 | 234,268,474 | 0.017 | 7.3 |
| Shell Rock | 2003 | 78,433,103 | 0.069 | 3.11 |
| High Mortality | 2003 | 250,184,633 | 0.03 | 3.11 |
| All | 1999, 2000, 2003, 2013 | 1,552,939,034 (-9\%) | 0.02 (11\%) | - |
| Very Low Mortality | 1999, 2000, 2003, 2013 | 160,310,704 (-67\%) | 0 (0\%) | 2.41 |
| Low Mortality* - Round Island | 1999, 2000, 2003, 2013 | 18,976,477 (-67\%) | 0 (0\%) | 2.41 |
| Low Mortality* - Arnolds/Upper Arnolds | 1999, 2000, 2003, 2013 | 206,920,093 (13\%) | 0.022 (-12\%) | 8.26 |
| Medium Mortality Market | 1999, 2000, 2003, 2013 | 466,485,568 (13\%) | 0.021 (-13\%) | 8.26 |
| Medium Mortality Transplant | 1999, 2000, 2003, 2013 | 265,076,384 (13\%) | 0.015 (-12\%) | 8.26 |
| Shell Rock | 1999, 2000, 2003, 2013 | 208,314,288 (166\%) | 0.026 (-62\%) | 8.26 |
| High Mortality | 1999, 2000, 2003, 2013 | 226,855,520 (-9\%) | 0.033 (10\%) | 2.82 |

Table 3.2. Power function model parameters describing density-dependent capture efficiency.

| Model | Parameter | Coefficient | Standard <br> error | T <br> statistic | P value |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Power function; density-dependence |  |  |  |  |  |
|  |  |  |  |  |  |
|  | $\alpha$ | 1.9104 | 0.3252 | 4.12 | 0.0013 |
| $\phi$ | 0.5190 | 0.0781 | 5.012 | $<0.0001$ |  |

## FIGURES - CHAPTER 3



Figure 3.1. Map of sampling locations.


Figure 3.2. Length frequency (A) and probability density length distribution (B) for oysters collected at the same locations with a commercial oyster dredge and patent tongs.


Figure 3.3. Box and whiskers plot of mean capture efficiency of a commercial oyster dredge estimated in 2000, 2003, and 2013 at the same sampling locations. Bold horizontal lines represent the mean, boxes encompass the interquartile range, black whiskers extend to the $5^{\text {th }}$ and $95^{\text {th }}$ percentiles, and dots are outliers.


Figure 3.4. Box and whiskers plot of mean capture efficiency of a commercial oyster dredge at different oyster reef locations along the Delaware Bay, USA. Reef locations are organized on the x -axis from the lower bay (left side) to the upper bay (right side). Empty black boxes around the reef names represent the regional grouping of bedspecific catchability coefficients applied in the 2015 Delaware Bay oyster stock assessment based on data collected from the 2003 experiments (Powell et al. 2007). Shaded, offset boxes, represent the regional groupings of bed-specific catchability coefficients identified as statistically appropriate with this work. Bold horizontal lines represent the mean, boxes encompass the interquartile range, black whiskers extend to the $5^{\text {th }}$ and $95^{\text {th }}$ percentiles, and dots are outliers.


Figure 3.5. Survey gear capture efficiency as a function of true oyster density. Error bars represent the standard deviation from 1,000 bootstrap simulations. Line indicates the best fit power model estimated by weighted nonlinear least squares. See methods section for details.

## CHAPTER 4

## ALTERNATIVE MODELS TO ESTIMATE DENSITY OF EASTERN OYSTERS CRASSOSTREA VIRGINICA USING A HYDRAULIC DREDGE


#### Abstract

Catchability $(q)$ is a parameter used in stock assessments to describe how much of the population is caught by a single unit of fishing or survey effort. In some cases, $q$ can be density-dependent, causing survey CPUE to be hyperstable across a range of animal densities. In these instances, CPUE can be an unreliable proxy for true density in the sampled area. In Delaware Bay, USA, there is a natural, along-bay gradient in the density of eastern oysters Crassostrea virginica. In addition, the $q$ of the survey dredge used in the assessment of the oyster population there is density-dependent. Using data collected during two separate field experiments, one in 2003 and one in 2013, I evaluated three potential methods for estimating true oyster density in an area sampled with an oyster dredge. For the first model, CPUE was corrected for by applying spatially-explicit $q$ coefficients that account for the along-bay gradient in density. In the second, CPUE was corrected for by estimating $q$ in situ for each tow using a logistic model fit to catch composition and tow covariates. For the third model, CPUE data were ignored entirely and a model that accounted for the proportion of the sample composition that was made up of oysters was applied to estimate oyster density in situ for each tow. Models were trained on one set of experimental data and tested on an independent set. The mean square error (MSE) was calculated for each train-test pairing and the modeling approach with the lowest average MSE score was chosen as


the best approach. The model that utilized the proportion of the total catch that was made up of oysters to estimate the oyster density performed best. Where time and financial constraints prevent a more quantitative approach that measures CPUE and estimates $q$ for the sampling gear, or where $q$ varies at fine spatial and temporal scales, a simpler evaluation of the relative components that comprise a tow may adequately describe the density of oysters in the sampled area.

## Introduction

Catchability $(q)$ is a parameter used in stock assessments to describe how much of the population is caught by a single unit of fishing or survey effort (Arreguin-Sanchez 1996). Fishery and survey $q$ can vary in space and time, sometimes due to changes in the density of the sampled organism (e.g., Rose and Kulka 1999; Godo et al. 1999; Kotwicki et al. 2014). In such instances, interpreting survey or fishery catch-per-unit effort (CPUE) is difficult because they are rarely proportional to the actual abundance (Hilborn and Walters 1992; Maunder et al. 2006).

The conditions that produce density-dependent $q$ in observations collected from fishing data are well established (Hilborn and Walters 1992; Rose and Kulka 1999). For example, as abundance declines, fish may aggregate in a behavioral response to declining abundance, and fishermen will likely aggregate around the fish, causing CPUE to remain constant while abundance declines. This makes abundance especially difficult to estimate from fishery-dependent data because it can result in fishery CPUE being hyperstable at different levels of abundance (Erisman et al. 2011).

Evidence for density-dependent $q$ is now becoming more prominent in fisheryindependent survey data as well (Godo et al. 1999; Kotwicki et al. 2013, 2014). While the behavior of the survey gear may be standardized, reducing or eliminating the influence of human behavior, the surveyed animal may still respond to that gear differently at varying densities (Kotwicki et al. 2014). This can make relative abundance or CPUE indices generated from a fishery-independent survey unreliable as well.

The $q$ for commercial dredges used to survey oyster reefs in Delaware Bay varies spatially and temporally (Powell et al. 2002, 2007) likely due to changes in oyster density (Chapter 3). Furthermore, density-dependent $q$ leads to survey CPUE being hyperstable at low oyster density (Chapter 3). This raises the question then, how does one confidently estimate the density of oysters in an area sampled with a dredge?

Highly variable dredge $q$ has been identified for other oyster populations (Fu et al. 2016; Marenghi et al. 2017). One recommended course of action to account for this is to treat oyster resource surveys as relative indices with prior probabilities that incorporate uncertainty in dredge efficiency (Fu et al. 2016). However, this approach generates highly uncertain abundance estimates, making it difficult to interpret how an oyster resource is responding to prescribed management actions or environmental drivers (Ashton-Alcox 2016).

In this chapter, given the density-dependence in $q$ of the survey gear used to assess the oyster population in Delaware Bay, described in Chapter 3, I evaluate the accuracy of three different approaches to estimating true oyster density $(t d)$ in the sampled area from a survey dredge tow. In the first approach, herein referred to as the average
model (AM), the estimated density (ed) was calculated by multiplying the sample CPUE by a region-specific, mean $q$. The AM approach is what is used to calculate ed in the sampled area for the stock assessment of oysters in Delaware Bay (Ashton-Alcox 2016) and is supported by statistical analyses of the spatial trend in oyster density along the bay gradient (Powell et al. 2002, 2007; Chapter 3). However, the AM approach depends on having an independent, experimentally-derived estimate of $q$, and since $q$ is densitydependent (Chapter 3), this approach requires $q$ estimates be periodically updated to account for any temporal changes in density in a given region.

In the second approach, herein referred to as the logistic model (LM), a tow-specific $q$ was modeled by logistic regression using a list of tow and catch composition variables, and applied to CPUE to calculate ed. When catch composition and variables that describe the conditions of the tow explain some of the variability in $q$, this information can potentially be used to estimate $q$ in situ for each individual tow (Powell et al. 2007). Since the LM approach estimates catchability directly for each tow in situ, this method does not require periodic reassessment of $q$.

The third approach evaluated, herein referred to as the power model (PM), does not utilize survey CPUE data, but rather models ed in the sampled area directly as a power function of the proportion of the survey dredge tow that was made up of oysters. Since the PM approach relies only on an estimate of the contents in a given tow, it does not require an estimate of CPUE for the tow or independent estimates of $q$. This approach is akin to an historic management rule called the " $40 \%$ rule" used in the management of the Delaware Bay oyster fishery from 1953 to 1996, before $q$ had been estimated. Under
this rule, if the average survey dredge catch in a given area was comprised of less than 40\% oysters, the fishery in that area would close (Ford 1997).

## Material and Methods

## Field sampling

Survey dredge CPUE and true oyster density, $t d$, were estimated at twenty-four locations in Delaware Bay in 2003 and 2013 (Figure 4.1). Detailed protocol for estimating CPUE and $t d$ can be found in Chapter 3, but briefly, the F/V Howard $W$. Sockwell was used to tow the survey dredge for 1-minute at each sampling location. If the dredge was full after the 1-minute tow, the tow time was reduced to 45 s so that the total number of bushels of material collected in the swept area could be determined. Swept area and an estimate of the total number of oysters caught in the dredge were used to calculate CPUE as oysters $/ \mathrm{m}^{2}$ for each tow. To estimate $t d$, six, $0.89 \mathrm{~m}^{2}$ patent tong grabs, assumed to be $100 \%$ efficient, were collected parallel to, but not inside, the tow track and $t d$ was estimated as total oysters $/ \mathrm{m}^{2}$ from these six grabs.

For each dredge tow, a one bushel subsample was collected, brought back to the lab, and sorted into catch component classes of oysters, cultch (oyster shell), boxes (dead oysters with the hinge still intact), and debris. Volumes and weights of each catch component were recorded and all oysters and boxes in each sample were counted and individual lengths measured. In addition, wire scope, swept area, total catch in bushels, and change in bottom depth were recorded for each tow.

For each tow, this approach resulted in an observation of survey dredge CPUE (or apparent density (ad)), an estimate of $t d$ adjacent to the survey tow, and a record of tow
and catch composition data that could be evaluated for their effect on survey dredge performance.

Data Analysis - Model Descriptions
Average Model (AM)
The model estimated density $e d_{t_{r_{i}}}$ for each test data set $t$, region $r$, and tow $i$ was

$$
\begin{equation*}
e d_{t_{r_{i}}}=q_{t r_{r}} * a d_{t_{r_{i}}} \tag{1}
\end{equation*}
$$

where for each tow $i$, the apparent density, or CPUE, $a d_{i}$ of oysters in the tow track was

$$
\begin{equation*}
a d_{i}=\frac{o p b_{i} * b_{i}}{s a_{i}} \tag{2}
\end{equation*}
$$

where $o p b$ represents the number of oysters collected in a 37-L bushel subsample, $b$ represents the total number of bushels in the tow, and sa represents the swept area $\left(\mathrm{m}^{2}\right)$ covered by the tow. The mean catchability parameter $q_{t r_{r}}$ for each training data set $t r$ and region $r$ was

$$
\begin{equation*}
q_{t r_{r}}=\frac{1}{n} \sum_{i=1}^{n}\left(\frac{1}{e_{i_{t r_{r}}}}\right) \tag{3}
\end{equation*}
$$

where for each tow $i$

$$
\begin{equation*}
e_{i}=\frac{a d_{i}}{t d_{i}} \tag{4}
\end{equation*}
$$

Finally, the density of oysters $t d_{i}$ collected parallel to tow track $i$ was

$$
\begin{equation*}
t d_{i}=\frac{\sum_{j=1}^{6} o_{j}}{5.34} \tag{5}
\end{equation*}
$$

where $j$ represents a $0.89 \mathrm{~m}^{2}$ tong sample collected parallel to tow $i$ and $o$ represents the number of oysters collected in each tong sample $j$.

Logistic Model (LM)

Powell et al. (2007) used data from the 2003 experiment to evaluate whether information collected during the tow, information from the catch composition, and information about the fishing intensity in the sampled area could be used to model towspecific $q$ of the dredge in situ. This approach is attractive because it does not rely on having independent observations of $q$ or require periodic reevaluation of $q$. In addition, it takes advantage of the available information that describes the tow and catch conditions. Finally, it allows for $q$ to be estimated at the individual tow level, instead of being a fixed, region-specific coefficient. With the LM approach, I build on this previous work by reevaluating the influence of the same set variables used in the Powell et al. (2007) analysis, but by including data from the 2013 experiment as well, by including additional independent catch and tow composition variables, and by altering the modeling approach from estimating $q$ with linear multiple regression to estimating the probability of capture $e$ with multiple logistic regression.

In many cases, the size of the organism being surveyed provides an accurate estimate of capture probability because survey and fishing gears are designed to retain individuals above a certain size threshold (Millar 1992). However, this is not true for oysters (Powell et al. 2007; Marenghi et al. 2017; Chapter 3), likely because oysters grow in interconnected reefs and the probability that a given clump of oysters will be retained by the dredge is a function of the size of the entire clump and not of the size of any one oyster in that clump. For this reason, the mean length of oysters caught in the dredge tow was not included as a potential variable in the LM analyses. Catch composition variables that were evaluated included the proportion of the 37-L bushel sub-sample that was made up of cultch, the area (length*width in mm ) of the ten largest
clumps in each sub-sample, where clump refers to a particle of material in the subsample, and the total haul volume (in bushels). Variables evaluated that were related to the tow included the change in depth from the beginning to the end of the tow, the wire scope, calculated as the amount of chain let out by the boat divided by the average depth for the tow, the total swept area covered, and a variable that measures the relationship between the direction of the tow and the direction and intensity of the tide, herein referred to as the tow-tide variable. The tow-tide variable $t t_{i}$ for each tow $i$ was calculated as

$$
\begin{equation*}
t t_{i}=\operatorname{dir}_{i}{ }^{*} t i d e_{i}, \tag{6}
\end{equation*}
$$

where the adjusted tow direction $\operatorname{dir}_{i}$ for tow $i$ was

$$
\begin{equation*}
d i r_{i}=45-t d i r_{i} \tag{7}
\end{equation*}
$$

for actual tow direction $t d i r_{i}<136$, and

$$
\begin{equation*}
d i r_{i}=t d i r_{i}-225 \tag{8}
\end{equation*}
$$

for $136 \leq t d i r_{i} \leq 315$, and

$$
\begin{equation*}
d i r_{i}=405-t d i r_{i} \tag{9}
\end{equation*}
$$

for $t d i r_{i}>315$. The magnitude of the tide $t i d e_{i}$ for equation (6) was calculated as

$$
\text { tide }_{i}=T_{i}-6,(10)
$$

for ebbing tides, and

$$
\begin{equation*}
\text { tide }_{i}=T_{i} \tag{11}
\end{equation*}
$$

for flooding tides, where $T_{i}$ represents the time in hours since slack tide at the time and location of tow $i$. The final variable evaluated was weighted industry coverage in the
sampled area. This was estimated as a weighted average of the fishing effort on the reef from the three years prior to when a given tow was conducted.

For each test data set $t$, the estimated density $e d_{t_{i}}$ was calculated separately for each tow $i$. This transforms equation (1) to

$$
\begin{equation*}
e d_{t_{i}}=q_{t_{i}} * a d_{t_{i}} \tag{12}
\end{equation*}
$$

where apparent density $a d_{t_{i}}$, or CPUE, is still estimated by equation (2), but where $q_{t_{i}}$ is now estimated separately for each individual tow $i$ and test data set $t$ in situ by

$$
\begin{equation*}
q_{t_{i}}=\frac{1}{e_{t_{i}}} \tag{13}
\end{equation*}
$$

and

$$
\begin{equation*}
e_{t_{i}}=\left[\frac{\exp \left(\beta_{0_{t r}}+\beta_{1 t r} x_{1 t_{i}}+\beta_{2 t r} x_{2 t} \ldots \beta_{j_{i}} \beta_{t r} x_{t_{i}}\right)}{1+\exp \left(\beta_{0}{ }_{t r}+\beta_{1 t r} x_{1 t_{i}}+\beta_{2 t r}{ }_{t r} x_{t_{t}} \ldots \beta_{j_{t r}} x_{j_{t}}\right)}\right], \tag{14}
\end{equation*}
$$

where $\beta_{0_{t}}$ is the estimated y-intercept parameter for test data set $t, x_{j_{t}}$ is a vector of catch composition, tow, and industry effort variables, as described above, $j$ for test data set $t$, and $\beta_{j_{t r}}$ is a vector of $j$ estimated parameters for each $x_{t r}$ in training data set $t r$. For each training data set $t r$ the best set of variables for vector $x_{j_{t}}$ was estimated using backward, stepwise Akaike Information Criteria

## Power Model (PM)

A preliminary analysis of oyster density across different variables measured during the field experiments revealed that oyster density increased in a non-linear, predictable fashion, with the proportion of the 37-L bushel sub-sample from each tow that was
made up of oysters. Therefore, the oyster volume relative to the total volume $o v_{i}$ in the bushel sub-sample was used to calculate the estimated oyster density $e d_{t_{i}}$ for each test data set $t$ in the sampled area for tow $i$ directly with

$$
\begin{equation*}
e d_{t_{i}}=\alpha_{t r} * o v_{t_{i}}{ }^{\theta_{t r}} \tag{15}
\end{equation*}
$$

where $\alpha_{t r}$ and $\theta_{t r}$ are estimated parameters for training data set $t r$.

## Data Analysis - Evaluating Model Performance

To evaluate the accuracy of each approach in estimating $t d$ in the sampled area, I split the experimental data up six ways, creating three pairs of training and test data sets. In the first pairing, the 2003 experimental data served as training data and the 2013 experimental data served as test data. In this case, the 2003 experimental data were used to estimate the parameters of the models. The models parameterized with the 2003 data were then used calculate $e d$ in the sampled area for the 2013 data and this model-estimated density was evaluated against $t d$ for the 2013 data. For a second pairing, this process was repeated using the 2013 data as training data and the 2003 experimental data as test data. For the final pairing, 24 of the 48 observations (including data from both the 2003 and 2013 experiments), were drawn at random and labeled as the training data set, while the remaining 24 observations were labeled as the test data set. For each training-test combination, the mean squared error $M S E_{m}$ for each model $m$ was calculated as,

$$
\begin{equation*}
M S E_{m}=\frac{1}{n} \sum_{i=1}^{n}\left(\widehat{t d}_{i}-e d d_{m_{i}}\right)^{2} \tag{16}
\end{equation*}
$$

where $\widehat{t d_{l}}$ represents what was estimated to be the true density in the sampled area from the tongs for tow $i$ and $e d_{m_{i}}$ represents the density estimated by model $m$ for tow
i. The approach with lowest average MSE score across all three training-test data combinations was selected as the best modeling approach.

## Results

The parameters for each modeling approach varied across the three training data sets (Table 4.1). For the AM approach, mean $q$ coefficients calculated from experiments conducted in the lower bay were always lower than those from experiments conducted in the upper bay and $q$ varied in the upper bay across the different training data sets.

For the LM approach, the variables that contributed significantly to the capture efficiency ( $e$ ) varied with the training data set (Table 4.1). Only two variables, total haul volume and the proportion of the subsample made up of cultch, were significant predictors of $e$ for all three training data sets. A higher proportion cultch in the subsample and a higher total catch volume resulted in a higher capture efficiency, and this was consistent across all three training data sets (Figure 4.2).

The PM approach achieved the lowest mean MSE score and outperformed all other models in all but one pairing, the AM 2003 training/2013 test data combination (Table 4.2). The mean MSE score for the AM approach was inflated by a high 2013 training/2003 test pairing MSE score (Table 4.2). The LM approach achieved a consistently poorer fit than either the AM or PM approach. The LM 2003 training/2013 test pairing, in particular, resulted in an order of magnitude poorer fit $(131,440)$ than any training-test pairing of any of the three modeling approaches. The LM approach typically produced large overestimations of $e d$ relative to $t d$, while the AM approach produced a disproportionate number of underestimates (Figure 4.3).

The $t d$ in the sampled area increased with an increase in the proportion of the bushel subsample that was made up of oysters (Figure 4.4). PM parameters estimated from a random draw of observations from the 2003 and 2013 experiments performed best in estimating the $t d$ in the sampled area (Table 4.2, Figure 4.4).

## Discussion

Fishery-independent surveys of natural resources are historically believed to provide indices that are proportional to abundance (Godo 1994; Kotwicki et al. 2014) and these indices are utilized regularly in stock assessments and models of population and community dynamics. However, we know now that even when collected using a standardized, fishery-independent survey design, CPUE may still not be proportional to abundance because $q$ of the survey gear can be highly variable in space and time (Hoffman et al. 2009; Wilberg et al. 2010; Kotwicki et al. 2014; Chapter 3).

Density-dependence in survey $q$ offers an interesting challenge because to convert CPUE or $a d$ in the sampled area to $t d$, one would need to know $t d$ a priori since $q$ is density-dependent. However, with an estimate of the $t d$ in the sampled area, there would be no need for $q$ or $a d$. Since oyster $t d$ cannot be estimated directly in an area sampled with an oyster dredge because the dredge is not $100 \%$ efficient (Powell et al. 2003, 2007; Marenghi et al. 2017), and since the capture efficiency of the dredge varies with density (Chapter 3), with this work I evaluated three alternative approaches to applying a singular $q$ coefficient to estimate $t d$ of oysters in an area sampled with a dredge. In the first, I attempted to control for density-dependence in $q$ in a direct way with spatially-explicit $q$ coefficients (the AM model). In the second I use an indirect
application to estimate $q$ for each tow in situ using available information about the tow and catch composition (the LM model). In the final application, I ignored CPUE data all together and adapted a version of a historically-applied method to estimate $t d$ in situ from the proportion of the bushel sub-sample that was made up of oysters (the PM model).

I expected the LM model to perform best because it utilized a suite of variables that described the tow conditions and catch composition, as well as the exploitation history in the sampled area, to estimate $q$. This fine scale information, however detailed, turned out to be uninformative relative to the information contained in the competing models. Not only did the LM approach not perform best, but it performed significantly worse than either of the alternative approaches. Tow and environmental conditions, as well as catch composition, are known to influence survey catchability for marine and freshwater species with a variety of life histories (Hoffman et al. 2009; Somerton et al. 2013; Sagarese et al. 2016; Korman and Yard 2017), and this is true for surveys of oyster reefs as well (Powell et al. 2007). However, since $q$ is density-dependent, and since neither density nor some proxy for density are included as independent variables in the LM approach, perhaps the poor performance of this method relative to the other two should not have been surprising.

The AM model, which outperformed the LM model by a large margin, also did not include density as a model parameter, however, by utilizing regional mean $q$ coefficients for a population where density varies in a predictable way along the bay gradient, this model did contain a proxy for density. The regional $q$ coefficients could be viewed as pseudo-density-dependent since density varies by region. While the
application of spatially-explicit $q$ coefficients appear to be rare in the assessment of oyster populations (but see Ashton-Alcox 2016), this approach likely has widespread application to other dredge surveys where oyster density occurs non-randomly in space.

Since CPUE is not proportional to true abundance when $q$ is density-dependent, it should probably not be surprising that the one application that ignores CPUE data entirely, the PM model, performed best. While one could devise a situation where this model would generate highly biased estimates of $t d$, such situations occurred at low enough frequency that the mean MSE for the PM model was lower than either of the other two applications evaluated. For instance, if a given tow covered a $100 \mathrm{~m}^{2}$ area and the only thing the dredge picked up over that tow was 1 oyster, the apparent density would be 0.01 oysters $/ \mathrm{m}^{2}$ and the proportion of the sample made up of oysters would be $100 \%$. Depending on the training data set used in the estimation, this would have resulted in an ed of between 142 and 362 oysters $/ \mathrm{m}^{2}$. This estimate would almost certainly be biased extremely high given that a single oyster was collected over a 100 $\mathrm{m}^{2}$ area.

To my knowledge, there are no other instances where a PM approach, or even something similar, has been used to estimate oyster density for a survey. The Delaware Bay oyster fishery used to be managed on a single reference point. If $40 \%$ or more of the average catch was made up of oysters, then the fishery in that area would remain open, but if the catch composition fell below $40 \%$ oysters, it would close (Ford 1997). For all four PM models this suggests $40 \%$ and lower would have equated to 50 oysters $/ \mathrm{m}^{2}$ and lower. While the " $40 \%$ rule" was a relative index of the catch
composition, perhaps it worked well because it provided and accurate and quantitative proxy for when $t d$ fell below the 50 oysters $/ \mathrm{m}^{2}$ threshold.

Where $q$ varies at fine spatial and temporal scales, the PM model may offer advantages over more traditional applications for estimating oyster density in an area sampled with a dredge. First, a significant amount of additional time and effort is needed to quantify both the swath area and the number of oysters for a given tow to calculate CPUE. Since both the AM and LM models require a measure of CPUE, and since both performed more poorly relative to the PM method, the dedicated time and resources spent to estimate CPUE may not be necessary. In addition, while the AM model performed nearly as well as the PM model, it requires some independent and up-to-date estimate of $t d$ over a coarse region. With increasing spatial shifts in density along the bay gradient over time, the application of the AM model will become progressively poorer. Since the PM model does not require independent knowledge of $t d$, similar shifts in density along the bay gradient should not affect the accuracy of the AM approach.

## Acknowledgements

I thank the personnel at the New Jersey Division of Fish and Wildlife Bureau of Shellfisheries, especially, Russell Babb, Jason Hearon, and Craig Tomlin for vessel support and for assisting with the field experiments. I am also grateful to the staff at the Haskin Shellfish Research Laboratory for helping in the field and for helping to sort and process samples in the lab. Finally, I would like to thank the members of the Delaware

Bay Oyster Stock Assessment Review Committee for providing helpful comments on the analyses presented herein.

## Literature Cited - Chapter 4

Arreguin-Sanchez, F. 1996. Catchability: a key parameter for fish stock assessment. Reviews in Fish Biology and Fisheries 6: 221-242.

Ashton-Alcox, K., D. Bushek, J. Gius, J. Morson, and D. Munroe. 2016. Report of the 2017 Stock Assessment Workshop (14th SAW) for the New Jersey Delaware Bay Oyster Beds. Haskin Shellfish Research Laboratory, Port Norris, NJ. 158pp.

Erisman, B. E., L. G. Allen, J. T. Claisse, D. J. Pondella, E. F. Miller, and J. H, Murray. 2011. The illusion of plenty: hyperstability masks collapses in two recreational fisheries that target fish spawning aggregations. Canadian Journal of Fisheries and Aquatic Sciences 68 (10): 1705-1716.

Ford, S. 1997. History and present status of molluscan shellfisheries from Barnegat Bay to Delaware Bay: In: The history, present condition, and future of the molluscan fisheries of North and Central America and Europe, Volume 1. Eds. C. L. MacKenzie Jr., V. G. Burrell Jr., A. Rosenfield, and W. L. Hobart. US Department of Commerce, NOAA Technical Report NMFS, Seattle, WA.

Fu, D., A. Dunn, K. P. Michael, and J. Hills. 2016. The development and performance of a length-based stock assessment of Foveaux Strait oysters (Ostrea chilensis, OYU 5) in southern New Zealand, and application to management. Fisheries Research 183: 506517.

Godo, O. R. Factors affecting the reliability of groundfish abundance estimates from bottom trawl surveys. In: Marine fish behavior in capture and abundance estimation. Eds. A. Ferno and S. Olsen. 166-199.

Godo, O. R., S. J. Walsh, and A. Engas. 1999. Investigating density-dependent catchability in bottom-trawl surveys. ICES Journal of Marine Science 56: 292-298.

Hilborn, R. and C. J. Walters. 1992. Quantitative fisheries stock assessment: choice, dynamics, and uncertainty. Chapman and Hall, New York, NY.

Hoffman, J. C., C. F. Bonzek, and R. J. Latour. 2009. Estimation of bottom trawl catch efficiency for two demersal fishes, Atlantic croaker and white perch, in Chesapeake Bay. Marine and Coastal Fisheries: Dynamics, Management, and Ecosystem Science 1: 255269.

Korman, J. and M. D. Yard. Effects of environmental covariates and density on the catchability of fish popululations and interpretation of catch per unit effort trends. Fisheries Research 189: 18-34.

Marenghi, F., K. A. Alcox, R. Wong, B. Reynolds, and G. Ozbay. 2017. Dredge efficiency on natural oyster grounds in Delaware Bay and its application in monitoring the Eastern oyster (Crassostrea virginica) stock in Delaware Bay, USA. Fisheries Research 186: 292-300.

Maunder, M. M., Sibert, J. R., Fonteneau, A., Hampton, J., Kleiber, P., and Shelton, J. H. 2006. Interpreting catch per effort data to assess the status of individual stocks and communities. ICES Journal of Marine Science 63 (8): 1373-1385.

Powell, E. N., K. A. Ashton-Alcox, J. A. Dobarro, M. Cummings, and S. E. Banta. 2002. The inherent efficiency of oyster dredges in survey mode. Journal of Shellfish Research 21: 691-695.

Powell, E. N., K. A. Ashton-Alcox, and J. N. Kraeuter. 2007. Reevaluation of eastern oyster dredge efficiency in survey mode: application in stock assessment. North American Journal of Fisheries Management 27: 492-511.

Rose, G. A. and D. W. Kulka. 1999. Hyperaggregation of fish and fisheries: how catch-per-unit effort increased as the northern cod (Gadus morhua) declined. Canadian Journal of Fisheries and Aquatic Sciences 56: 118-127.

Sagarese, S. R., M. G. Frisk, R. M. Cerrato, K. A. Sosebee, J. A. Musick, and P. J. Rago. 2016. Diel variation in survey catch rates and survey catchability of spiny dogfish and thei pelagic prey in the Northeast U. S. continental shelf large marine ecosystem. Marine and Coastal Fisheries 8: 244-262.

Somerton, D. A., K. L. Weinberg, and S. E. Goodman. 2013. Catchability of snow crab (Chionoecetes opilio) by the eastern Bering Sea bottom trawl survey estimated using a catch comparison experiment. Canadian Journal of Fisheries and Aquatic Sciences 70 (12): 1699-1708.

Wilberg. M. W., J. T. Thorson, B. C. Linton, and J. Berkson. 2010. Incorporating timevarying catchability into population dynamic stock assessment models. Reviews in Fisheries Science 18 (1): 7-24.

## TABLES - CHAPTER 4

Table 4.1. Coefficients for model parameters for average (AM), logistic (LM), and power (PM) models. In parentheses next to each coefficient is the $p$-value associated with that parameter. NS = not selected by backward stepwise AIC (see Methods).

|  | Parameters | 2003 Training Data | 2013 Training Data | Random Training Data |
| :---: | :---: | :---: | :---: | :---: |
| Average Model |  |  |  |  |
| Lower Bay | $q$ | 3.04 | 3.30 | 3.15 |
| Upper Bay | $q$ | 6.48 | 10.55 | 6.97 |
| Logistic Model |  |  |  |  |
| Intercept | $\beta 0$ | -2.021 | -1.981 | 1.425 |
| Total Haul Volume (bushels) | $\beta 1$ | $0.5307(<0.001)$ | 0.6257 (<0.001) | 0.0817 (0.007) |
| Prop. of Subsample Made of Cultch | $\beta 2$ | 0.0441 (<0.001) | 0.0270 (<0.001) | 0.0253 (<0.001) |
| Average Clump Area in Subsample | $\beta 3$ | NS | -0.0001 (0.003) | 0.0001 (0.002) |
| Change in Depth | $\beta 4$ | NS | $0.2965(<0.001)$ | 0.0923 (0.01) |
| Scope | $\beta 5$ | -1.8471 (<0.001) | $0.3109(<0.001)$ | NS |
| Swept Area | $\beta 6$ | NS | -0.0264 (<0.001) | -0.0392 (<0.001) |
| Tow-Tide | $\beta 7$ | -0.0008 (<0.001) | NS | -0.0012 (<0.001) |
| Weighted Industry Coverage | $\beta 8$ | 0.2105 (<0.001) | -0.2764 (<0.001) | NS |
| Power Model |  |  |  |  |
|  | $\alpha$ | 362.43 (0.03) | 142.95 (0.003) | 264.15 (0.05) |
|  | $\theta$ | 2.22 (0.01) | 1.39 (0.009) | 2.16 (0.03) |

Table 4.2. Model performance estimated using the mean squared error (MSE) for each modeling approach (average, logistic, and power) and each training-test data combination. The overall reported mean for each method is the mean of the MSE scores for each training-test combination.

|  | Average | Logistic | Power |
| :---: | :---: | :---: | :---: |
| 2003 Training / 2013 Test | 1,389 | 18,599 | 2,074 |
| 2013 Training / 2003 Test | 18,119 | 131,440 | 4,218 |
| Random Training / Random Test | 2,312 | 12,421 | 1,935 |
| Mean | 7,273 | 54,153 | 2,742 |

## FIGURES - CHAPTER 4



Figure 4.1. Map of sampling locations.


Figure 4.2. Capture efficiency as a function of two variables, total haul volume (left panels) and proportion of the sub-sample that was made up of cultch (right panels) for all three training-test pairs of data $(A=2003$ training data, $B=2013$ training data, $C=$ random training data).


Figure 4.3. Residuals (model-estimated density ed - true density $t d$ ) calculated from three approaches, a logistic model (LM) , a power model (PM), and an average model (AM), to estimating true density in the sampled area. See Methods for model descriptions.


Figure 4.4. The true density $(t d)$ of oysters in the sampled area as a function of the proportion of the sample that was made up oysters. (A) 2003 data were used to train and estimate power model (PM) parameters for the predicted line while the points plotted are the 2013 observations, (B) 2013 data were used to train and estimate power model parameters for the predicted line while the points represent the 2003 data, and (C) a random draw of 24 of the 48 experimental observations (2003 and 2013 experimental data) were used to train and estimate parameters for the predicted line while the points represent the remaining 24 data points.

## CONCLUSIONS

Our ability to evaluate the response of a fished population to changes in the ecosystem or changes in fishing exploitation rate is dependent on how confident we are in stock assessment results. Output from stock assessments, including abundance and biological reference points, are highly sensitive to misspecification of fishery and survey catchability and selectivity (Harley et al. 2001; Chen et al. 2003; Francis 2011; Jonsson et al. 2013; Butterworth et al. 2014; Cadrin et al. 2016). Despite this, the processes that drive patterns in selectivity and catchability remain poorly understood for many fisheries (Sampson 2013; Maunder and Piner 2014; Wang and Maunder 2017).

Traditionally, selectivity and catchability are parameters that are estimated inside the stock assessment model where they conform with the model input data and assumptions. However, since catchability and selectivity can be highly correlated with other model parameters (Arreguin-Sanchez 1996) and can be highly variable in space and time (Wilberg et al. 2010), more recently a call for experimental and field observation-based approaches to estimates of selectivity and catchability are becoming common. For example, Somerton et al. (1999) state, "Cooperative studies by experimentalists and modelers are needed to derive a better understanding of the situations in which experimentally derived estimates of catchability can improve knowledge of stock size or fishing mortality rates", and Cadrin et al. (2016) suggest, "Considering the sensitivity of stock assessments to the assumed form of selectivity and estimates of catchability, as well as their importance for fishery management advice, we
suggest that the selection pattern and catchability (or relative efficiency) should be evaluated from a field experiment whenever possible to reduce uncertainties in stock assessments."

One could view this dissertation, in part, as an answer to these calls. However, I suggest that we not only endeavor to derive estimates of catchability and selectivity through experimentation and observation, but that we should attempt to understand the underlying ecology that drives variability in these parameters so that we can better understand and predict how shifts in life history, distribution, and behavior will influence perceived dynamics of a population. Wang and Maunder (2017), state in reference to assessment model misspecification, "Therefore, we recommend that more work be done to ensure that models are correctly specified. Unfortunately, there is a lack of understanding and uncertainty in the fundamental biological and fishing processes of most, if not all, fish stocks, making the removal of model misspecification difficult." The unique characteristics of a species require similarly unique applications and an understanding of the underlying ecological processes that make them unique. While it is likely not feasible, nor necessary, to develop a unique stock assessment modeling approach for every species, research on the underlying ecology of each individual species could lead to species-level applications of parameters like selectivity and catchability that account for unique life history within more general modeling frameworks.

My work is not the only research attempting to intersect underlying ecological processes with parameters used in stock assessments, of course. For instance, the most recent butterfish (Peprilus triacanthus) stock assessment used a thermal niche model to
develop temperature-dependent estimates of catchability that account for the availability of butterfish to the survey gear in the water temperature where it was deployed (Adams et al. 2014). In this application, the organisms's unique thermal niche is being used to develop realistic estimates of survey catchability when and where the survey occurred. Additional examples exist for temperature- and density-dependent catchability, predator-dependent catchability, and selectivity patterns that account for unique migration patterns (Wilberg et al. 2010; Chakraborty et al. 2012; O’Boyle et al. 2016). Each method is similar in that the knowledge of the underlying ecology of the organisms is used to define direct estimates, or estimate bounds, for catchability and selectivity.

In Chapter 1 I identified a selectivity pattern in the recreational summer flounder fishery that was significantly skewed toward female fish relative to the selectivity patterns in the commercial fishery or the survey. As discussed, this pattern likely results from sex- and age-specific movement and habitat use. In Chapter 2, given the identified selectivity pattern in the recreational fishery and its influence on the catch composition, I evaluated alternative management actions that could reduce fishing pressure on large, fecund female fish. The unique characteristics of summer flounder, that they likely separate out by sex and age in space and time, require a separate selectivity pattern be modeled for this fishery than for the commercial catch or the survey. Furthermore, given this behavior results in a highly biased female catch in the recreational fishery, management action may need to be taken to evaluate alternatives measures that could reduce fishing pressure on female fish.

In Chapter 3 I discovered that we cannot treat oysters as we would surf clams or scallops relative to survey gear catchability. Oysters have a unique catchability characteristic such that changes in density lead to changes in gear performance, and this is likely related directly to their need to grow in a three-dimensional reef structure. One of the unique ecological characteristics of oysters, that they grow in three-dimensional reefs, requires a unique application to how we sample, assess, and manage them. In Chapter 4, I worked on developing these applications and evaluating the performance of each so that a method that accounts for density-dependence in catchability could be incorporated into the stock assessment of oysters in Delaware Bay.

Without an understanding of summer flounder availability relative to fishing effort, one would never reevaluate the sex- and age-specific selectivity patterns in the recreational fishery. Similarly, without knowing how oysters interact with an oyster dredge at high density, one would never seek to develop density-dependent models of catchability for that survey. Hare (2014) calls for "a multi-hypothesis, integrative and multidisciplinary approach to fishery assessment and management" and Able (2016) states, "The development of our understanding of fish and other marine fauna, including my own over several decades, has proceeded from basic natural history to ecology and evolution, but we often need to return to natural history to address deficiencies in our attempts to manage fisheries, conserve habitats, and model ecosystems". I agree with both sentiments and I hope this work contributes evidence in support of the notion that accurate assessment and sustainable management of marine resources depends on our ability to collaborate as fishermen, stock assessment
scientists, ecologists, ocean modelers, resource managers, biologists, and natural
historians.

## Literature Cited - Summary

Able, K. W. 2016. Natural history: an approach whose time has come, passed, and needs to be resurrected. ICES Journal of Marine Science 73 (9): 2150-2155.

Adams, C. F., T. J. Miller, J. P. Manderson, D. E. Richardson, and B. E Smith. 2014. Butterfish 2014 Stock Assessment. Northeast Fisheries Science Center Reference Document 15-06.

Arreguin-Sanchez, F. 1996. Catchability: a key parameter for fish stock assessment. Reviews in Fish Biology and Fisheries 6: 221-242.

Butterworth, D. G., R. A. Rademeyer, A. Brandao, H. F. Geromont, and S. J. Johnston. 2014. Does selectivity matter? A fisheries management perspective. Fisheries Research 158: 194-204.

Cadrin, S. X., G. R. DeCelles, and D. Reid. 2016. Informing fishery assessment and management with field observations of selectivity and efficiency. Fisheries Research 184: 9-17.

Chakraborty, S., S. Pal, and N. Bairagi. 2012. Predator-prey interaction with harvesting: mathematical study with biological ramifications. Applied Mathematical Modeling 36: 4044-4059.

Chen, Y. L. Chen, and K. I. Stergiou. 2003. Impacts of data quantity on fisheries stock assessment. Aquatic Sciences 65: 1-7.

Francis, R. I. C. C. 2011. Data weighting in statistical fisheries stock assessment models. Canadian Journal of Fisheries and Aquatic Sciences 68: 124-1138.

Hare, J. A. 2014. The future of fisheries oceanography lies in the pursuit of multiple hypotheses. ICES Jounral of Marine Science 71 (8): 2343-2356.

Harley, S. J., R. A. Myers, and A. Dunn. 2001. Is catch-per-unit-effort proportional to abundance? Canadian Journal of Fisheries and Aquatic Sciences 58: 1760-1772.

Jonsson, T. M. Setzer, J. G. Pope, and A. Sandstrom. 2013. Addressing catch mechanisms in gillnets improves modeling of selectivity and estimates of mortality rates: a case study using survey data on an endangered stock of Arctic char. Canadian Journal of Fisheries and Aquatic Sciences 70: 1477-1487.

Maunder, M. and K. R. Piner. 2014. Contemporary fisheries stock assessment: many issues still remain. ICES Journal of Marine Science 72 (1): 7-18.

O'Boyle, R, M. Dean, and C. Legault. 2016. The influence of seasonal migration on fishery selectivity. ICES Journal of Marine Science 73 (7): 1774-1787.

Sampson, D. B. and R. D. Scott. 2011. A spatial model for fishery age-selection at the population level. Canadian Journal of Fisheries and Aquatic Sciences 68(6): 1077-1086.

Sampson, D. B. 2013. Fishery selection and its relevance to stock assessment and fishery management. Fisheries Research 158: 5-14.

Wang, S.-P. and M. N. Maunder. 2017. Is down-weighting composition data adequate for dealing with model misspecification, or do we need to fix the model? Fisheries Research 192: 41-51.

Wilberg. M. W., J. T. Thorson, B. C. Linton, and J. Berkson. 2010. Incorporating timevarying catchability into population dynamic stock assessment models. Reviews in Fisheries Science 18 (1): 7-24.


[^0]:    * Published as: Morson, J. M., E. A. Bochenek, E. N. Powell, E. C. Hasbrouck, J. E. Gius, C. F. Cotton, K. Gerbino, and T. Froehlich. 2015. Estimating the sex composition of the summer flounder catch using fishery independent data. Marine and Coastal Fisheries: Dynamics, Management, and Ecosystem Science 7: 393-408.

[^1]:    * Published as: Morson, J. M., D. Munroe, R. Harner, and R. Marshall. 2017. Evaluating the potential for a sex-balanced harvest approach in the recreational summer flounder Paralichthys dentatus fishery. North American Journal of Fisheries Management: dx.doi.org/10.1080/02755947.2017.1362490

